

T H E S I S

SOME BIOCHEMICAL STUDIES OF  
CARBONDISULFIDE TOXICITY IN RATS

Submitted for the degree of -  
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(in requirement of partial fulfilment)

by

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TO

HUMANITY

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LOVE

BIRLA INSTITUTE OF TECHNOLOGY AND SCIENCE, PILANI (RAJASTHAN )..

C E R T I F I C A T E

This is to certify that the thesis entitled  
"SOME BIOCHEMICAL STUDIES OF CARBONDISULFIDE TOXICITY  
IN RATS" and submitted by SUBHASH CHANDER SHARMA  
ID No. 76S87501, for the award of Ph.D. degree of  
the institute,embodies original work done by him  
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## ABBREVIATIONS

CS <sub>2</sub>	Carbondisulfide
CA	Catecholamines
DA	Dopamine
NE	Norepinephrine
5-HT	5-hydroxytryptamine
MAO	Monoamine Oxidase
HMP	Hexose monophosphate Shunt
de-dtc	diethyl- dithio carbamate
dm-dtc	dimethyl dithio carbamate
edtc	ethyl dithio carbamates
G-SH	Reduced glutathione
FDPase	Fructose, 1-6 diphosphatase
ATPase	Adenosine Triphosphatase
SDH	Succinic dehydrogenase
ANSA	Alpha amino naphthyl - sulfonic acid
TTC	2,4,6 - Triphenyl Tetrazolium Chloride

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In the biosphere, pollution is caused by substances that exist in nature i.e the whole range of semisynthetic or synthetic substances like molasses, polymers, industrial solvents etc. The industrial pollutants are spread over through deliberate or careless handling of raw materials or waste products. Although nature is a readymade laboratory to deal fully with the first category of substances spread over by physical and natural resources, the other category of pollutants pose a complex problem since nature cannot deal fully to remove them. The rapid introduction of more and more new compounds due to industrialization has further complicated the problem. The ecotoxicological problems arising out of this situation have demanded scientific probe in this direction.

Carbondisulfide ( $CS_2$ ), an inorganic compound capable of dissolving many organic substances, is a very useful solvent. Its main use in the industry is as a chemical intermediate in the xanthation of cellulose in the viscose rayon manufacturing and as a fumigant. Besides, it is encountered most frequently in the chemical laboratories where it is extensively used as a solvent. The low boiling point of  $CS_2$  impregnates the working environment with its vapours, causing thereby the problem of pollution and a serious health hazard to mankind. The main risks of  $CS_2$  are by vapour inhalation and fire explosion. Carbondisulfide is reputed for



for its central nervous system depressant effects i.e. reduction of motor activity, lethargy etc. Coma and death follow excessive inhalation, though instances of such a serious consequence are very rare. However, persistent alteration of central and peripheral nervous system functions have been reported to occur as a result of single massive over exposure. A series of less severe exposures (Teisinger, 1972), may also result in similar alterations. Symptoms range from headache, fatigue and listlessness to loss of memory, anorexia and reduction in sexual function. In more severe cases, cardiovascular disorders, hallucinations and other symptoms resembling psychosis have been reported. Of these, the more interesting manifestations of chronic CS<sub>2</sub> intoxication is loss of reflexes; giving first indication of nerve damage. ( Lieben, 1974).

The workers who are exposed to very high concentrations of CS<sub>2</sub> have been reported to manifest similar neurological symptoms and cardiovascular disorders ( Seppalainen et.al. 1966 ; Tiller et.al. 1968). These symptoms and signs have been known to be directly related to the intensity and period of exposure to the intoxicant. Only few attempts have been made so far to elucidate the mechanisms of CS<sub>2</sub> hazards in a living system. These studies have also not clearly shown any relationship between the mechanism of CS<sub>2</sub> intoxication and biochemical picture of an animal. Therefore, In the present

work, some biochemical modifications due to CS<sub>2</sub> toxicity have been studied to correlate the mechanism of CS<sub>2</sub> toxicity with carbohydrate metabolism aberrations. In the present work various biochemical parameters dealt with, using rats as test organisms are :

1. Alteration in carbohydrate metabolism and its enzymes in liver and brain.
2. Alterations in tissues biogenic amines level under CS<sub>2</sub> toxicity.
3. Interrelationship of tissues biogenic amines with monoamine oxidase activity and reduced glutathione ( G-SH ).
4. Alterations in trace element metabolism under the stress of carbondisulfide.

The thesis is divided into six chapters. First chapter deals with the review of the existing literature and the second with the materials and methods adopted for carrying the biochemical studies. The third chapter includes the results and observations while the fourth chapter is devoted to discussion. The fifth chapter includes a summary of the work and conclusions drawn from this study. The sixth chapter lists relevant references pertaining to the present work.

## 1.1 EPIDEMIOLOGICAL STUDIES OF CARBONDISULFIDE TOXICITY :-

The problem evolved on the human health from carbondisulfide toxicity gained recent extensive scientific impetus from the presentation of toxicological data of carbondisulfide at the International Symposium at Prague (1966). A major recommendation of the Symposium was to conduct the comprehensive epidemiological studies on the morbidity and mortality of workers exposed to toxic vapours in the viscose rayon industry for several years ( Brieger, 1967; Viglaini, 1967)

Cavalleri et al (1966), on the endocrinological findings in the young workers exposed to carbondisulfide, reported an increased urinary excretion of 17- ketosteroids and 17-hydroxy-steroids. Savic (1967) reported pathological changes in the eyes caused by carbondisulfide. Tolonen et al (1975) found the disturbed ocular microcirculation and suggested that occupational exposures to carbondisulfide were not the basis of the intensity of physiological abnormalities but the severity of manifestations of vasculopathy was related to the quantity of CS<sub>2</sub> in environment. The safety margin of CS<sub>2</sub> recommended by American Conference of Governmental Industrial Hygeinists is 20 ppm. The incidences of death by coronary heart disease in males due to exposure of carbondisulfide were higher than that deaths in unexposed men due to this abnormality and other unexposed workers of the same



industry ( Tiller et al, 1968; Herneberg et al, 1970; Tolonen et al , 1975). In an another study Herneberg and Norman (1973) confirmed the finding in both sexes. The risk of death rose with increasing exposure. These findings confirmed the earlier mortality studies, and strongly supported the hypothesis of causal relationship between CS<sub>2</sub> exposures and coronary heart diseases.

