

---

# Chapter 1

---

Introduction



## **1. Background: Diabetes mellitus**

Diabetes mellitus (DM) is a category of metabolic disorders characterized by hyperglycemia due to deficiencies in insulin secretion, insulin action or both.<sup>1</sup> As a result of ageing, urbanization and associated lifestyle changes, the global prevalence of DM is increasing rapidly.<sup>2</sup> DM is recognized as the eighth leading cause of death worldwide and is spreading very rapidly, particularly in low- and middle-income countries, with an overall prevalence rate of 8.5 percent in adults.<sup>3</sup> According to the International Diabetes Federation, the number of people diagnosed with diabetes worldwide increased explosively from 151 million in 2000 to 463 million in 2019 and this number is projected to reach 578 million by 2030, and 700 million by 2045.<sup>4</sup> In 2019, more than one million children and adolescents were reported to have type 1 diabetes mellitus (T1DM). Total healthcare spending on diabetes in 2019 was USD 760 billion and is expected to increase to USD 845 by 2040.<sup>5-7</sup> Such soaring figures need aggressive research not only to develop new molecules for efficient diabetes treatment, but also to develop alternative therapeutic approaches through the use of nanotechnology to overcome the problems associated with traditional approaches to the drug delivery.

In the development of DM multiple pathogenic processes are involved. This range from autoimmune disruption of the pancreatic  $\beta$ -cells with consequent deficit of insulin to abnormalities that result of insulin resistance. Deficient insulin action on target tissues is the cause of these metabolic anomalies.<sup>8</sup> Deficient insulin action results from inadequate insulin secretion and/or diminished tissue responses towards insulin at one or more steps in the complex pathways of hormone action. Impairment of insulin secretion and defects in insulin action frequently coexist in the same patient, and it is often unclear which abnormality, if either alone

or both, is the primary cause of the hyperglycemia.<sup>9</sup> There are four clinical classes of diabetes including type 1, type 2, other specific types of diabetes (genetic defects in  $\beta$ -cell function or insulin action, disease of exocrine pancreas, drug- or chemically induced diabetes) and gestational diabetes mellitus (GDM).<sup>10,11</sup> Majority of diabetes cases fell into two broad groups of etiopathogens. In one type, T1DM, the cause is an absolute insulin secretion deficiency. Individuals at increased risk of developing this type of diabetes can often be identified by serological evidence of an autoimmune pathological process occurring in the pancreatic islets and by genetic markers.<sup>12</sup> In the other, far more predominant category, type 2 diabetes mellitus (T2DM), the cause is a combination of insulin resistance and an insufficient compensatory insulin secretory response.<sup>1,13</sup> For the purpose of this thesis, discussion will be mainly focused on T1DM.

## **2. T1DM: causes, current status and treatment**

T1DM is triggered by an autoimmune reaction in which the body's immune system damages the insulin-producing  $\beta$  cells of the pancreas.<sup>14</sup> This results in either little or no insulin being released by the body. The origins of this inflammatory mechanism are not well known but a probable scenario is that the combination of genetic vulnerability (conferred by a wide number of genes) and an environmental stimulus, such as a viral infection, initiates the autoimmune reaction.<sup>15</sup> Once stimulated, macrophages secrete multiple inflammatory cytokines including interleukin- $1\beta$  (IL- $1\beta$ ), interleukin-12 (IL-12) and tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) arising from stimulated T cells.<sup>12,16</sup> Such cytokines destroy the  $\beta$ -cells by causing oxygen-free radicals, nitric oxide and lipid peroxides within the  $\beta$ -cells and enhance Th1 cell-mediated inflammatory

responses.<sup>17</sup> Toxins, or certain dietary factors, were also associated with development of T1DM. The condition can occur at any age although T1DM is more severe in children and adolescents.

Strong glycemic regulation in T1DM decreases the risk of development and progression of late diabetic microvascular complications and also has long-term beneficial effects on the development and progression of nephropathy, hypertension, cardiovascular diseases and atherosclerosis.<sup>18</sup> Existing therapies have limited effectiveness, limited tolerability and substantial mechanism-based side effects such as weight gain and episodes of hypoglycemia. Moreover, only a handful of the treatments available which can sufficiently tackle underlying risks, such as obesity and/or insulin resistance. Thus, newer approaches are the need of the hour with greater emphasis on exploring delivery approaches that rely on physiological responses (e.g., glucose-mediated insulin secretagogues) without substantial weight gain.<sup>19,20</sup>

Despite the development of active anti-hyperglycemic agents, the major challenges in successful diabetes treatment include improving the existing therapies to maintain an acceptable and controlled glucose level and resolving the long-term complications associated with diabetes.

### ***2.1. T1DM and insulin: treatment and limitations***

The gold standard in T1DM therapy is functional insulin therapy with a system in basal-bolus insulin. Different insulin regimens used for treating T1DM patients include short-acting, long-acting, and premixed human insulin and insulin analog preparations. Short-acting analogs of insulin include Insulin lispro (Humalog ®), Aspart (Novorapid ®) and Glulisine (Apidra ®) that work for up to 3-5 hours. Long-acting analogs of insulin include Insulin Glargine (Lantus ®) and Detemir (Levemir ®) with an effect lasting less than 24 hours.<sup>21</sup> However, intense insulin therapy also raises the risk of hypoglycemia. Despite the emergence of modern insulin analogs

with physiologically appropriate absorption profiles and less routine bioavailability differences relative to older insulin preparations; varying blood glucose levels, which often raise the risk of hypoglycemia, remain a serious threat. There are shortcomings in T1DM treatment with prolonged insulin therapy and many patients do not obtain the required blood glucose levels and metabolic targets.<sup>22,23</sup> Firstly, insulin tackles only partly the paradoxical and pathophysiological glucagon abundance. Secondly, in individuals with T1DM, the gastric emptying rate is changed and even the fastest acting mealtime insulin peaks are too late to balance postprandial glucose absorption, leading to significant postprandial glucose excursions.<sup>24,25</sup> Thirdly, intense insulin therapy is often associated with weight gain, potentially increasing cardiovascular risks, leading to hyperglycemia.<sup>26</sup> Obesity is a massive problem in T1DM, with an approximate prevalence of about 50 percent in some developing countries.<sup>27</sup> Thus, novel non-insulin adjunct therapies need to be explored in patients with T1DM.

## ***2.2. Conventional oral anti-diabetic formulations: treatment and limitations***

The best treatment for T1DM is currently combination therapy using medications with actions complementary to insulin that can boost glycated hemoglobin (HbA1c), reduce the risk of hypoglycemia, cardiovascular disorders and weight loss. Glucagon-like peptide-1 (GLP-1) receptor agonists and pramlintide inhibit glucagon secretion, and slow gastric emptying.<sup>28,29</sup> Dipeptidyl peptidase-4 (DPP-4) inhibitors raise endogenous GLP-1 concentration by 2-3 times, potentiating glucose-dependent insulin release and glucagon inhibition.<sup>30</sup> Insulin resistance is evident in even lean T1DM patients. Thiazolidinedione primarily enhances insulin sensitivity and this effect is also associated with weight gain, while sulfonylureas only potentiate insulin secretion and therefore no significant impact on glucose levels or insulin dose will be expected in

patients without  $\beta$ -cell function with these medications. Metformin also decreases the production of hepatic glucose and causes a minor weight loss, but its treatment for T1DM patients is not recommended in any international guidelines as it raises the risk of adverse gastrointestinal events.<sup>31</sup> Sodium-glucose co-transporter-2 (SGLT2) inhibitors reduce reabsorption of renal glucose, resulting in substantial glucose excretion in urine. Since the mode of action of SGLT2 inhibitors is insulin independent, if used in patients with T1DM a significant impact of these drugs on glucose regulation can be anticipated.<sup>32</sup>

These oral anti-diabetic medications have some drawbacks and are usually used in combination with insulin in the treatment of T1DM patients. Solubility and permeability challenges are quite popular among these anti-diabetic drugs which are already present in the market. Some oral anti-diabetics like sulfonylureas, have poor solubility which contributes to less bioavailability and hence a frequent dosage regime that results in non-compliance by the patient due to the missed dose.<sup>33</sup> At the other hand, metformin, frequently used as a first-line medication for T2DM, belongs to the Biopharmaceutical Classification System (BCS) class III and is thus highly soluble but poorly permeable leading to its slow and insufficient absorption.<sup>34,35</sup> Shortcomings seen with several other anti-diabetic medications are short half-life (3-5 h) with thiazolidinedione like pioglitazone resulting in poor bioavailability and decreased therapeutic efficacy.<sup>36,37</sup> Repaglinide, often used as an alternative drug as an adjunct therapy, also has a limited half-life (~1 h) and has to be given three times a day, frequently contributing to patient non-compliance.<sup>38</sup> The marketed formulations of GLP-1 agonists that is Byetta<sup>®</sup> for exenatide and Victoza<sup>®</sup> for liraglutide, have a limited half-life and are given solely by s.c. route which often causes discomfort to the patient like injection site (ISR) reactions such as swelling,

pruritus, local pain and chances of infection. In fact, these medications induce nausea and vomiting too.<sup>39-41</sup> In addition to the aforementioned issues, some of the well-known and commonly used anti-diabetic drugs often cause serious hypoglycemia and weight gain. Moreover, such a therapeutic approach does not allow for completely controlled homeostasis of glucose and may result in disorders of the cardiovascular system over time. Hence it is of utmost importance for diabetic patients to find alternative and better therapies using novel drug delivery systems (NDDS).