Martinez and Farina (1969), in a clinical examinations of seventy five workers exposed to high levels of CS<sub>2</sub> for many years in viscose factories reported 83 percent workers having vasculopathy in encephalic region, 22 percent having disturbance of renal activity, 5 percent having renal hypertension, and 7 percent of diabetes mellitus. Higher incidences of angina pectoris and hypertension with electrocardio graphic abnormalities were also observed by Herneberg et al (1970). The incidence of diabetes mellitus among workers exposed to the risk of carbondisulfide poisoning were more than the normal persons (Ferreiro 1969). Szott, et al (1969) estimated the urinary excretion of uropepsin and reported an increase of uropepsin from 46-92 units per hour, the latter value being close to that observed in cases with pepsin ulceration. These studies further, showed some correlation between uropepsin elimination and duration of exposure to carbondisulfide.

Hanninen (1971), performed a battery of psychological tests to three different groups of workers, one with carbon-disulfide poisoning, second exposed to carbondisulfide but without known poisoning and the third group consisted of unexposed workers. A prominent and significant reduction in performances involving speed vigilance, manual dexterity and intelligence was noticed in the first two groups. The group exposed to only low CS<sub>2</sub> concentration showed impairment but the changes were less severe in comparison to poisoned group. Further, in this work the poisoned group could reliably be distinguished from the exposed workers in two ways i.e. latent and manifest. The latent and manifest poisoning differed not only in intensity but also in quality as, latent poisoning was characterised by traits indicative of depressive mood, slight motor disturbances and intellectual impairments whereas clinically manifested poisoning resulted in lowered vigilance, diminished intellectual activity, diminished rational control, retarded speed and motor disturbances. Seppalainen et al (1974) also reported the neurotoxicity of long term exposure to CS<sub>2</sub> by measuring the maximum motor conduction velocity and conduction velocity of slower motor fibers.

Herneberg et al (1969) studied plasma and erythrocyte changes in zinc and magnesium levels in Finish and Norwegian workers exposed to CS<sub>2</sub> to study the racial variations. The



magnesium levels were slightly reduced in erythrocytes whereas plasma magnesium level increased in exposed Finnish workers while, zinc concentration is lowered slightly. No such differences in plasma and erythrocyte magnesium and zinc were found in Norwegian men by these workers. Carbondisulfide also caused depletion of serum zinc by an increase in rate of zinc excretion, and an increase in all serum protein fractions. The effects were temporary and improved on cessation of exposure ( EI' Gazzar et al 1973).

Herneberg et al (1971) examined blood lipids, glucose tolerance, and plasma creatinine in viscose rayon industry workers as a part of planned study of diseases. This study included workers exposed to an average available CS<sub>2</sub>, 20-40 ppm in 1950's and 10-30 ppm in 1960's. The only difference reported between the exposed workers and matched controls were in fasting glucose; the differences bear a direct correlation with the exposure time and exposure index and in mean plasma creatinine which was higher in exposed group.

Goto et al. (1971) selected exposed workers from 11-Japanese viscose rayon plants and reported a higher blood sugar levels trend in advance age group of exposed workers. A corticosteroid priming glucose tolerance test indicated a subclinical defect in carbohydrate metabolism resembling a mild diabetogenic action in some apparently healthy carbondisulfide exposed workers.



The incidences of prevalence of retinal microaneurisms which depended on the exposure to carbondisulfide were also reported to be increased. Higher blood sugar levels with increasing grade of microaneurisms had been reported. Unfortunately no information to the intensity of carbondisulfide exposure and its relation to microaneurisms has been reported.

Mancuso and Locke (1972) studied social aspects of workers in CS<sub>2</sub> environment at one of the rayon factory and found an excess of suicides in workers first employed between the years 1938 and 1948. In addition to an increased cases of recorded suicides, there were a number of deaths recorded due to other causes which strongly resembled suicides.

Sakurai (1972) reported temporary increase of blood pressure in a Japanese viscose rayon plant for more than ten years and found reduction of blood pressure, when workers were removed from carbondisulfide environment to an improved environment. Prevovska and Zvolisky (1973), reported that continuous exposure to carbondisulfide could make the persons as invalid and deaths had been due to organic complications of vascularsclerosis. They therefore, interpreted CS<sub>2</sub> as a possible atherogenic agent.

Murashko (1974) observed the deleterious effect of carbon-disulfide on digestive organs of workers at viscose fibre plant and suggested a special diet for workers exposed to carbondisulfide.

Herneberg and Tolonen ( 1976) reported that the altered conditions with the excess of  $CS_2$  could bring about reversible risk for future fatal attacks of coronary heart diseases which were minimized by removing the persons from  $CS_2$  exposure. This study, therefore, reflected that  $CS_2$  exert its direct toxic effect on myocardium. Andrazewska et al. (1976) examined visual space localization for upper and lower extremities and the trunk in man. The neuropathological action of  $CS_2$  on these organs and parts of the body was attributed again to its direct manifestations in these body organs. Sugimoto et al. (1976) reported temporary retinopathy due to  $CS_2$  in workers exposed for a long duration which disappeared or improved after cessation from  $CS_2$  environment.

The incidence of angina and the increase of blood pressure i.e., the diastolic and systolic, were significantly higher among the exposed workers and race dependant because it was shown in this study Finish workers were more prone to  $CS_2$  toxicity than Japanese. The E.C.G. studies showed that the left ventricular ejection time was shortened and isovolumic contraction time was prolonged in workers exposed industrially to carbondisulfide for a period of eighteen years ( Franco, 1976). The possible mechanism of this cardiac defect was attributed to the interference of  $CS_2$  with Kreb's Cycle or with catecholamine metabolism.



Gullko and Pikisllskaya (1977) revealed further, that prolonged contact with  $CS_2$  promoted the development of cardiovascular disorders like unstable arterial pressure with tendency to hypertension, an increase of propagation rate of the pulse wave in elastic blood vessels, dilation and intensification of the aortal shadow. Disorders of the lipid metabolism are a possible precondition for the development of early atherosclerosis. Therefore, they suggested that prolonged effect of carbondisulfide may lead to the development of early atherosclerosis. Tarlov et al. (1977) reported that cardiac disorders depended on the functional condition of the nervous system and were nonspecific. They also found typical neurotic functional cardiopathy and dystrophic changes in 521 workers of viscose rayon factory, exposed to  $CS_2$ . For these patients they recommended, beta-blocking agents and tranquilizers.