### **3. Nanomedicines for diabetes: significance and status**

NPs are colloidal drug delivery systems that involve nanocrystals, polymeric nanoparticles, polymeric micelles, solid lipid NPs, nanosuspensions, and multilayer nanoparticles in the size range of 10-1000 nm in diameter. They can be in the form of a matrix system in which the drug is dispersed all across the particles, or as a reservoir system in which the drug is enclosed to a cavity covered by a polymeric membrane.<sup>42-44</sup>

Nano formulations not only enhance the drug's solubility but can have many other advantages such as decreased dosage, rapid onset of action, controlled release of drugs, minimized side effects, targeted drug distribution, enhanced half-life of the drug, decreased patient to patient variability as well as improved bioavailability, and thereby can solve several of the drawbacks of existing anti-diabetics.<sup>45-48</sup> To mention a few examples; many studies indicated an increased area under curve (AUC) and higher bioavailability of sulfonylureas when delivered by nanoformulations due to its enhanced solubility.<sup>49-51</sup> Several sustained release nanoformulations were also reported to address the permeability problems of metformin.<sup>52-54</sup> Hasan et al. observed a two-fold improvement in AUC of metformin niosomes relative to pure

drug solution.<sup>55</sup> Nanotherapy has found a way to overcome the short half-life obstacle; pioglitazone nanostructured lipid carriers embedded in a transdermal patch (TDP) substantially reduced BGL to 24 hr relative to the commercial formulation (6 hrs).<sup>56</sup> Nanoformulations have also been effective in addressing limitations linked to high dose frequency by allowing the medication to be released sustainably. A sustained release of the repaglinide from its nanoformulations revealed a robust hypoglycemic effect relative to the already marketed preparations.<sup>38,57</sup> Furthermore, when delivered orally, nanoformulations of GLP-1 analogs demonstrated an improved hypoglycemic effect with comparable AUC compared to s.c. administration of drug solution.<sup>48,58</sup>

Most interestingly, nano formulations often act at the molecular level to facilitate cellular drug uptake or block efflux pathways such as P-glycoprotein (P-gp) pump or via targeting specific receptors, thus further improving the pharmacokinetic and pharmacodynamic profile of anti-diabetic drugs. TPGS-based repaglinide nanocrystals inhibited the P-gp efflux pump by rigidizing the membrane lipid bilayers as well as inhibited the CYP3A4 enzyme which is responsible for metabolizing the drug in the liver, contributing to a substantial improvement in repaglinide bioavailability.<sup>59,60</sup> Glibenclamide's bioavailability has also been enhanced by formulating SLNs containing Compritol 888 ® ATO which form chylomicrons and thus enhance the drug's lymphatic transport. Improved surface area and decreased efflux transport by these SLNs are also responsible for the drug's improved bioavailability.<sup>61</sup> In another study on preventing drug efflux from the cells, glibenclamide SNEDDS showed better uptake and therefore higher bioavailability of the drug by inhibiting efflux transporters of ATP-binding cassette (ABC) that are known to transport glibenclamide out of the cells. This was because of



the presence of Tween 80, Cremophor RH, TPGS and Brij 30 in glibenclamide SNEDDS.<sup>50</sup> Up on oral administration of Fc receptor targeting exenatide NPs showed increased hypoglycemic activity compared to pure drug injection by s.c route and greater gastrointestinal retention compared to unmodified NPs. Fc receptors exist in the small intestine as well as colon and have an expanded surface region for NP absorption.<sup>62</sup> In addition, exenatide loaded albumin and dextran NPs demonstrated higher oral relative bioavailability (77%) due to higher lymphatic absorption of dextran effectively binding to the dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin (DC-SIGN) receptor family.<sup>48</sup> **Table 1.1** discusses different types of nanoformulations for treatment of DM.

#### **4. Lisofylline (LSF) as a potential molecule for treatment of autoimmune disorders**

LSF was originally developed and tested to reduce cellular damage due to autoimmunity, hypoxia and ischemic reperfusion.<sup>63</sup> LSF has been used to overcome morbidity and mortality during serious infections associated with cancer chemotherapy and for treatment of acute lung injury after severe trauma.<sup>64,65</sup> It has also been reported for its therapeutic efficacy in early treatment of diabetes wherein, it enhances glucose-stimulated insulin secretion,<sup>66,67</sup> causes reversal of insulin insensitivity and glucose-induced phosphorylation of the insulin receptor.<sup>68</sup> The protective role of LSF in diabetes is mainly attributed to the promotion of mitochondrial metabolism in  $\beta$ -cells, normalizing the membrane potential of mitochondria and thus stimulating energy production.<sup>69,70</sup> Mitochondrion controls cell apoptosis<sup>71</sup> and regulates  $\beta$  cell insulin secretion.<sup>14</sup> This broad spectrum of activity suggests that LSF bears significant clinical utility in preventing both T1DM and T2DM.<sup>70,72</sup> Considering the immense therapeutic potential and

multiple pharmacological activities of LSF as stated above, appropriate and improved therapy of LSF is of paramount significance.

### **5. LSF: Currently under development for treatment of diabetes**

LSF has well reported benefit in T1DM. Striffler and Nadler have demonstrated that LSF decreased IL-1 $\beta$  induced islet dysfunction in isolated pancreatic islets along with maintaining insulin secretion. Furthermore, co-incubation of LSF with insulin secreting, rat insulinoma cells, INS-1 in the presence of pro-inflammatory cytokines, IL-1 $\beta$ , TNF- $\alpha$  and IFN- $\gamma$ , restored glucose-stimulated insulin secretion and mitochondrial metabolism to control levels.<sup>72</sup> LSF treatment in the non-obese diabetic (NOD) mouse model of T1DM delayed the onset of diabetes.<sup>73</sup> In T2DM prediabetic mice, LSF administered at 25 mg/kg, intraperitoneally (IP), twice daily potentiated glucose-mediated insulin secretion possibly by stimulating the functioning of residual  $\beta$  cells.<sup>74</sup>

LSF administration in obese Zucker rats that are well known models for obesity and insulin resistance, reduced p-STAT4 in visceral adipose tissues and showed improvement in metabolic profile of Zucker rats by reducing fasting plasma glucose and improving insulin sensitivity. LSF treatment increased feed efficiency with concomitant increased lipid storage in adipose tissue, which may be beneficial in preventing deposition of ectopic (nonadipose) lipids.<sup>75</sup> Yang et al. investigated LSF and exendin-4 (Ex-4) combination (LSF (27 mg/kg/day) and Ex-4 (18 nM/day) were delivered by the Alzet osmotic minipumps to provide a 28 day consistently systemic administration by s.c. route) simultaneously block the autoimmune cytokine damage along with simultaneously supplying a growth-promoting stimulus for  $\beta$ -cells in the non-obese diabetic (NOD) mouse model. The results demonstrated that LSF and exendin-4 combined therapy could effectively and efficiently reverse insulin-dependent autoimmune diabetes in the

NOD mouse model. Although, actual mechanism behind synergistic effect of LSF and Ex-4 was not known. Here, Ex-4 works as an activator of the GLP-1 receptor by activating cyclic AMP and protein kinase A while LSF improves mitochondrial function and blocks the expression of STAT-4 activation in target tissues.<sup>76</sup>

Several researchers have demonstrated the efficacy of LSF in diseases that are either mediated by altered lipid profile or induced by pro-inflammatory cytokines including diabetes. Nonetheless, the low potency, poor oral bioavailability, and short half-life of LSF hinder its clinical translation. Research efforts aiming at improving these shortcomings of LSF are still in infancy.<sup>77</sup>

In spite of being a potent molecule, LSF is quite less explored in research and very few reports are available focusing on the physicochemical and pharmacokinetic issues associated with LSF. In one such study reported in the year 2006, Cui et al. have synthesized 32 analogs based on the structural motif of LSF wherein, only two of these analogs were found to be effective in protecting  $\beta$ -cells from cytokine-induced injury and maintaining insulin secretory ability in cell culture based evaluation.<sup>69</sup> Nonetheless, no *in vivo* pharmacokinetic and pharmacodynamics data is reported on these analogs till date to determine if the synthesized analogs improved the metabolic stability and oral bioavailability of LSF. We did not come across any other study highlighting the approaches to modify LSF or deliver it by either conventional or novel drug delivery systems.

## **6. Drug delivery challenges associated with LSF**

The broad spectrum of activity of LSF suggests its significant clinical potential but in spite of being a potent molecule it poses certain major challenges that limit its clinical development.

### **6.1. High aqueous solubility**

LSF has a high aqueous solubility (~60 mg/mL in water) which hinders its encapsulation into any delivery system.<sup>72</sup> Due to its hydrophilicity, it exhibits low intracellular absorption, short half-life (~ 0.75-1.17 h), rapid clearance, sub-optimal distribution and poor pharmacokinetics.

### **6.2. Extensive first pass metabolism and short half-life**

LSF undergoes rapid interconversion into PTX necessitating a high dose and frequent dosing for its therapeutic action.<sup>78</sup> LSF has a low (to non-existent) oral bioavailability in humans (~5.9 %) due to its extensive first pass metabolism.<sup>68</sup> Due to these pharmacokinetic issues, it is required in high doses inspite of being potent.

Nadler *et al.* have reported the anti-diabetic potential of LSF in streptozotocin (STZ) induced diabetic model at a dose of 25 mg/kg, i.p., twice daily.<sup>72</sup> Similarly, Yang Z. *et al.* reported the effectiveness of LSF in diabetes prevention in multiple low dose STZ induced diabetic mice model at a dose of 25 mg/kg, i.p. twice daily, for 14 consecutive days.<sup>74</sup> Combination delivery of LSF and  $\beta$ -cell growth factor, exendin-4 has been explored for reversal of autoimmune diabetes in NOD mice wherein, LSF was administered at 27 mg/kg/day by s.c. route using osmotic mini pump for 28 days.<sup>76</sup> In clinical trials of LSF in T1DM, LSF has been administered at a single dose of 9 mg/kg by continuous IV infusion or at 12 mg/kg by continuous subcutaneous infusion over a 10 hour period during the alternate period 1 week apart.<sup>79</sup> Apart from T1DM, in other ongoing clinical trials of LSF for treatment of allogeneic bone marrow transplants<sup>80</sup>, acute lung injury and acute respiratory distress syndrome, the drug is administered

**Table 1.1**

Nanoformulations in diabetes: significance and status

<b>Drug</b>	<b>Drug Delivery System</b>	<b>Dose and ROA</b>	<b>Remarks</b>	<b>Ref</b>
<b>Nanoparticles (NPs)</b>				
	Bioadhesive NPs	-----	The formulations of exenatide- or insulin-loaded NPs demonstrated substantial reduction of glycemia in patients with diabetes beginning from 2 h post oral administration.	81
	Chitosan PGA NPs (250 nm and 25 mV)	30 IU/Kg, oral	NPs were prepared by incorporating a solution composed of insulin and -y-PGA to trimethyl chitosan or chitosan using a tripolyphosphate, magnesium sulphate, etc. via process of mild ionic gelation.	82
	PLGA NPs (247 nm and -16.7 mV)	30 & 50 mg/kg orally	Protect insulin from gastric pH. ~25% insulin release was observed after 24 h of incubation at pH 1 but comparatively higher release in PBS 7.4 (~35% in 24 h). NPs showed substantial reduction in glycemia that was sustained for 24 h.	83
<b>Insulin</b>	PLGA and HPMCP55 NPs (180 nm)	50 IU/kg, Oral	The formulation of NPs demonstrated a decrease of up to > 80% in glycemia, and also a determined relative bioavailability of 11.3% in diabetic rats.	84
	PLGA NPs consisting of chitosan and stabilized by pluronic 188 (133 nm and 40 mV)	Oral	NPs demonstrated moderately biological adhesion to the rat intestine and comparatively higher reduction in serum glucose levels compared to non-chitosan PLGA NPs. The relative bioavailability was found to be 10.5% and 7.6%, respectively.	85
	Polycaprolactone and Eudragit RS blend NPs (700 nm and +40 mV)	50 IU/kg, oral	NPs demonstrated a reduction in glycemia of up to 53 per cent at 50 IU / kg, which decreased with increased dosage	86
	Chitosan & $\gamma$ -poly(glutamic acid) NPs (245 nm and 27 mV)	-----	NPs exposed the close junctions of the Caco-2 cell monolayer in vitro transiently and released insulin. Radiolabeled insulin has been shown to be ingested and detected as early as 30 min after administration in the kidney and urinary bladder.	87
<b>Metformin</b>	Chitosan and gum arabic NPs (146.5 $\pm$ 8.7 nm)	40 mg/kg orally	Significant reduction in fasting blood glucose as compared to pure metformin (150 mg/kg) administered orally, for 21 days	52

	Alginates NPs (60 – 150 nm)	1.5 g/kg	Significant reduction in blood glucose level as compared to pure metformin (150 mg/kg, i.p.)	54	
	Chitosan based non-composite films containing mesoporous MCM-41 (5.8 nm) and MCM-41 (5.93 nm)-aminopropylsilane NPs	----	Prolonged release till 15 days, no cellular toxicity	53	
	Polaxamer 188 and Eudragit L 100 NPs (138.8 nm)	----	Enhanced drug solubility by 10 folds than pure drug and 97.5% drug released within 60 min	88	
<b>Pioglitazone</b>					
			<i>PK parameters</i>		
		5 mg/kg, orally	AUC <sub>(0-∞)</sub> (ng h/mL)	t <sub>1/2</sub> (h)	C <sub>max</sub> (ng/mL)
<b>Glibenclamide</b>	HPMC K15M and lactose NPs (168.6 nm)		71680.24±63.6	5.73±0.42	9428.42±897.8
			172383.64±237.2	9.57±0.23	24451.14±2170.5
			AUC <sub>(0-∞)</sub> , fasted state µg h/L	AUC <sub>(0-∞)</sub> , fed state µg h/L	
<b>Repaglinide</b>	Soluplus® (SLPS) and Kolliphor™ E-TPGS nanocrystals, (304±6 nm for 1% w/v SLPS (TD-A) and 331±34 nm for SLPS and 0.5% w/v of TPGS (each) (TD-B))	2 mg/kg, orally	25.76±4.17	18.57±6.13	59
			257.75±7.32	241.63±13.51	
			384.07±9.54	355.88±10.69	
			<i>PK parameters</i>	MRT (h)	T <sub>max</sub> (h)
		50 µg/kg, s.c.	AUC (ng h/mL)		C <sub>max</sub> (ng/mL)
<b>Exenatide</b>	Mesoporous silica NPs (SBA-15) (920±120 nm)		1.71±0.07	1.14±0.07	1.16±0.09
	Fc modified polyethylene glycol-poly (lactic-co-glycolic NPs (130±5.2 nm)		8.77±0.76	21.3±0.99	0.39±0.01
		100 µg/kg, orally			1.38±0.22
			Hypoglycemic effect for 12 h when compared to 8 h by s.c. administration of free drug solution (10 µg/kg). Higher gastric residence time (24 h), when compared with unmodified NPs (10h)		
<b>Self emulsifying drug delivery systems</b>					

<b>Pioglitazone</b>	Capryol 90, Transcutol HP, Cremophor ELP SMEDDS (< 50 nm)	----	60% drug release in 24 h, when compared with marketed preparation (17%), and pure drug	91
<b>Glipizide</b>	Phosphatidylcholine (Phosal 53 MCT), Tween 80, and Transcutol P SEDDS (55.94 nm) and S-SEDDS (78.03 nm)	800 µg/kg, i.g.	After OGTT, improved reduction in BGL within 30 min, when compared with pure drug suspension (800 µg/kg), i.g., higher bioavailability	92
<b>Glimepiride</b>	Miglyol® 821, Tween 80, PEG 400, aerosol 200 S-SNEDDS (152 nm)	1 mg/kg, orally	Increased AUC <sub>(0-24h)</sub> for L and S-SNEDDS 248.88±52.22 and 234.64±32.22 respectively as compared to pure drug 128.77±54.25, and marketed formulation 207.20±34.16, orally (1 mg/kg)	51
<b>Glibenclamide</b>	Aerosil 200 (carrier for L-SNEDDS) SNEP (143.6 ± 3.46 nm)	3 mg/kg, orally	In rabbits: significantly higher AUC <sub>0-24</sub> of SNEPs 220.43±44.22 mg h/mL, when compared with L-SNEDDS (3 mg/kg), 139.34±34.14 mg h/mL and pure drug (3 mg/kg) 103.52± 22.60 mg h/mL, orally	93
<b>Micro- and Nano-emulsion</b>				
			<i>PK parameters</i>	
			AUC <sub>0-48h)</sub> (µg min/mL)	T <sub>max</sub> (min)
			t <sub>1/2β</sub> (min)	C <sub>max</sub> (µg/mL)
<b>Glimepiride</b>	Capryol 90, Cremophor RH 40), Transcutol micro-emulsion (38.9 ± 17.46 nm).	5 mg/kg, i.g.	Drug suspension 857.43 Glimepiride complex 3384.2 Microemulsion 6242.76	1690.5 1107.8 4687.8
<b>Repaglinide</b>	Sefsol-218, Tween 80 and Transcutol nanoemulsion (76.23 nm)	1 mg/kg, orally	1 mg/kg dose was able to reduce the blood glucose level by a maximum of 67%	37.6 36.3 74.6
<b>Solid lipid nanoparticles (SLNs) and Nanostructure lipid carriers (NLCs)</b>				
<b>Insulin</b>	Stearic acid and soya phospholipid NPs coated with octaarginine (162 nm and 30 mV)		NPs improved the permeability of insulin in Caco-2 cells by 18.4 folds. Decreased fasting blood glucose by 70% compared to initial fasting blood glucose at 1.5 h and determined relative bioavailability was 13.9%	95

NPs with pluronic F127 & glycerylpalmitostearate (305 nm and -17 mV)	NPs in fasted diabetic rats demonstrated a decrease of ~35 percent in glycyemia at 5 to 8 h post administration resulting in a calculated 6 percent bioavailability.	96
<b>Glibenclamide</b>		
SLNs of Tween 80, Pluronic F68, Phospholipon 90 G (201.5±1.33 nm)	<p><i>PK parameters</i></p> <p>AUC<sub>(0-∞)</sub> (mg h/mL)</p> <p>t<sub>1/2</sub> (h)</p> <p>MRT (h)</p> <p>C<sub>max</sub> (µg/mL)</p> <p>T<sub>max</sub> (h)</p>	61
5 mg/kg, orally	<p>Pure drug suspension</p> <p>94.63±6.65</p> <p>9.07±0.79</p> <p>11.28±0.48</p> <p>7.57±0.64</p> <p>4</p>	61
SLN	<p>333.23±88.9</p> <p>12.69±2.94</p> <p>13.36±1.64</p> <p>18.87±1.11</p> <p>2</p>	2
SLNs of Precirol® and lecithin (100.2±4.2 nm)	Rapid onset of action in just 1h and sustained effect till 8 h	97
Lipohydrogel NPs (LHNs) prepared by coating chitosan on solid lipid nanoparticles (SLNs) (287±14 nm)	Burst release of drug was less in LHN as compared with SLN and free drug solution. Reduced toxicity when compared with SLN	98
NLC of Precirol ATO 5, Miglyol, Tween 80 and Polaxomer 188 (161±4 nm)	Synthesized NLC were able to stimulate enteroendocrinal L cells to release endogenous GLP-1. Ex vivo studies on human jejunal tissue did not report any toxicity; no Lactate dehydrogenase release with no effect on concentration of ATP	99
<b>Vesicular systems</b>		
<b>Insulin</b>	<p>Hepatic-directed vesicle (HDV-1 liposome)</p> <p>In patients with T2DM under stable metformin therapy, the insulin-loaded liposome HDV-1 was well tolerated and resulted in a significant reduction in postprandial glucose excursions</p> <p>Formulations</p> <p>AUC<sub>(0-48h)</sub> (ug h/mL)</p> <p>C<sub>max</sub> (ug/mL)</p> <p>T<sub>max</sub> (h)</p>	100
0.05 to 0.4 U/kg	<p>Free gimepiride in saline, 2 mg/kg, orally</p> <p>1.436±1.65</p> <p>0.394±0.142</p> <p>2</p>	101
Span 60, cholesterol niosomes (371.8±23.20 nm)	Marketed tablet (Amaryl) in saline (1 mg/mL), orally	2
10.42±2.32	1.412±0.0212	2