## 1.2 EXPERIMENTAL STUDIES OF $CS_2$ TOXICITY :-

### 1.2.1 HISTOLOGICAL STUDIES OF $CS_2$ TOXICITY :-

Carbondisulfide poisoning achieved by administering  $CS_2$  in vegetable oil for three weeks did not produce much deformation but a considerable restorative process in somatic tissue was observed (Klishov et al. 1970). However, Barilyerk and Vasileva (1975) studied the effect of small concentration of  $CS_2$  and hydrogen sulfide on intrauterine development of rats. They reported that the exposure of female rats to  $CS_2$  before and

during gestation was lethal to embryos at the pre and post implantation stages. The effect of CS<sub>2</sub> on embryonic stage was also noticed when male rats were inhaled with CS<sub>2</sub> before mating with untreated females. The embryonic abnormality due to CS<sub>2</sub> was suggested to the defect caused in the genitourinary and bone systems of embryo. Disturbances in ossification, blood formation and dystrophic changes in liver and Kidneys were also observed.

Gondzik (1976) reported irreversible changes in microstructure of gonads and in seminiferous tubules of testis by CS<sub>2</sub>. However, he observed that the concentration of CS<sub>2</sub> is an important factor as there were no alterations in the testicles and kidneys of the rat after exposure to lower concentration vapors of air for 20 weeks and in man working in an atmosphere of CS<sub>2</sub> containing 0.02- 0.06 mg CS<sub>2</sub>/ liter.

Seppalainen et al. (1976) examined the electrophysiological findings in rat given chronic carbondisulfide inhalation. The development of neuropathy was monitored bi-monthly by measurement of motor conduction velocity in sciatic nerve. A slight but statistically significant slowing of motor conduction velocity was observed on second week. No further reduction occurred during the next three weeks but after 8 to 10 weeks the difficulties in voluntary control of hind limbs



with marked slowing of motor conduction velocity was demonstrated. However, permanent alteration in motor conduction velocity did not occur in animals exposed to carbondisulfide for five weeks.

Greenberg (1977) reported the ulcer formation at the site of injection of  $CS_2$  subcutaneously, which increased in size after the last injection. The lesion seen with the first or second dose caused the infiltration of neutrophils in the epidermis around the ulcer and lateral dermal invasion from ulcer margin by hyperplastic epidermal cell masses.

Zajaczek et al (1977) explored the effects of  $CS_2$  on chromosomes structure in vitro and mitotic capability of leukocytes in man and rat. A complete inhibition of mitosis in the cultures of peripheral blood leukocytes and some structural disturbances in chromosomes were observed. Solecka et al . (1977) reported a significant increase in the agglutination test of leukocytes in rabbits breathing carbondisulfide vapors for many days. Mihalache et al.(1977) reported the myocardial lesions similar to those in myocardial ischemia without vascular lesions whose severity was directly related to the increased exposure time. This experiment therefore, suggested that the lesions may result from a direct toxic action of carbondisulfide on myocardial fibres or from inhibition of myocardial circulation.

Juntunen Japanese et al. (1977) showed demonstrable acetylcholinesterase activity by CS<sub>2</sub> intoxication which was distributed in myoneural junctions of both exposed and unexposed rats. In both groups, intense enzyme activity was localised at the level of postsynaptic membrane of myoneural junction. No enzyme activity has been seen outside the zone of myoneural junctions. In the terminal axons, signs of various degrees of degeneration were also reported to be present with widened synaptic clefts because of schwann cells interposition. Apparently, CS<sub>2</sub> poisoning primarily altered the presynaptic structures of myoneural junctions whereas postsynaptic side remained relatively intact.

Dietzmann & Lass ( 1977) reported myelin sheath disruptions within longitudinal track systems of the spinal cord, disruptions of individual ganglion cells in all brain regions, and elective parenchymal necrosis in frontal and parietal cerebral cortices. The histochemical assays of monoamine oxidase, ATPase, glucose phosphatase, acetylcholinesterase and succinic dehydrogenase activities in the entire central nervous system revealed no change in the enzymes activities however, succinic-dehydrogenase and acetylcholinesterase revealed reduction in the activities within elective parenchymal necrotic region.

Wronska-Nofer et al. (1978) reported four fold elevation of total ( free and esterified ) serum cholesterol



with a simultaneous rise in phospholipids. In rabbits exposed to carbondisulfide and fed with atherogenic diet increase in cholesterol was 40 to 100 percent and triglyceride was 100 percent. Gross atheromatosis of the aorta and enhanced histocoronary lesions were observed as a result of CS<sub>2</sub> intoxication. Further, evidence was presented of an enhanced susceptibility of arterial wall to the atherogenic diet under the influence of chronic CS<sub>2</sub> intoxication with special reference to the individual regional susceptibility of arterial coronary vessels. But Verigin et al. (1978) reports on the development of atherosclerotic process under experimental conditions, showed that CS<sub>2</sub> did not alter blood serum cholesterol, phospholipids or triglycerides level in rabbits maintained on either normal or cholesterol supplemented diet. No morphological changes indicative of atherosclerotic process were observed after CS<sub>2</sub> treatment and normal diet.

### 1.2.2. BIOCHEMICAL STUDIES OF CARBONDISULFIDE TOXICITY :-

#### 1.2.2.1 ENZYMATIC CHANGES DUE TO CS<sub>2</sub> TOXICITY :-

In the recent years much of the interesting toxicological studies have been directed to the biochemical studies to underline the cause of various disturbances produced by carbondisulfide. Hockin et al. (1968) observed reversible biochemical changes in lens and retina of rabbit eyes which was related to the period of exposure to CS<sub>2</sub>

of the animals. The ATP content of retina was reduced by 8 percent and 22 percent after 7 days and 14 days respectively. In the lens the activity of aldolase, glucose 6- phosphate dehydrogenase, hexokinase and glucokinase were decreased and those of malate dehydrogenase, glutamate- pyruvic transminase, sorbitel dehydrogenase were increased shortly after CS<sub>2</sub> treatment. However, the activities of malate dehydrogenase, sorbitol dehydrogenase had returned to normal level after 14 days of treatment. The irreversible changes had been suggested to be important for the genesis of cataract.