Niosomes		9.633±1.19	0.316±0.032	6	
Egg phosphatidylcholine, 1,2-dimyristoylphosphatidylcholine, 1,2-dipalmitoylphosphatidylcholine, 1,2-ditetraolylphosphatidylcholine liposomes (64 ± 6 nm)					
<b>Co-delivery of metformin and glipizide</b>					
Increased release of glipizide from 3% to 12% and of metformin from 35% to 64%, in 1 h					
<b>Nanoformulations in Transdermal patches</b>					
<b>Insulin</b>	Chitosan NPs by polyelectrolyte complex formation (110 nm and 21.63 mV)	Transdermal, 2 IU	AUC <sub>(0-60)</sub> (μIU h/mL)	C <sub>max</sub> (μIU/mL)	T <sub>max</sub> (h)
			2614.08	46.68	2
			3153.36	45.80	8
<b>Pioglitazone</b>	NLC of Carbopol, Tween 80, labrasol, triethanolamine and apifil (166.05 nm)	Transdermal patch	AUC <sub>(0-∞)</sub> (ng h/mL)	C <sub>max</sub> (ng/mL)	T <sub>max</sub> (h)
			578.21± 18.45	65.67±61.41	2.14±0.21
			1461.54± 76.34	54.19±14.67	8.57±1.98
		<i>PK parameters</i>		<i>T<sub>max</sub> (h)</i>	
<b>Glimepiride</b>	Cholesterol, ethanol, Propylene glycol, Hydroxypropyl methyl cellulose ethosomes (61 nm)	Transdermal, (1.76 cm <sup>2</sup> ), 1 mg	AUC <sub>(0-∞)</sub> (ng min/mL)	C <sub>max</sub> (ng/mL)	T <sub>max</sub> (h)
			215.71	21.53	4
			397.55	135.16	2.5
			1187.48	46.09	2.5
		3-fold increase in total drug release			
<b>Glibenclamide</b>	Chitosan and poloxamer 188 nanocrystals in transdermal patch (TDP)	3mg, Transdermal	Drug permeation (μg/cm <sup>2</sup> )	Cumulative drug release (μg/cm <sup>2</sup> )	
			107±4.3 (at 0.5 h) and	362±25.25	105

(429 nm)	loaded microcrystals	119±8.6 (at 1 h)		
	Glibenclamide loaded nanocrystal	148±5.43 (at 0.5 h) and 177±10.6 (at 1 h)	498±33.35	
	In diabetic rats: Hypoglycemic effect at 24h was significantly higher when compared with oral glibenclamide or glibenclamide loaded microcrystals TDP			
		AUC <sub>(0-∞)</sub> (ng h/mL)	t <sub>1/2</sub> (h)	C <sub>max</sub> (ng/mL)
	Oral suspension of marketed preparation (Daonil®)	3042.11 ±216.3	8.88 ±0.0	448.65±32.65
	Glibenclamide conventional gel, 1mg	1166.54±120.42	13.6	98.96±15.16
	NLC	4141.61±265.32	17.3	265.63±18.96
			7	4±1.62
			21	4±0.91
			1	
			49	
NLC of Capryol™ 90 (120.69±2.13 nm)	1 mg, Transdermal			

### Miscellaneous nanoformulations

Layersomes using poly(allylamine hydrochloride), poly(acrylic acid) and folic acid (266.2±10 nm and 25.4 mV)	50 IU/kg, Oral		The developed FA-Ins-layersomes exhibited: Excellent stability in simulated biological fluids and hypoglycemic response in diabetic rats. Prolonged hypoglycemia up to 18 h, indicative of easy-to-administer, patient-friendly oral formulation that can combat diabetes with improved therapeutic profile.	106
Glucose responsive poly(ethylene glycol)-block-poly[(2-phenylboronic esters 1,3-dioxane-5-ethyl) methylacrylate] (MPEG-block-PPBDEMA) micelles (50 nm and -40.2 mV)	--		The block polymers containing phenylborate ester that responded to changes in the glucose concentration at neutral pH and act as self-regulated insulin delivery	107
Metformin conjugated	150		Prolong drug release, nanotubes-maintained hypoglycemia for a much longer	108

### Insulin

### Metformin

	carbon nanotubes	mg/kg, orally	time	
<b>Repaglinide</b>	Repaglinide-phospholipid complex enriched micelles using poloxamer 188 (525.79±23.62 nm)	2 mg/kg, for 7 days, orally	83.02% reduction in blood glucose level, by the optimized formulation, while only 55.40% reduction by marketed tablet (2mg/kg), orally for 7 days	38
<b>Exenatide</b>	Poly (ethylene glycol)- <i>b</i> -brush poly(L-lysine) polymer (24 nm)	5 µg/kg, s.c.	Blood glucose level maintained for 7 days, as compared to free drug solution (4.5h)	109, 110

at a dose of 3 mg/kg with a maximum of 300 mg intravenously (infusion) for 10 min for every 6 h till 31 days.<sup>111</sup> These studies and reports prove the immense therapeutic potential of LSF but also illustrate a difficult and patient non-compliant dosage regimen of LSF attributed to its short half-life and rapid clearance. Thus, orally active formulation of the drug if available could provide a major relief to patients with T1DM.

## **7. Drug-fatty acid conjugates and delivery systems**

Oral drug delivery route is one of the most convenient and commonly used drug administration routes. But there are several drugs that exhibit poor bioavailability upon delivering them orally. One of the appropriate strategies used to address this problem is the use of drug-fatty acid conjugates. Drug-fatty acid conjugates are the drug molecules which have been covalently modified along with a fatty acids. Such conjugates have shown many benefits including enhanced oral bioavailability, improved tumor targeting, decreased toxicity and improved drug loading into delivery carriers.<sup>112-114</sup> Fatty acids consist of hydrocarbon chain and a reactive carboxylic acid group which have been conjugated with the drugs to make them hydrophobic. The fatty acids are used because of their characteristic properties such as biocompatibility, additional functional roles in drug targeting or self-assembly and chemical flexibility for modification. The widely used technique is to conjugate the carboxylic end of the fatty acid with a drug's hydroxyl or amine group to form a stable ester or amide linkage.<sup>115</sup> Many fatty acids and their derivatives were used for the development of conjugates (**Table 1.2**) and these conjugates have been delivered using different carriers including self-assembled systems (without carriers), liposomes, emulsions, lipid nanoparticles, micelles, and polymer NPs etc.

**Table 1.2**

Drug-fatty acid conjugates and their delivery systems.

S.No.	Fatty acid	Drug	Delivery system	Ref.
1	Conjugated linoleic acid	Paclitaxel	Liposome	116
2	Stearic acid	Gemcitabine	Polymer nanoparticle lipid nanoparticle, micelle	117,118,119
		5-fluorouracil	Lipid nanoparticle	120
3	Lauric acid	Cytarabine	Self-assembled nanofiber	121
4	Oleic acid	Docetaxel	Nanostructured lipid carrier	122
		Paclitaxel	Emulsion	123
		Imiquimod	Cream	124
5		Dexamethasone	Emulsion	125
		Paclitaxel	Emulsion	126
	Palmitic acid	Doxorubicin	Micelle	127
		TGX-221	Micelle	128
		siRNA	Polymer nanoparticle	129
		Capecitabine	Lipid nanoparticle	130
6		Doxorubicin	No carrier	131
		Paclitaxel	No carrier	132
	DHA	Lovastatin	No carrier	133
		10-hydroxycamptothecin	No carrier	134
7	Myristic acid	Cabotegravir	Nanoparticles	135
	Hexanoic acid, Octanoic acid, Decanoic acid, and Dodecanoic acid			136
8		Entecavir	No carrier	
9	Octadecanoic acid	Gemcitabine	Nanoassembly	137

### 7.1. Lisofylline (LSF): suitable candidate for conjugate formation

LSF is a potent hydrophilic drug with reported benefit in T1DM and its aqueous solubility is approx. 60 mg/mL which makes it difficult to be formulated into any nano drug delivery system. Apart from solubility, major concern with LSF is its interconversion to its parent drug pentoxifylline (PTX).<sup>138</sup> Delivery of LSF by any conventional approach fails to solve the problems of drug metabolism and its poor PK parameters. Interconversion of LSF

and PTX is mainly attributed to free hydroxyl group found in LSF side chain which is gets oxidized in the presence of oxidoreductase enzymes (**Figure 1.1**). Considering this, conjugation of LSF with a hydrophobic moiety such as a fatty acid appears to be a preferable strategy that could consume the free hydroxyl group of LSF and further it could be delivered using a nano-formulation. Conjugation of LSF with hydrophobic moieties like polymer or fatty acids can impart hydrophobicity to LSF, the resulting conjugate can be encapsulated into any nano drug delivery system and can also reduce the excessive metabolism of LSF providing an overall enhanced efficacy.

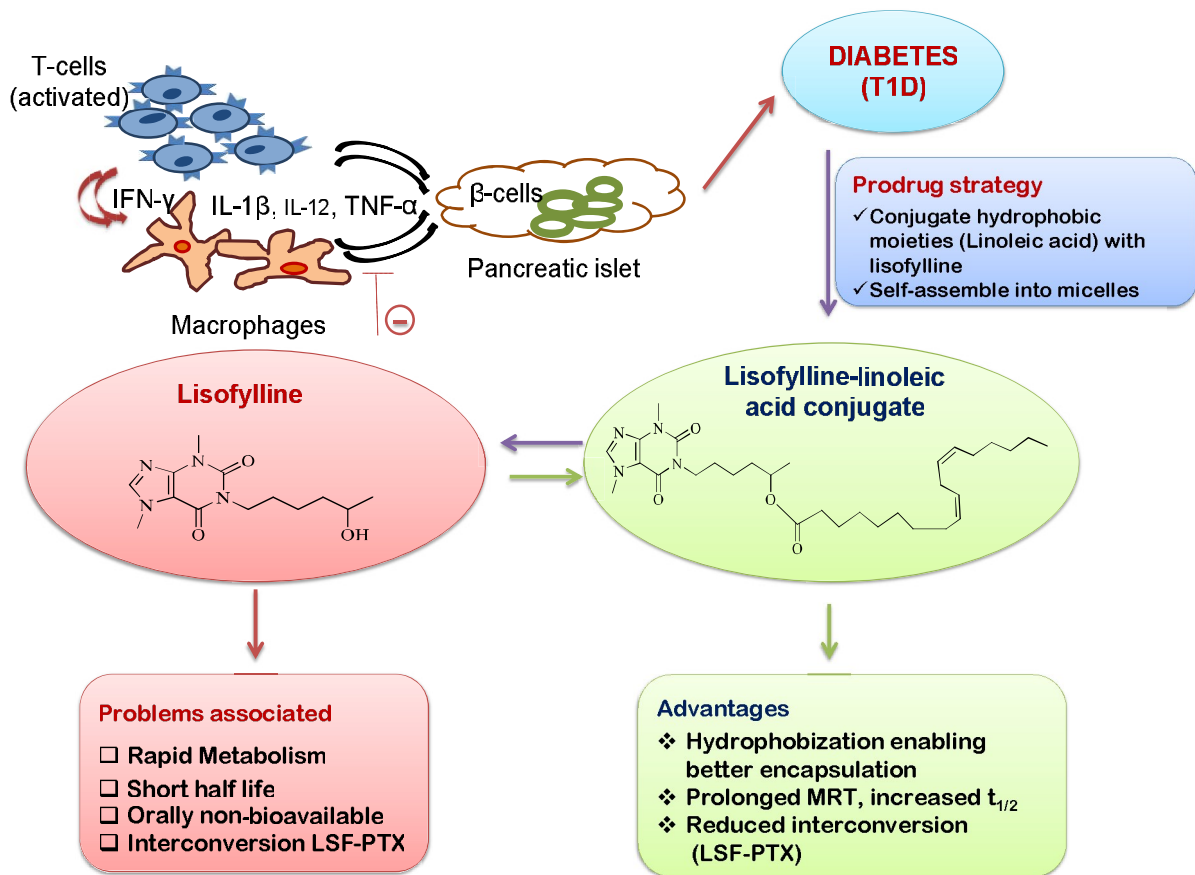
## **8. Objectives of the present research and development endeavor**

The objective of this work was to overcome the challenges associated with LSF by synthesizing LSF-fatty acid conjugate and to deliver it by a suitable nanoformulation for effective treatment of T1DM (**Figure 1.1**). The advantages of preparing a fatty acid conjugate of LSF are, *a) impart hydrophobicity to the drug to enable its efficient encapsulation into the delivery system and, b) reduce drug metabolism by protecting its hydroxyl group and thus prolonging its half-life*. Further, an oral delivery system for LSF-fatty acid conjugate nanoformulation was envisaged, developed and evaluated.

The specific goals of this research work are outlined below:

1. Analytical and bioanalytical method development and validation of LSF and PTX
2. Synthesis, characterization and evaluation of LSF-linoleic acid (LSF-LA) conjugate
  - i. Synthesis and characterization of LSF-LA
  - ii. Self-assembly of LSF-LA conjugate into micelles (LSF-LA SM)
  - iii. *In-vitro* evaluation of LSF-LA SM in cell culture
  - iv. Pharmacokinetics and *in-vivo* efficacy studies of LSF-LA SM in T1DM animal model

3. Development and evaluation of polymeric nanoformulation of LSF-LA conjugate
  - i. Development of polymeric micelle formulation of LSF-LA (LSF-LA PLM)
  - ii. *In-vitro* evaluation of LSF-LA PLM in cell culture
  - iii. Pharmacokinetics and *in-vivo* efficacy studies of LSF-LA PLM in T1DM animal model
4. Designing of an oral tablet dosage form of LSF-LA PLM
  - i. Scale-up and lyophilization of LSF-LA PLM
  - ii. Preparation and characterization of LSF-LA PLM tablets
  - iii. Pharmacokinetic studies of LSF-LA PLM tablets



**Figure 1.1** Mechanism of action of LSF in T1DM, problems associated with LSF and our proposed strategy and its advantages.



## **Bibliography**

1. Association, A. D. Diagnosis and classification of diabetes mellitus. *Diabetes care* **2014**, *37*, (Supplement 1), S81-S90.
2. Chen, L.; Magliano, D. J.; Zimmet, P. Z. The worldwide epidemiology of type 2 diabetes mellitus--present and future perspectives. *Nature reviews. Endocrinology* **2011**, *8*, (4), 228-36.
3. Wild, S.; Roglic, G.; Green, A.; Sicree, R.; King, H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes care* **2004**, *27*, (5), 1047-1053.
4. Williams, R.; Karuranga, S.; Malanda, B.; Saeedi, P.; Basit, A.; Besançon, S.; Bommer, C.; Esteghamati, A.; Ogurtsova, K.; Zhang, P. Global and regional estimates and projections of diabetes-related health expenditure: Results from the International Diabetes Federation Diabetes Atlas. *Diabetes Research and Clinical Practice* **2020**, 108072.
5. Kaveeshwar, S. A.; Cornwall, J. The current state of diabetes mellitus in India. *Australas Med J* **2014**, *7*, (1), 45-48.
6. Verma, P.; Pathak, K. Therapeutic and cosmeceutical potential of ethosomes: An overview. *J Adv Pharm Technol Res* **2010**, *1*, (3), 274-282.
7. Federation, I. D. *IDF Diabetes Atlas*; International Diabetes Federation: Brussels, Belgium, 2017.
8. Lowe, W. L., Diabetes mellitus. In *Principles of molecular medicine*, Springer: 1998; pp 433-442.
9. Association, A. D. 2. Classification and diagnosis of diabetes. *Diabetes care* **2017**, *40*, (Supplement 1), S11-S24.
10. Kauffman, T. L.; Scott, R. W.; Barr, J. O.; Moran, M. L., *A Comprehensive Guide to Geriatric Rehabilitation:[previously entitled Geriatric Rehabilitation Manual],Chapter 46 - Diabetes*. Elsevier Health Sciences: 2014.
11. Association, A. D. 2. Classification and diagnosis of diabetes: standards of medical care in diabetes—2018. *Diabetes care* **2018**, *41*, (Supplement 1), S13-S27.

12. Pozzilli, P.; Signore, A. The reconstructed natural history of type 1 diabetes mellitus. *Nature Reviews Endocrinology* **2019**, *15*, (5), 256-257.
13. Chatterjee, S.; Khunti, K.; Davies, M. J. Type 2 diabetes. *The Lancet* **2017**, *389*, (10085), 2239-2251.
14. Knip, M. Type 1 diabetes mellitus is a heterogeneous disease. *Nature Reviews Endocrinology* **2017**, *13*, (9), 1.
15. Fatima, N.; Faisal, S. M.; Zubair, S.; Ajmal, M.; Siddiqui, S. S.; Moin, S.; Owais, M. Role of pro-inflammatory cytokines and biochemical markers in the pathogenesis of type 1 diabetes: correlation with age and glycemic condition in diabetic human subjects. *PloS one* **2016**, *11*, (8).
16. Grunnet, L. G.; Mandrup-Poulsen, T. Cytokines and type 1 diabetes: a numbers game. *Diabetes* **2011**, *60*, (3), 697-699.
17. Knip, M. Diabetes: Loss of  $\beta$ -cell mass—an acute event before T1DM presentation? *Nature Reviews Endocrinology* **2017**, *13*, (5), 253.
18. Control, D.; Trial, C. Intensive diabetes treatment and cardiovascular outcomes in type 1 diabetes: the DCCT/EDIC study 30-year follow-up. *Diabetes care* **2016**, *39*, (5), 686-693.
19. Chaudhury, A.; Duvoor, C.; Reddy Dendi, V. S.; Kraleti, S.; Chada, A.; Ravilla, R.; Marco, A.; Shekhawat, N. S.; Montales, M. T.; Kuriakose, K.; Sasapu, A.; Beebe, A.; Patil, N.; Musham, C. K.; Lohani, G. P.; Mirza, W. Clinical Review of Antidiabetic Drugs: Implications for Type 2 Diabetes Mellitus Management. *Frontiers in Endocrinology* **2017**, *8*, 1-12.
20. Moller, D. E. New drug targets for type 2 diabetes and the metabolic syndrome. *Nature* **2001**, *414*, (6865), 821-827.
21. Lechleitner, M.; Hoppichler, F. Insulin therapy. *Wiener Medizinische Wochenschrift* **2011**, *161*, (11), 300-304.
22. Aljawarneh, Y. Associations between Physical Activity, Health-Related Quality of Life, Regimen Adherence, and Glycemic Control in Jordanian Adolescents with Type 1 Diabetes. **2018**.

23. Lachin, J. M.; Orchard, T. J.; Nathan, D. M. Update on Cardiovascular Outcomes at 30 Years of the Diabetes Control and Complications Trial/Epidemiology of Diabetes Interventions and Complications Study. *Diabetes care* **2014**, *37*, (1), 39-43.
24. Halland, M.; Bharucha, A. E. Relationship Between Control of Glycemia and Gastric Emptying Disturbances in Diabetes Mellitus. *Clinical Gastroenterology and Hepatology* **2016**, *14*, (7), 929-936.
25. Woerle, H. J.; Albrecht, M.; Linke, R.; Zschau, S.; Neumann, C.; Nicolaus, M.; Gerich, J. E.; Göke, B.; Schirra, J. Impaired Hyperglycemia-Induced Delay in Gastric Emptying in Patients With Type 1 Diabetes Deficient for Islet Amyloid Polypeptide. *Diabetes care* **2008**, *31*, (12), 2325-2331.
26. Kilpatrick, E. S.; Rigby, A. S.; Atkin, S. L. Insulin Resistance, the Metabolic Syndrome, and Complication Risk in Type 1 Diabetes. "Double diabetes" in the Diabetes Control and Complications Trial **2007**, *30*, (3), 707-712.
27. Conway, B.; Miller, R. G.; Costacou, T.; Fried, L.; Kelsey, S.; Evans, R. W.; Orchard, T. J. Temporal patterns in overweight and obesity in Type 1 diabetes. *Diabetic Medicine* **2010**, *27*, (4), 398-404.
28. Chapman, I.; Parker, B.; Doran, S.; Feinle-Bisset, C.; Wishart, J.; Strobel, S.; Wang, Y.; Burns, C.; Lush, C.; Weyer, C.; Horowitz, M. Effect of pramlintide on satiety and food intake in obese subjects and subjects with type 2 diabetes. *Diabetologia* **2005**, *48*, (5), 838-848.
29. Meier, J. J.; Nauck, M. A. Glucagon-like peptide 1(GLP-1) in biology and pathology. *Diabetes/Metabolism Research and Reviews* **2005**, *21*, (2), 91-117.
30. Madsbad, S.; Krarup, T.; Deacon, C. F.; Holst, J. J. Glucagon-like peptide receptor agonists and dipeptidyl peptidase-4 inhibitors in the treatment of diabetes: a review of clinical trials. *Current Opinion in Clinical Nutrition & Metabolic Care* **2008**, *11*, (4), 491-499.
31. Inzucchi, S. E.; Bergenstal, R. M.; Buse, J. B.; Diamant, M.; Ferrannini, E.; Nauck, M.; Peters, A. L.; Tsapas, A.; Wender, R.; Matthews, D. R. Management of hyperglycaemia in type 2 diabetes: a patient-centered approach. Position statement of the American Diabetes Association