Bond and Matteis (1969) reported that 1 ml/kg oral dose of CS<sub>2</sub> persistently suppressed the O- dealkylation of p- nitroanisole and hydroxylations of phenobarabital and aniline in rats with rapid loss of hepatic cytochrome P- 450 content which resulted in an increased liver size and protein content. They proposed that the prolongation of phenobarbital sleeping time by CS<sub>2</sub> was due to suppression of this enzyme.

Bond et al. (1969) studied the interference of drug metabolism by CS<sub>2</sub> exposure. They suggested that a metabolite probably a dithiocarbamate, rather than free CS<sub>2</sub> itself, could be the causative agent for the liver microsomal depressions. Prior treatment with phenobarbital allow CS<sub>2</sub> to be metabolised more rapidly in animals and potentiation of hepatotoxicity of CS<sub>2</sub> was observed. Freundt and Dreher (1969) noticed the depression of N- demethylase and nitroreductase following CS<sub>2</sub>



vapor inhalation at low concentrations independently of morphological changes in the liver. Prior equilibration of rats to air levels of  $CS_2$  significantly prolonged hexobarbital induced hypnosis and depressed the microsomal oxidation or demethylation of drugs which was suggestive by the reduced urinary elimination of metabolites like 4- amino antipyrine and trichloroethylene after administration of antipyrine.

Mack and Freundt (1970) stated that inhibition of drug metabolising enzymes activity by  $CS_2$  was reversible, regression of the depressed enzyme activity and excretion deficits occurred soon after the cessation of exposure. Small amount of inhaled  $CS_2$  also produced depression in the rate of aminopyrine N- demethylation, as reflected by reduced 4- amino antipyrine excretion.

Caligaris et al. (1972) reported that approximately 45% of labelled  $CS_2$  ( $S^{35}$ ) bound to the microsomes incubated with CN, and was released in a form which appeared identical to SCN. Incubation of microsomes with unlabelled  $CS_2$  followed by incubation with  $CN^{14}$  also resulted in the recovery of what appeared to be  $C^{14}$  labelled SCN. A portion of 'S' released in microsomal metabolism of  $CS_2$  had been suggested with- SH groups of cysteine residues in microsomal proteins to form hydrosulfide.

Murasko & Gubskis ( 1976) reported the modification of liver and small intestine enzymes in rats chronically poisoned with carbondisulfide. A decrease in the activities of alpha-keto- glutaric dehydrogenase and succinic dehydrogenase in liver mitochondria with disrupted membrane digestive function of small intestine was indicated by reduction in dipeptidase activity and glucose absorption. The reduction in liver and intestinal enzymes activity were more severe with higher concentrations of CS<sub>2</sub> inhaled.

Tereshchenko et al. (1976) reported a decrease in erythrocyte--SH group level in persons poisoned by CS<sub>2</sub>, whereas erythrocytic -SH group content increased in healthy workers, if exposed to CS<sub>2</sub> for a long time.

Ol'khovskaya and Kazan (1976) found the change in the activity of blood enzymes in patients with chronic CS<sub>2</sub> poisoning, especially in those with polyneuritis symptoms. The serum and erythrocyte levels of cholinesterase, aldolase, ceruplasmin and total and reduced glutathione decreased as compared with those of normal subjects.

Sidorowicz et al.(1977) found a reduction in haemoglobin concentration, osmotic resistance and ATP level where as AMP, ADP and 2,3 diphosphoglycerate levels increased corroborating the haemolytic effect of CS<sub>2</sub> in human beings exposed chronically to

CS<sub>2</sub>. The disturbances in the activity of key enzymes of anaerobic glycolytic cycle and decrease in ATP level caused hemolysis. The increased levels of 2,3 diphosphoglycerate showed a profound tissue hypoxia which was suggested to be a compensating mechanism. The dysfunction of anaerobic glycolysis was observed to reduce the erythrocyte function which was contemplated with neurological disturbances and premature atheromatosis.

Savolainen and Jarvisalo (1977), reported the specific binding of CS<sub>2</sub> metabolites to microsomal proteins in rat liver. Binding of CS<sub>2</sub> by microsomal proteins of phenobarbitone pretreated and untreated rats following an intraperitoneal injection of radiolabelled CS<sub>2</sub> indicated that the principal binding microsomal protein of CS<sub>2</sub> after its oxidative decomposition was cytochrome P- 450.

ELL - Dessoukey et al. (1977) examined the serum enzyme changes associated with CS<sub>2</sub> hepatotoxicity in experimental animals. Rats treated with CS<sub>2</sub> showed elevation of serum GPT, GOT and serum alkaline - phosphatase. Rats treated similarly showed a slight to moderate rise in serum lactate dehydrogenase.

Jarvisalo and Savolainen (1977) found that the binding of S<sup>35</sup> was considerably higher than that of C<sup>14</sup>, three hour after injection. There was a significant fall in P- 450 whereas microsomal epoxide hydrolase activity of UDP glucuronyl transferase increased significantly in phenobarbitone pretreated rats.



They suggested that changes in P-450 results from the binding of metabolites of  $CS_2$  to cytochrome and its subsequent degradation. The increase in UDP glucuronyl transferase resulted probably from activated perturbation in the structure of microsomal membranes by metabolism of carbondisulfide.

Wagner et al. (1977) examined the levels of alpha-I antitrypsin in children from areas of different air pollution. They had found that lowering of alpha-I antitrypsin activity was associated mainly with high levels of  $SO_2$  and  $CS_2$ .

Murashko et al. (1977), studied the effect of inhaled  $CS_2$  on digestive enzymes. Carbondisulfide was found to disrupt enterokinase formation by small intestine, inhibition of enterokinase and alkaline phosphatase activity in large intestine which caused the appearance of mucus and nonalimentary proteins in the feces and disrupted regeneration process in the large and small intestines. Also it disrupted glucose absorption and inhibited the intestinal enzymes responsible for the hydrolysis of disaccharide and polysaccharides.

Dalvi and Howell (1978), reported no alterations in the concentration of P-450 cytochrome from liver microsomes by  $CS_2$ , malathione and parathione in phenobarbitone pretreated animals. But inhibition of oxidative desulfuration was recorded by the insecticides by microsomes from  $CS_2$  treated animals was attributed to the prior binding with P-450 of 'S' released from  $CS_2$  as



confirmed from the results of spectral binding of SKF 525- A, parathione and malathione. Thus CS<sub>2</sub>, parathione and malathione undergoes oxidative desulfuration in an analogous manner and the metabolism of the insecticides is impaired in the liver previously exposed to CS<sub>2</sub>.