- (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetologia* **2012**, *55*, (6), 1577-1596.
32. Wright, E. M.; Loo, D. D. F.; Hirayama, B. A. Biology of Human Sodium Glucose Transporters. *Physiological Reviews* **2011**, *91*, (2), 733-794.
  33. Seedher, N.; Kanojia, M. Co-solvent solubilization of some poorly-soluble antidiabetic drugs. *Pharm Dev Technol* **2009**, *14*, (2), 185-192.
  34. Cheng, C. L.; Lawrence, X. Y.; Lee, H. L.; Yang, C. Y.; Lue, C. S.; Chou, C. H. Biowaiver extension potential to BCS Class III high solubility-low permeability drugs: bridging evidence for metformin immediate-release tablet. *Eur J Pharm Sci* **2004**, *22*, (4), 297-304.
  35. Rojas, L. B. A.; Gomes, M. B. Metformin: an old but still the best treatment for type 2 diabetes. *Diabetol Metab Syndr* **2013**, *5*, (1), 6-21.
  36. Bhikshapathi, D.; Madhukar, P.; Kumar, B. D.; Kumar, G. A. Formulation and characterization of pioglitazone HCl self emulsifying drug delivery system. *Der Pharmacia Lettre* **2013**, *5*, (2), 292-305.
  37. Tripathi, K., Chapter 19-Insulin, oral hypoglycaemic drugs and glucagon. In *Essentials of medical pharmacology*, 7 ed.; JP Medical Ltd: New Delhi, 2013; pp 258-281.
  38. Kassem, A. A.; El-Alim, S. H. A.; Basha, M.; Salama, A. Phospholipid complex enriched micelles: A novel drug delivery approach for promoting the antidiabetic effect of repaglinide. *Eur J Pharm Sci* **2017**, *99*, 75-84.
  39. Araujo, F.; Fonte, P.; Santos, H. A.; Sarmiento, B. Oral delivery of glucagon-like peptide-1 and analogs: alternatives for diabetes control? *J Diabetes Sci Technol* **2012**, *6*, (6), 1486-1497.
  40. Kalra, S. Glucagon-like peptide-1 receptors agonists (GLP1 RA). *J Pak Med Assoc* **2013**, *63*, (10), 1312-1315.
  41. Filippatos, T. D.; Panagiotopoulou, T. V.; Elisaf, M. S. Adverse effects of GLP-1 receptor agonists. *Rev Diabet Stud* **2014**, *11*, (3-4), 202-230.
  42. Hamidi, M.; Azadi, A.; Rafiei, P. Hydrogel nanoparticles in drug delivery. *Adv Drug Deliv Rev* **2008**, *60*, (15), 1638-1649.

43. Mudshinge, S. R.; Deore, A. B.; Patil, S.; Bhalgat, C. M. Nanoparticles: emerging carriers for drug delivery. *Saudi pharmaceutical journal* **2011**, *19*, (3), 129-141.
44. De Jong, W. H.; Borm, P. J. A. Drug delivery and nanoparticles: Applications and hazards. *International Journal of Nanomedicine* **2008**, *3*, (2), 133-149.
45. Mudshinge, S. R.; Deore, A. B.; Patil, S.; Bhalgat, C. M. Nanoparticles: emerging carriers for drug delivery. *Saudi Pharm J* **2011**, *19*, (3), 129-141.
46. Bhatia, S., Nanoparticles types, classification, characterization, fabrication methods and drug delivery applications. In *Natural Polymer Drug Delivery Systems*, Springer International Publishing, 2016; pp 33-93.
47. Pathak, K.; Raghuvanshi, S. Oral bioavailability: issues and solutions via nanoformulations. *Clin Pharmacokinets* **2015**, *54*, (4), 325-357.
48. Soudry-Kochavi, L.; Naraykin, N.; Nassar, T.; Benita, S. Improved oral absorption of exenatide using an original nanoencapsulation and microencapsulation approach. *J Control Release* **2015**, *217*, 202-210.
49. Aslam, M.; Aqil, M.; Ahad, A.; Najmi, A. K.; Sultana, Y.; Ali, A. Application of Box–Behnken design for preparation of glibenclamide loaded lipid based nanoparticles: optimization, in vitro skin permeation, drug release and in vivo pharmacokinetic study. *J. Mol. Liq* **2016**, *219*, 897-908.
50. Liu, H.; Shang, K.; Liu, W.; Leng, D.; Li, R.; Kong, Y.; Zhang, T. Improved oral bioavailability of glyburide by a self-nanoemulsifying drug delivery system. *J Microencapsul* **2014**, *31*, (3), 277-283.
51. Mohd, A. B.; Sanka, K.; Bandi, S.; Diwan, P. V.; Shastri, N. Solid self-nanoemulsifying drug delivery system (S-SNEDDS) for oral delivery of glimepiride: development and antidiabetic activity in albino rabbits. *Drug deliv* **2015**, *22*, (4), 499-508.
52. Rani, R.; Dahiya, S.; Dhingra, D.; Dilbaghi, N.; Kim, K. H.; Kumar, S. Evaluation of anti-diabetic activity of glycyrrhizin-loaded nanoparticles in nicotinamide-streptozotocin-induced diabetic rats. *Eur J Pharm Sci* **2017**, *106*, 220-230.

53. Shariatinia, Z.; Zahraee, Z. Controlled release of metformin from chitosan-based nanocomposite films containing mesoporous MCM-41 nanoparticles as novel drug delivery systems. *J Colloid Interface Sci* **2017**, *501*, 60-76.
54. Kumar, S.; Bhanjana, G.; Verma, R. K.; Dhingra, D.; Dilbaghi, N.; Kim, K. H. Metformin-loaded alginate nanoparticles as an effective antidiabetic agent for controlled drug release. *The Journal of pharmacy and pharmacology* **2017**, *69*, (2), 143-150.
55. Hasan, A. A.; Madkor, H.; Wageh, S. Formulation and evaluation of metformin hydrochloride-loaded niosomes as controlled release drug delivery system. *Drug deliv* **2013**, *20*, (3-4), 120-126.
56. Alam, S.; Aslam, M.; Khan, A.; Imam, S. S.; Aqil, M.; Sultana, Y.; Ali, A. Nanostructured lipid carriers of pioglitazone for transdermal application: From experimental design to bioactivity detail. *Drug deliv* **2016**, *23*, (2), 601-609.
57. Akhtar, J.; Siddiqui, H. H.; Fareed, S.; Badruddeen; Khalid, M.; Aqil, M. Nanoemulsion: for improved oral delivery of repaglinide. *Drug Deliv* **2016**, *23*, (6), 2026-2034.
58. Li, X.; Wang, C.; Liang, R.; Sun, F.; Shi, Y.; Wang, A.; Liu, W.; Sun, K.; Li, Y. The glucose-lowering potential of exenatide delivered orally via goblet cell-targeting nanoparticles. *Pharm Res* **2015**, *32*, (3), 1017-1027.
59. Gadadare, R.; Mandpe, L.; Pokharkar, V. Ultra rapidly dissolving repaglinide nanosized crystals prepared via bottom-up and top-down approach: influence of food on pharmacokinetics behavior. *AAPS PharmSciTech* **2015**, *16*, (4), 787-799.
60. Rege, B. D.; Kao, J. P.; Polli, J. E. Effects of nonionic surfactants on membrane transporters in Caco-2 cell monolayers. *Eur J Pharm Sci* **2002**, *16*, (4-5), 237-246.
61. Elbahwy, I. A.; Ibrahim, H. M.; Ismael, H. R.; Kasem, A. A. Enhancing bioavailability and controlling the release of glibenclamide from optimized solid lipid nanoparticles. *J Drug Deliv Sci Technol* **2017**, *38*, 78-89.
62. Shi, Y.; Sun, X.; Zhang, L.; Sun, K.; Li, K.; Li, Y.; Zhang, Q. Fc-modified exenatide-loaded nanoparticles for oral delivery to improve hypoglycemic effects in mice. *Sci Rep* **2018**, *8*, (1), 726-735.

63. Singer, J. W.; Rursten, S. L.; Rice, G. C.; Gordon, W. P.; Bianco, J. A. Inhibitors of intracellular phosphatidic acid production: novel therapeutics with broad clinical applications. *Expert Opin Investig Drugs* **1994**, *3*, (6), 631-644.
64. Waxman, K.; Daughters, K.; Aswani, S.; Rice, G. Lisofylline decreases white cell adhesiveness and improves survival after experimental hemorrhagic shock. *Crit Care Med.* **1996**, *24*, (10), 1724-1728.
65. Abraham, E.; Bursten, S.; Shenkar, R.; Allbee, J.; Tuder, R.; Woodson, P.; Guidot, D. M.; Rice, G.; Singer, J. W.; Repine, J. E. Phosphatidic acid signaling mediates lung cytokine expression and lung inflammatory injury after hemorrhage in mice. *J Exp Med.* **1995**, *181*, (2), 569-575.
66. Masiello, P.; Broca, C.; Gross, R.; Roye, M.; Manteghetti, M.; Hillaire-Buys, D.; Novelli, M.; Ribes, G. Experimental NIDDM: development of a new model in adult rats administered streptozotocin and nicotinamide. *Diabetes* **1998**, *47*, (2), 224-229.
67. Leahy, J.; Bonner-Weir, S.; Weir, G. Abnormal glucose regulation of insulin secretion in models of reduced B-cell mass. *Diabetes* **1984**, *33*, (7), 667-673.
68. Bursten, S. L.; Federighi, D.; Wald, J.; Meengs, B.; Spickler, W.; Nudelman, E. Lisofylline causes rapid and prolonged suppression of serum levels of free fatty acids. *J Pharmacol Exp Ther.* **1998**, *284*, (1), 337-345.
69. Cui, P.; Macdonald, T. L.; Chen, M.; Nadler, J. L. Synthesis and biological evaluation of lisofylline (LSF) analogs as a potential treatment for Type 1 diabetes. *Bioorg Med Chem Lett.* **2006**, *16*, (13), 3401-3405.
70. Chen, M.; Yang, Z.; Wu, R.; Nadler, J. L. Lisofylline, a novel antiinflammatory agent, protects pancreatic  $\beta$ -cells from proinflammatory cytokine damage by promoting mitochondrial metabolism. *Endocrinology* **2002**, *143*, (6), 2341-2348.
71. Desagher, S.; Martinou, J.-C. Mitochondria as the central control point of apoptosis. *Trends in cell biology* **2000**, *10*, (9), 369-377.

72. Striffler, J. S.; Nadler, J. L. Lisofylline, a novel anti-inflammatory agent, enhances glucose-stimulated insulin secretion in vivo and in vitro: studies in prediabetic and normal rats. *Metabolism* **2004**, *53*, (3), 290-296.
73. Tersey, S. A.; Carter, J. D.; Rosenberg, L.; Taylor-Fishwick, D. A.; Mirmira, R. G.; Nadler, J. L. Amelioration of type 1 diabetes following treatment of non-obese diabetic mice with INGAP and lisofylline. *Journal of diabetes mellitus* **2012**, *2*, (2), 251.
74. Yang, Z.; Chen, M.; Fialkow, L. B.; Ellett, J. D.; Wu, R.; Nadler, J. L. The novel anti-inflammatory compound, lisofylline, prevents diabetes in multiple low-dose streptozotocin-treated mice. *Pancreas* **2003**, *26*, (4), e99-e104.
75. Balon, T. W.; Jasman, A. P.; Bursten, S. L.; Nadler, J. L. Lisofylline, a modulator of fatty acid metabolism, increases peripheral insulin sensitivity. *Diabetes* **1999**, *48*, (5), SA95-SA95.
76. Yang, Z.; Chen, M.; Carter, J. D.; Nunemaker, C. S.; Garmey, J. C.; Kimble, S. D.; Nadler, J. L. Combined treatment with lisofylline and exendin-4 reverses autoimmune diabetes. *Biochem Biophys Res Commun.* **2006**, *344*, (3), 1017-1022.
77. Yang, Z.; Chen, M.; Nadler, J. L. Lisofylline: a potential lead for the treatment of diabetes. *Biochem Pharmacol.* **2005**, *69*, (1), 1-5.
78. Wyska, E.; Pękala, E.; Szymura, O., Joanna. Interconversion and tissue distribution of pentoxifylline and lisofylline in mice. *Chirality* **2006**, *18*, (8), 644-651.
79. National Institutes of Health, A Safety, Tolerability and Bioavailability Study of Lisofylline After Continuous Subcutaneous (12 mg/kg) and Intravenous (9 mg/kg) Administration in Subjects With Type 1 Diabetes Mellitus. <https://clinicaltrials.gov/ct2/show/NCT01603121> (25 April 2019),
80. List, A.; Maziarz, R.; Stiff, P.; Jansen, J.; Liesveld, J.; Andrews, F.; Schuster, M.; Wolff, S.; Litzow, M.; Karanes, C. A randomized placebo-controlled trial of lisofylline in HLA-identical, sibling-donor, allogeneic bone marrow transplant recipients. *Bone Marrow Transplant.* **2000**, *25*, (3), 283-291.



81. Lee, W. W.; Lu, F., Therapeutic calcium phosphate particles and methods of making and using same. Google Patents: 2012.
82. Sung, H.-W.; Sonaje, K.; Liao, Z.-X.; Hsu, L.-W.; Chuang, E.-Y. pH-responsive nanoparticles shelled with chitosan for oral delivery of insulin: from mechanism to therapeutic applications. *Accounts of chemical research* **2012**, *45*, (4), 619-629.
83. Yang, J.; Sun, H.; Song, C. Preparation, characterization and in vivo evaluation of pH-sensitive oral insulin-loaded poly (lactic-co-glycolicacid) nanoparticles. *Diabetes, Obesity and Metabolism* **2012**, *14*, (4), 358-364.
84. Eldor, R.; Arbit, E.; Corcos, A.; Kidron, M. Glucose-reducing effect of the ORMD-0801 oral insulin preparation in patients with uncontrolled type 1 diabetes: a pilot study. *PloS one* **2013**, *8*, (4).
85. Zhang, X.; Sun, M.; Zheng, A.; Cao, D.; Bi, Y.; Sun, J. Preparation and characterization of insulin-loaded bioadhesive PLGA nanoparticles for oral administration. *European Journal of Pharmaceutical Sciences* **2012**, *45*, (5), 632-638.
86. Damgé, C.; Socha, M.; Ubrich, N.; Maincent, P. Poly ( $\epsilon$ -caprolactone)/eudragit nanoparticles for oral delivery of aspart-insulin in the treatment of diabetes. *Journal of pharmaceutical sciences* **2010**, *99*, (2), 879-889.
87. Sonaje, K.; Lin, K.-J.; Wey, S.-P.; Lin, C.-K.; Yeh, T.-H.; Nguyen, H.-N.; Hsu, C.-W.; Yen, T.-C.; Juang, J.-H.; Sung, H.-W. Biodistribution, pharmacodynamics and pharmacokinetics of insulin analogues in a rat model: oral delivery using pH-responsive nanoparticles vs. subcutaneous injection. *Biomaterials* **2010**, *31*, (26), 6849-6858.
88. Appana Chowdary, K.; Suravarapu, N. L. R.; Meddala, S. Formulation and characterization of pioglitazone hydrochloride nanoparticles. *World J Pharm Pharm Sci* **2015**, *4*, (4), 1638-1648.
89. Deshpande, R.; Gowda, D.; Vegesna, N. S. K. V.; Vaghela, R.; Kulkarni, P. The effect of nanonization on poorly water soluble glibenclamide using a liquid anti-solvent precipitation technique: aqueous solubility, in vitro and in vivo study. *RSC Adv* **2015**, *5*, (99), 81728-81738.

90. Chen, C.; Zheng, H.; Xu, J.; Shi, X.; Li, F.; Wang, X. Sustained-release study on Exenatide loaded into mesoporous silica nanoparticles: in vitro characterization and in vivo evaluation. *Daru* **2017**, *25*, (1), 1-8.
91. Pandey, V.; Kohli, S. SMEDDS of pioglitazone: Formulation, in-vitro evaluation and stability studies. *Future Journal of Pharmaceutical Sciences* **2017**, *3*, (1), 53-59.
92. Agrawal, A. G.; Kumar, A.; Gide, P. S. Self emulsifying drug delivery system for enhanced solubility and dissolution of glipizide. *Colloids Surf B Biointerfaces* **2015**, *126*, 553-560.
93. Bari, A.; Chella, N.; Sanka, K.; Shastri, N. R.; Diwan, P. V. Improved anti-diabetic activity of glibenclamide using oral self nano emulsifying powder. *J Microencapsul* **2015**, *32*, (1), 54-60.
94. Li, H.; Pan, T.; Cui, Y.; Li, X.; Gao, J.; Yang, W.; Shen, S. Improved oral bioavailability of poorly water-soluble glimepiride by utilizing microemulsion technique. *International Journal of Nanomedicine* **2016**, *11*, 3777-3788.
95. Zhang, Z.-H.; Zhang, Y.-L.; Zhou, J.-P.; Lv, H.-X. Solid lipid nanoparticles modified with stearic acid–octaarginine for oral administration of insulin. *International Journal of Nanomedicine* **2012**, *7*, 3333.
96. Yang, R.; Gao, R.-C.; Cai, C.-F.; Xu, H.; Li, F.; He, H.-B.; Tang, X. Preparation of gel-core-solid lipid nanoparticle: a novel way to improve the encapsulation of protein and peptide. *Chemical and Pharmaceutical Bulletin* **2010**, *58*, (9), 1195-1202.
97. Gonçalves, L. M. D.; Maestrelli, F.; Di Cesare Mannelli, L.; Ghelardini, C.; Almeida, A. J.; Mura, P. Development of solid lipid nanoparticles as carriers for improving oral bioavailability of glibenclamide. *Eur J Pharm Biopharm* **2016**, *102*, 41-50.
98. Ebrahimi, H. A.; Javadzadeh, Y.; Hamidi, M.; Barzegar Jalali, M. Development and characterization of a novel lipohydrogel nanocarrier: repaglinide as a lipophilic model drug. *The Journal of pharmacy and pharmacology* **2016**, *68*, (4), 450-458.
99. Shrestha, N.; Bouttefeux, O.; Vanvarenberg, K.; Lundquist, P.; Cunarro, J.; Tovar, S.; Khodus, G.; Andersson, E.; Keita, Å. V.; Dieguez, C. G. The stimulation of GLP-1 secretion and delivery of GLP-1 agonists via nanostructured lipid carriers. *Nanoscale* **2018**, *10*, (2), 603-613.

100. Lau, J. R.; Geho, W. B., Orally bioavailable lipid-based constructs. Google Patents: 2015.
101. Mohsen, A. M.; AbouSamra, M. M.; ElShebiney, S. A. Enhanced oral bioavailability and sustained delivery of glimepiride via niosomal encapsulation: in-vitro characterization and in-vivo evaluation. *Drug Dev Ind Pharm* **2017**, *43*, (8), 1254-1264.
102. Joshi, S.; Hussain, M. T.; Roces, C. B.; Anderluzzi, G.; Kastner, E.; Salmaso, S.; Kirby, D. J.; Perrie, Y. Microfluidics based manufacture of liposomes simultaneously entrapping hydrophilic and lipophilic drugs. *Int J Pharm* **2016**, *514*, (1), 160-168.
103. Nam, J.-P.; Choi, C.; Jang, M.-K.; Jeong, Y.-I.; Nah, J.-W.; Kim, S.-H.; Park, Y. Insulin-incorporated chitosan nanoparticles based on polyelectrolyte complex formation. *Macromolecular Research* **2010**, *18*, (7), 630-635.
104. Ahmed, T. A.; Khalid, M.; Aljaeid, B. M.; Fahmy, U. A.; Abd-Allah, F. I. Transdermal glimepiride delivery system based on optimized ethosomal nano-vesicles: Preparation, characterization, in vitro, ex vivo and clinical evaluation. *Int J Pharm* **2016**, *500*, (1-2), 245-254.
105. Ali, H. S.; Hanafy, A. F. Glibenclamide Nanocrystals in a Biodegradable Chitosan Patch for Transdermal Delivery: Engineering, Formulation, and Evaluation. *J Pharm Sci* **2017**, *106*, (1), 402-410.
106. Agrawal, A. K.; Harde, H.; Thanki, K.; Jain, S. Improved stability and antidiabetic potential of insulin containing folic acid functionalized polymer stabilized multilayered liposomes following oral administration. *Biomacromolecules* **2014**, *15*, (1), 350-360.
107. Yao, Y.; Zhao, L.; Yang, J.; Yang, J. Glucose-responsive vehicles containing phenylborate ester for controlled insulin release at neutral pH. *Biomacromolecules* **2012**, *13*, (6), 1837-1844.
108. Mirazi, N.; Shoaie, J.; Khazaei, A.; Hosseini, A. A comparative study on effect of metformin and metformin-conjugated nanotubes on blood glucose homeostasis in diabetic rats. *Eur J Drug Metab Pharmacokinet* **2015**, *40*, (3), 343-348.
109. Tong, F.; Tang, X.; Li, X.; Xia, W.; Liu, D. The effect of insulin-loaded linear poly (ethylene glycol)-brush-like poly (l-lysine) block copolymer on renal ischemia/reperfusion-induced lung