Merkur'eva, et al. (1978) observed the lysosomal and cytoplasmic enzyme changes in liver, kidneys, brain, aorta and blood serum, and carbohydrate levels in rabbits exposed to CS<sub>2</sub> for six weeks. These changes were indicative of possible neurotropic and hepatotropic activity of carbondisulfide.

Jarvisalo et al . (1978) concluded that the accelerated liver heme conversion to bile pigments caused by CS<sub>2</sub> and other sulfur containing drugs is related to the increased metabolism by oxidative desulfuration and ensuring the microsomal toxicity. The damage of the apoprotein of cytochrome P- 450 might have attained a reduced affinity for heme, thus, causing more liver heme being available for degradation.

#### 1.2.2.2. LIPIDS AND CS<sub>2</sub> TOXICITY :-

Wronska- Nofer (1966) studied the role of CS<sub>2</sub> in inducing the disturbances in lipid metabolism. Rats were exposed to CS<sub>2</sub> following forced feeding of 0.2 mg of cholesterol C<sup>14</sup>. The blood cholesterol levels at six, twelve, and twentyfour hours later were higher in CS<sub>2</sub> exposed rats. Differences were evident at 12 weeks and twice as great after 30 weeks. Carbondisulfide

increased the absorption of cholesterol from intestinal tract, and concomitantly speeded up its excretion via the bile.

Makarova et al. (1974) examined the effect of sulfur containing organic compounds on lipid metabolism in presence of CS<sub>2</sub>. The level of cholesterol and beta-lipoproteins was higher in the blood of workers exposed to CS<sub>2</sub> but methionine, 2-mercapto-benzathiazole, tetramethyl-thio-carbamoyl disulfide caused the protection to the increased cholesterol level.

Wronska Nofer (1976) studied the aortic phospholipid synthesis in rat administered CS<sub>2</sub> toxic dose. The rate of incorporation of phosphate precursor into phospholipids was investigated in normal and in CS<sub>2</sub> intoxicated group of rat. The specific activity of P<sup>32</sup> in phospholipids was measured in plasma and the aortic wall at various times after intravenous administration of 0.2 mcu phosphate. In CS<sub>2</sub> intoxicated rat the specific activity of aorta phospholipids was more than the normal rat. According to the suggestion of these workers, rise of aortic phospholipid activity probably, was not derived from increased blood phospholipid concentration but was attributed to an increased rate of phospholipid synthesis in aorta tissue.

Wronska- Nofer (1977) reported slight reduction in C<sup>14</sup> steroid excretion after 4-C<sup>14</sup> cholestenol administration at the end of seven months. This was primarily due to the decrease



of  $C^{14}$  bile acid excretion. However,  $C^{14}$  - cholesterol in feces was 35% higher in the exposed animals. But  $1-C^{14}$  was higher in liver and blood of exposed rats after  $1-C^{14}$  cholesterol administration. Laurman et al. (1977) also reported loss of body weight, increased cholesterol levels in aortic wall and skeletal muscles but no change in liver cholesterol level was observed.

Jarvisalo et al. (1977) commenting on the deleterious effects of subacute  $CS_2$  exposure on mouse liver thought that liver copper and phospholipid levels were increased. They reported interestingly, that microsomal enzyme activities were partially or totally restored inspite of continued exposure to  $CS_2$ . The diene conjugation of liver phospholipids enhanced which indicated, therefore, that the  $CS_2$  exposure damaged the membrane lipids continuously. The return of the enzyme activities to normal values had been attributed to decreased sensitivity to  $CS_2$  metabolites or to increased synthesis of enzymes. The increased diene conjugation of liver phospholipids indicated that the lipid environment of membrane bound enzymes changed during subacute exposure to carbondisulfide.

Cunningham & Holt (1977) reported a small but significant decrease in the rate of utilization of plasma free fatty acid (FFA). This decreased availability of FFA probably led to the increased catabolism of aminoacids and a large increase in urea



production. Wesolowska et al. (1978) observed the changes in the content of free and esterified cholesterol in serum and aorta. These changes were similar as observed in experimental sclerosis.

Balabaeven and Tabakova (1979) reported that during the entire period of pregnancy, carbondisulfide inhalation raised the level of FFA in the liver of mothers and decreased the level of triglycerides and phospholipids both in mother and fetus . They concluded that in mothers the level of FFA was most sensitive to carbondisulfide whereas in fetus the level of triglycerides was more labile than cholesterol.

#### 1.2.2.3. PROTEINS AND CARBONDISULFIDE TOXICITY :-

Woyke (1969) reported that DNA and RNA content of human liver was decreased in the cytoplasmic fraction and nuclei fractions by CS<sub>2</sub> injection. But there was a rise of DNA with the fall of RNA in neutrophils and lymphocytes.

Savolainen and Jarvisalo (1977) noticed the increased activity of rat brain acid proteinase. It accompanied with the changes in leucine C<sup>14</sup> turnover as well as in RNA content from 1 to 4 hour , after exposure. The changes were, however, more conspicuous in cerebellum than in other parts of the brain. The activities of creatinkinase, nonspecific cholinesterase, displaced only subtle changes as seen in cerebral homogenates and serum. These reports, thus, are indicative for the transient changes of brain protein metabolism with a single dose of carbondisulfide.

Jarvisalo et al. (1977) observed that there was an initial  $C^{14}$  - leucine uptake inhibition in phenobarbitone pretreated rat. This decreased  $C^{14}$  - leucine uptake was accompanied by decreased RNA levels and led them to conclude that  $CS_2$  affected protein synthesis as well as protein turnover .

EL'Dessoukey et al. (1977) reported a state of hypoaminoacidemia and hyperaminoacid urea in various groups of rat treated with toxic dose of  $CS_2$  for various weeks. However, this effect was transient as the cessation of  $CS_2$  intoxication led to the improvement of amino acid patterns of urine and blood in rat.

#### 1.2.2.4 CARBOHYDRATES AND $CS_2$ TOXICITY :-

There is a paucity in the literature about the full evidences regarding carbondisulfide effect on carbohydrate metabolism. However, Minden et al. (1970) reported a typical condition of tissue acidosis in  $CS_2$  poisoning with hypolacticemia. Kurzinger and Freundt (1969) reported an increase in the hepatic lactate and glycogen diminution.