- injury through downregulating hypoxia-inducible factor. *International Journal of Nanomedicine* **2016**, *11*, 1717-1730.
110. Tong, F. Preparation of exenatide-loaded linear poly (ethylene glycol)-brush poly (l-lysine) block copolymer: potential implications on diabetic nephropathy. *International Journal of Nanomedicine* **2017**, *12*, 4663-4678.
111. Wiedemann, H. P.; Arroliga, A. C.; Komara, J.; Denver, V.; Welsh, C.; Fulkerson, W. J.; MacIntyre, N.; Mallatratt, L.; Sebastian, M.; Sladen, R. Randomized, placebo-controlled trial of lisofylline for early treatment of acute lung injury and acute respiratory distress syndrome. *Crit. Care Med.* **2002**, *30*, (1), 1-6.
112. Irby, D.; Du, C.; Li, F. Lipid–drug conjugate for enhancing drug delivery. *Molecular pharmaceutics* **2017**, *14*, (5), 1325-1338.
113. Thanki, K.; Prajapati, R.; Sangamwar, A. T.; Jain, S. Long chain fatty acid conjugation remarkably decreases the aggregation induced toxicity of Amphotericin B. *International journal of pharmaceutics* **2018**, *544*, (1), 1-13.
114. Thanki, K.; Date, T.; Jain, S. Improved oral bioavailability and gastrointestinal stability of amphotericin B through fatty acid conjugation approach. *Molecular pharmaceutics* **2019**, *16*, (11), 4519-4529.
115. Date, T.; Paul, K.; Singh, N.; Jain, S. Drug–Lipid Conjugates for Enhanced Oral Drug Delivery. *AAPS PharmSciTech* **2019**, *20*, (2), 41.
116. Du, R.; Zhong, T.; Zhang, W.-Q.; Song, P.; Song, W.-D.; Zhao, Y.; Wang, C.; Tang, Y.-Q.; Zhang, X.; Zhang, Q. Antitumor effect of iRGD-modified liposomes containing conjugated linoleic acid–paclitaxel (CLA-PTX) on B16-F10 melanoma. *International Journal of Nanomedicine* **2014**, *9*, 3091.
117. Gupta, A.; Asthana, S.; Konwar, R.; Chourasia, M. An insight into potential of nanoparticles-assisted chemotherapy of cancer using gemcitabine and its fatty acid prodrug: a comparative study. *Journal of biomedical nanotechnology* **2013**, *9*, (5), 915-925.

118. Chung, W.-G.; Sandoval, M. A.; Sloat, B. R.; Lansakara-P, D. S.; Cui, Z. Stearoyl gemcitabine nanoparticles overcome resistance related to the over-expression of ribonucleotide reductase subunit M1. *Journal of controlled release* **2012**, *157*, (1), 132-140.
119. Wang, Y.; Fan, W.; Dai, X.; Katragadda, U.; Mckinley, D.; Teng, Q.; Tan, C. Enhanced tumor delivery of gemcitabine via PEG-DSPE/TPGS mixed micelles. *Molecular pharmaceutics* **2014**, *11*, (4), 1140-1150.
120. Yu, B.-T.; Sun, X.; Zhang, Z.-R. Enhanced liver targeting by synthesis of N 1-stearyl-5-Fu and incorporation into solid lipid nanoparticles. *Archives of pharmacal research* **2003**, *26*, (12), 1096-1101.
121. Liu, J.; Zhao, D.; Ma, N.; Luan, Y. Highly enhanced leukemia therapy and oral bioavailability from a novel amphiphilic prodrug of cytarabine. *RSC advances* **2016**, *6*, (42), 35991-35999.
122. Sun, B.; Luo, C.; Li, L.; Wang, M.; Du, Y.; Di, D.; Zhang, D.; Ren, G.; Pan, X.; Fu, Q. Core-matched encapsulation of an oleate prodrug into nanostructured lipid carriers with high drug loading capability to facilitate the oral delivery of docetaxel. *Colloids and Surfaces B: Biointerfaces* **2016**, *143*, 47-55.
123. Lundberg, B.; Risovic, V.; Ramaswamy, M.; Wasan, K. A lipophilic paclitaxel derivative incorporated in a lipid emulsion for parenteral administration. *Journal of controlled release* **2003**, *86*, (1), 93-100.
124. Sharma, A.; Sharma, D.; Baldi, A.; Jyoti, K.; Chandra, R.; Madan, J. Imiquimod-oleic acid prodrug-loaded cream reduced drug crystallinity and induced indistinguishable cytotoxicity and apoptosis in mice melanoma tumour. *Journal of Microencapsulation* **2019**, *36*, (8), 759-774.
125. Daull, P.; Paterson, C. A.; Kuppermann, B. D.; Garrigue, J.-S. A preliminary evaluation of dexamethasone palmitate emulsion: a novel intravitreal sustained delivery of corticosteroid for treatment of macular edema. *Journal of ocular pharmacology and therapeutics* **2013**, *29*, (2), 258-269.

126. Goldstein, D.; Gofrit, O.; Nyska, A.; Benita, S. Anti-HER2 cationic immunoemulsion as a potential targeted drug delivery system for the treatment of prostate cancer. *Cancer research* **2007**, *67*, (1), 269-275.
127. Li, F.; Snow-Davis, C.; Du, C.; Bondarev, M. L.; Saulsbury, M. D.; Heyliger, S. O. Preparation and characterization of lipophilic doxorubicin pro-drug micelles. *JoVE (Journal of Visualized Experiments)* **2016**, (114), e54338.
128. Zhao, Y.; Duan, S.; Zeng, X.; Liu, C.; Davies, N. M.; Li, B.; Forrest, M. L. Prodrug strategy for PSMA-targeted delivery of TGX-221 to prostate cancer cells. *Molecular pharmaceutics* **2012**, *9*, (6), 1705-1716.
129. Sarett, S. M.; Kilchrist, K. V.; Miteva, M.; Duvall, C. L. Conjugation of palmitic acid improves potency and longevity of siRNA delivered via endosomolytic polymer nanoparticles. *Journal of Biomedical Materials Research Part A* **2015**, *103*, (9), 3107-3116.
130. Gong, X.; Moghaddam, M. J.; Sagnella, S. M.; Conn, C. E.; Danon, S. J.; Waddington, L. J.; Drummond, C. J. Lamellar crystalline self-assembly behaviour and solid lipid nanoparticles of a palmityl prodrug analogue of Capecitabine—A chemotherapy agent. *Colloids and Surfaces B: Biointerfaces* **2011**, *85*, (2), 349-359.
131. Wang, Y.; Li, L.; Jiang, W.; Yang, Z.; Zhang, Z. Synthesis and preliminary antitumor activity evaluation of a DHA and doxorubicin conjugate. *Bioorganic & medicinal chemistry letters* **2006**, *16*, (11), 2974-2977.
132. Bradley, M. O.; Webb, N. L.; Anthony, F. H.; Devanesan, P.; Witman, P. A.; Hemamalini, S.; Chander, M. C.; Baker, S. D.; He, L.; Horwitz, S. B. Tumor targeting by covalent conjugation of a natural fatty acid to paclitaxel. *Clinical Cancer Research* **2001**, *7*, (10), 3229-3238.
133. Siddiqui, R. A.; Harvey, K. A.; Xu, Z.; Natarajan, S. K.; Davisson, V. J. Characterization of lovastatin-docosahexaenoate anticancer properties against breast cancer cells. *Bioorganic & medicinal chemistry* **2014**, *22*, (6), 1899-1908.
134. Wang, Y.; Li, L.; Jiang, W.; Larrick, J. W. Synthesis and evaluation of a DHA and 10-hydroxycamptothecin conjugate. *Bioorganic & medicinal chemistry* **2005**, *13*, (19), 5592-5599.

135. Zhou, T.; Su, H.; Dash, P.; Lin, Z.; Dyavar Shetty, B. L.; Kocher, T.; Szlachetka, A.; Lamberty, B.; Fox, H. S.; Poluektova, L.; Gorantla, S.; McMillan, J.; Gautam, N.; Mosley, R. L.; Alnouti, Y.; Edagwa, B.; Gendelman, H. E. Creation of a nanoformulated cabotegravir prodrug with improved antiretroviral profiles. *Biomaterials* **2018**, *151*, 53-65.
136. Jung, H. J.; Ho, M. J.; Ahn, S.; Han, Y. T.; Kang, M. J. Synthesis and physicochemical evaluation of entecavir-fatty acid conjugates in reducing food effect on intestinal absorption. *Molecules* **2018**, *23*, (4), 731.
137. Jin, Y.; Lian, Y.; Du, L. Self-assembly of N-acyl derivatives of gemcitabine at the air/water interface and the formation of nanoscale structures in water. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* **2012**, *393*, 60-65.
138. Wyska, E.; Pękala, E.; Szymura-Oleksiak, J. Interconversion and tissue distribution of pentoxifylline and lisofylline in mice. *Chirality* **2006**, *18*, (8), 644-651.



This document was created with the Win2PDF "print to PDF" printer available at <http://www.win2pdf.com>

This version of Win2PDF 10 is for evaluation and non-commercial use only.

This page will not be added after purchasing Win2PDF.

<http://www.win2pdf.com/purchase/>