Cunningham and Holt (1977) studied the turnover of blood glucose in the rat after acute  $CS_2$  intoxication and reported a reduction in blood glucose concentration in  $CS_2$  treated rats as compared to control. These studies made them to suggest that the rate of gluconeogenesis was unchanged in  $CS_2$  treated but starved rats while lowering of blood glucose in  $CS_2$  treated non-starved rats is due to an increase in futile cycling at the triosephosphate stage of glycolysis, rather than increased clearance of



glucose from blood.

#### 1.2.2.5. BIOGENIC AMINES AND CS<sub>2</sub> TOXICITY :-

Aleksander et al. (1966) studied the behaviour of certain biogenic amines in subjects exposed to CS<sub>2</sub> and recorded an increase in elimination of 5- hydroxyindole acetic acid in urine of those workers exposed to toxic levels of CS<sub>2</sub>. They also reported an increase of the blood levels of various catecholamines.

Maj and Vetulani (1970) studied the pharmacological properties of N-N disubstituted dithiocarbamates and their effect on brain catecholamines and reported a reduction in the brain norepinephrine (NE) content and spontaneous activity. Dimethyldithiocarbamate (dmdtc) and diethyldithiocarbamate (dedtc) elevated the brain dopamine level whereas dibutyldithiocarbamate (dbdte) depressed it. Dimethyldithiocarbamate and dedtc potentiated the convulsive effect of pentalenetetrazole whereas dedtc exerted a protective effect showing thereby that ethyldithiocarbamate(edtc) is not formed during CS<sub>2</sub> poisoning.

Maj Jerzy et al. (1970) also studied the effect of disulfiram, de-dtc and dm-dtc on serotonin and 5 HIAA levels in rat brain and reported that at lower dose of de-dtc and dm-dtc did not change the levels of serotonin and 5 HIAA significantly in brain. However, higher dose of de-dtc increased the 5 HIAA content by 143% without effecting the serotonin content but still



higher dose of de-dtc decreased the serotonin content without effecting the level of 5 HIAA . Disulfiram and dm-dtc decreased serotonin and 5-HIAA in the animals. Ethyldithio-carbamate decreased the serotonin content of hippocampus by 74% while it increased the serotonin content in hypothalamus by 72%. They concluded that all the sulfur containing substances effect the serotonin level in different ways and pattern.

Magos and Jarvis (1970) reported that rats exposed to 2 mg CS<sub>2</sub>/litter of air for two days showed a reduction in their brain NE and elevation in DA concentration. Exposure of ten days further, decreased the NE level but DA concentration returned to control level. Dopamine concentration in adrenals after 10 days of exposure were 67% higher than control animals suggesting a reduction in DA beta-hydroxylase enzyme. Exposure to CS<sub>2</sub> did not effect tyrosine level and MAO-activity in the brain, whereas, they found that sod-de-dtc inhibited the MAO activity as well as increased the tyrosine level.

Magos and Jarvis ( 1970) in another work studied the effects of diethyldithiocarbamate and CS<sub>2</sub> on brain tyrosine levels. They found that de-dtc increased the level of DA and tyrosine whereas decreased NE level in brain. Tyrosine level like that of DA was maximal 1 hour after the injection of de-dtc and the increase was followed by a rapid decrease to 80% of the control value. A similar two phased change in brain

level of tyrosine was also observed after repeated exposure to CS<sub>2</sub>. The increased DA level in the 1st phase of their action might be due to slowed conversion of tyrosine to dopa suggesting due to stimulation of feed back mechanisms, resulting in a subsequent decrease in brain NE and DA levels.

Magos (1971) reported further, the amphetamine induced stereotype in CS<sub>2</sub> exposure and diethyl dithiocarbamate treated male rats. CS<sub>2</sub> exposure increased brain DA levels and decreased the NE levels. Pretreatment of Sod. diethyl dithiocarbamate increased the duration but not the intensity of subsequent amphetamine induced behaviour.

Diliberto and Distifano (1973) examined the effects of 2-mercaptoethyl guanidine and other compounds on NE synthesis by adrenal medullary granules. This compound like reserpine phosphate inhibited NE formation from 1-C<sup>14</sup> labelled DA by bovine adrenal medullary granules. Reserpine prevented DA uptake by granules whereas 2-mercapto ethylguanidine inhibited dopamine beta-hydroxylase. The presence of copper was found to stimulate dopamine beta-hydroxylase. Diliberto and Distefano (1975) studied the mechanism and kinetics of the inhibition of dopamine beta-hydroxylase (DBH) by 2-mercaptoethyl guanidine and suggested that 2- mols of inhibitor combines with 2 Cu<sup>++</sup> ions in the active site and thus inhibit DBH. The intramolecular distance between anionic site of inhibitor and beta-hydroxylation site of dopamine was coinciding with the intramolecular distance between the



positive charge of the inhibitor and the site of binding of dopamine beta-hydroxylase.

Andrea and Guido (1974) reported the effect of chronic environmental stress on 5-HT receptors of smooth muscles and suggested that exposure of newly born animals to a chronic stress induced a faster maturation of 5-HT receptors sensitivity with reduced activity of MAO.

Stankovic et al. (1975) studied the effect of CS<sub>2</sub> and found that the amount of 5-HIAA in the urine of exposed workers was about 4 to 6 times higher than in the control group. It was therefore, concluded by these workers that CS<sub>2</sub> exert its marked effect on tryptophan metabolism.

Michael and Victor (1977) proposed a mechanism for the action of carbondisulfide according to which inhibition of DBH activity by CS<sub>2</sub> was mediated by the action of dithiocarbamates formed from the endogenous amino acids and catechol amines.

#### 1.2.2.6 TRACE ELEMENTS AND CS<sub>2</sub> TOXICITY :-

Djuric et al (1968) reported the significant increased excretion of zinc in the urine of persons exposed to carbon-disulfide in viscose rayon industry.

Massoud (1977) reported the effect of CS<sub>2</sub> intoxication on the levels of serum copper and ceruplasmin. Intramuscular injection of CS<sub>2</sub> in rat slightly increased serum copper and ceruplasmin



levels but prolonged treatment of  $CS_2$  caused these values to be decreased due to intoxication caused by the loss of appetite in animals. After the cessation of  $CS_2$  administration, animals regained their appetites and showed a slight reversal of serum copper and ceruplasmin though the levels were still below normal which suggested local action rather than the effect on serum values.

EL'Dessoukey and Awadallah (1977) reported that serum zinc, iron, calcium, magnesium levels decreased, while serum potassium levels increased under the effect of  $CS_2$  intoxication. The reduction in serum zinc, iron, calcium and magnesium levels had been due to a decrease in dietary minerals because of appetite loss and elevated potassium levels caused by tissue destruction. They reported these changes to be reversible.

Michael and Victor (1977) studied the binding of dithiocarbamates with copper of the enzyme dopamine- beta-hydroxylase, while these dithiocarbamates are formed by the reaction of  $CS_2$  with free amino groups available in cellular environment. They stated that it is the complexing of copper with dithiocarbamates which caused the inhibition of copper containing enzymes.

#### 1.2.2.7. PROPOSED MODE OF ACTION OF CARBONDISULFIDE :-

Stripp et al. (1969) reported that  $CS_2$  undergoes partial metabolic conversion into dithiocarbamates. The suspicion that microsomal inhibition by  $CS_2$  depended on the formation of these

metabolites has been strengthened by the fact that dithiocarbamates or related substances behaved as active microsomal inhibitors thereby inducing diminished tolerance to many drugs.

Tessinger (1971) discussed the mechanism of  $CS_2$  toxicity in relation to pyridoxine or vitamin  $B_6$  deficiency. They stressed that  $CS_2$  toxicity caused the deficiency of Vitamin  $B_6$  and its impaired metabolism caused the inhibition of monoamineoxidase and related enzymes.

Matteis and Seawright (1976) reported the liver toxicity of  $CS_2$  and possible significance of its metabolism by oxidative desulfuration. They concluded that reactive sulfur liberated during the oxidative desulfuration of  $CS_2$  might bound to cellular components and initiated toxic changes in the liver. This accounted for the loss of cytochrome P-450 and also for the centrilobular hydrophic degeneration which was also seen in vivo. Savolainen and Vainio (1976) examined the high binding of sulfur of  $CS_2$  in spinalcord axonal fraction. The sulfur was found to be bound to spinal cord axons in larger fractions than by spinal cord homogenate. In contrast, the carbon from  $CS_2$  was not significantly bound by axons. So they reported that toxic effect of  $CS_2$  may be due to sulfur containing intermediates formed in the nervous system.

Savolainen and Jarvisalo (1977) studied the binding of  $CS_2$  in CNS of control and phenobarbitone pretreated rats.

They found that more sulfur atoms were bound to rat brain. The phenobarbitone pretreatment increased the cerebral binding of sulfur and decreased that of carbon. This finding suggested that a considerable amount of CS<sub>2</sub> is retained in the nervous system and phenobarbitone pretreatment may also alter the brain metabolism of carbondisulfide.

Meckenna et al (1977) described the metabolism of inhaled CS<sub>2</sub> in rat. They found that elimination of CS<sub>2</sub> from the rat during the post exposure period was rapid and occurred primarily through kidneys. Accumulation of these acid labile metabolites upon repeated exposures and their presence in all tissues suggested that they may play a role in the toxicity of carbondisulfide.

Savolainen and Vainio (1977) reported the CS<sub>2</sub> binding to protein fraction of spinal neurofilaments of rat. So the binding of CS<sub>2</sub> to neurofilaments may contribute to the development of neural manifestations in chronic CS<sub>2</sub> poisoning.

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CHAPTER - 2

MATERIALS AND METHODS

Albino rats of either sex ( Haffkins strain ) weighing between 150-200 grms. were employed throughout this study. The selection of the animals was made at random basis. These animals were fed with pellet diet supplied by Hindustan Levers, Bombay, alongwith cereals and green vegetables. The access to water supply to animals was not restricted at any stage during this work.

GROUPING OF RATS :- For each of the experiments the animals were divided into two groups, i.e. the first group constituted carbondisulfide treated animals and other group served as control. Two types of carbondisulfide treatments were made in this work. 1. Acute 2. Chronic. In each of the treatments, six to seven animals formed a group.

ACUTE TREATMENT :- Five different groups of animals consisting of 6 to 7 animals in each group were taken for this treatment. The carbondisulfide ( AR-grade) in a dose of 1890 mg/kg, body weight was administered intraperitoneally in CS<sub>2</sub> treated group and the control group was injected with equivalent volume of vehicle. The physical observations on the gait, activity, and other bodily activities were made at an interval of 1,2,3 and 4 hours after CS<sub>2</sub> treatment. A group which was not sacrificed, died after five hours. Brain, liver, heart and blood was taken immediately and preserved in the deepfreeze for biochemical studies.

CHRONIC TREATMENT :- Rats were treated for four weeks, 6 days per week with carbondisulfide emulsion in groundnut oil 25 mg/kg intraperitoneally whereas control group was injected with equivalent volume of simple groundnut oil. Carbondisulfide emulsion was made by dissolving 1.0 ml of carbondisulfide in 100 ml of groundnut oil ( Postman Brand ). The animals were sacrificed by puncturing intrajugular vein and the blood was collected in an oxalated tubes, then the head was separated from the body. The brain, liver, heart , kidneys, pancreas, lungs and testis were taken in a dried and clean beaker to be preserved in a deepfreeze at  $-7^{\circ}\text{C}$  (  $\pm 2^{\circ}\text{C}$  ) till these were used for further processing for various biochemical parameters. The following behavioural observations and biochemical methods of analysis were done for each of the parameters.

### 2.1. BEHAVIOURAL OBSERVATIONS :-

Rats were trained in a jumping box for conditional avoidance response for two weeks. After  $\text{CS}_2$  treatment, conditional avoidance response was noted. In addition to this the eye movements, limb movements , gait activity and breathing movements were observed in acute as well as chronically treated rats.

### 2.2. GLYCOGEN ESTIMATION :-

Glycogen estimation in liver and brain was done by the method originally used by Pfluger (1905) and modified by Good, et.al.



(1933). The process depends on the digestion of tissues in hot concentrated KOH, precipitation of glycogen with acid, and determination of glucose in the hydrolysate as a reducing sugar.

Procedure :- About 0.5 to 1.0 Gram. of exactly weighed tissue was put in a 15.0 ml pyrex centrifuge tubes and reduced it to fine particles in a tissue homogeniser. After delivering the sample in a test tube, the tube was kept in a boiling water bath for 30 minutes to dissolve the tissue. 0.5 ml of saturated sodium sulfate was added and the glycogen was precipitated by the addition of 1.1 to 1.2 vol of 95% ethanol. The contents were stirred with a rod, and the rod washed with a small quantity of 60% ethanol. The tubes and contents were heated on a water bath again until the mixture was boiled, thereafter it was cooled and centrifuged at 3000 rpm. The supernatant was decanted and the aliquots were allowed to drain off. The precipitated glycogen was dissolved in water and the total volume was made up to 250 ml in a volumetric flask in each case. 0.5 ml of this solution and 4.5 ml of water was transferred to a set of tubes. In a separate tube, was introduced 5.0 ml of glucose standard containing 100 ug of glucose. To another tube was added 5.0 ml of water which served as a blank.

The tubes were submerged in the ice cold water. 10.0 ml of 0.2% anthrone reagent in (95%)  $H_2SO_4$  was added in each tube, and the reactants were mixed on vortex mixer. These cold tubes

were covered with glass marbles and heated for 10 minutes in a boiling water bath. They were then cooled immediately in a bath containing ice cold water and read on a spectrophotometer Spectronic-20 at 620 nm, after adjusting with the blank.

Calculations :-

Amount of glycogen in aliquot (ug)

$$= \frac{100 \times U}{1.11 \times S}$$

U = optical density ( O.D.) of unknown test solution.

S = O.D. of Standard glucose 100 ug/ml.

1.11 = It is the factor determined by Morris (1948), for the conversion of glucose to glycogen.

2.3. BLOOD GLUCOSE, ESTIMATION :-

Blood glucose determination depends upon the reducing nature of glucose, which reduces cupric ions to cuprous ions, that in turn reduce phosphomolybdic acid to phosphomolybdous acid of blue color.

Procedure :- Determination of blood glucose was done by the method of Folin & Wu (1926). Folin- Wu protein free filtrate was prepared by adding 9 volumes of the mixture of 8 parts N/12 H<sub>2</sub>SO<sub>4</sub> and one part 10% sodium tungstate directly to the blood. After mixing it was filtered. The clear filtrate was taken in Folin- Wu tube. Two ml of alkaline copper solution was added in all the tubes. The tubes were kept in a boiling

waterbath for six minutes. After cooling, 2.0 ml of phosphomolybdic acid solution was added to all the tubes. The contents were diluted up to 25.0 ml and optical density was read at 420 nm, on Spectronic- 20 . 2.0 ml of standard glucose solution ( 1 mg/ 1 ml of glucose ) and a blank containing 2.00 ml of distilled water were run alongwith the test samples.

#### Calculations :-

$$\frac{\text{OD of Test}}{\text{OD of Standard}} \times \text{mg of glucose in standard}$$

$$\times \frac{100}{2} = \text{mgs. of glucose per 100 ml of blood.}$$

#### 2.4. PROTEIN ESTIMATION :-

Procedure :- (i) Total plasma proteins : These were determined by the method of Lowry and Rosenberg (1951). 1.0 ml of plasma was diluted to 500 ml in a volumetric flask. 1.0 ml of diluted plasma was added in 5.0 ml of alkaline copper sulfate solution and allowed to stand for 10 minutes at room temperature. Similarly, in tubes marked standard, 1.0 ml of freshly prepared bovine serum albumin solution ( 100 ug/ml) was taken, and in blank 1.0 ml of distilled water was added. In both, standard and blank, 5.0 ml of alkaline copper sulfate solution was added and allowed to stand for 10 minutes. After this, 0.5 ml of folin cicalteau reagent was added in all tubes and mixed on a vortex mixer. All the set was allowed to stand for half an hour. The optical density was read at 740 nm on Spectronic-20



( using red lamp ).

(ii) For tissue proteins :- 10% homogenate of brain and liver was prepared in distilled water and further diluted 20 times and the above method described for plasma was followed.

Calculations :-

$$\frac{\text{Optical density of Test}}{\text{Optical density of standard}} \times \text{ug of proteins in standard}$$

$$\times \frac{\text{Dilution factor}}{\text{Vol of plasma or tissue extract}} = \text{ug of proteins/ ml. -}$$

of test solution

#### 2.5. SERUM -ALPHA- AMYLASE :-

Alpha amylase is an enzyme secreted by pancreas and salivary glands. It hydrolyses starch to disaccharides, maltose. Starch but not the maltose forms a blue colloidal complex with iodine in solution, and the intensity of this color is directly proportional to the concentration of the starch. The blue color produced by the starch substrate when combined with iodine, is measured after incubation with serum and compared with blank. The decrease in color is proportional to the amylase activity. Iodometric method by Carway (1959) depending on this principle was followed to determine the activity of serum alpha-amylase.

Reagents :-

- (i) Stable buffered starch substrate pH 7.0 :- 13.3 gm of anhydrous disodium phosphate and 4.3 gm of benzoic acid in about 250 ml of water was dissolved and was boiled. 0.20 gms. of Merck's solution starch was mixed in 5.0 ml of cold water and it was added to the boiling mixture. Boiling was continued for one minute. After cooling to room temperature, pH was adjusted to 7.0. Then the contents were diluted to 500.0 ml with distilled water.
- (ii) Stock solution of Iodine 0.1 N :- 3.56 gm Potassium iodate and 45 gm of potassium iodide was dissolved in 800 ml of water. To this was added slowly, with mixing 9.0 ml of concentrated HCl ( 12 N) and diluted to one liter with water.
- (iii) Working Iodine Solution :- ( 0.01 N ):-59.0 gm of sodium fluoride was dissolved in 350.0 ml of water. To this was added 50.0 ml of stock iodine solution and diluted to 500 ml mark in a volumetric flask.

Procedure :-

5.0 ml of starch substrate was pipetted in to 50 ml graduated tubes, marked test and blank. All the tubes were placed in water bath at 37°C for 5 minutes to warm up the contents. 0.1 ml of serum was pipetted in the bottom of the tubes labelled test. Allow the reaction to proceed for exactly

7½ minutes. No serum was added in the blank but instead 0.1 ml of distilled water was added. After 7½ minutes the tubes were removed from water bath and the volume was made up to 40.0 ml mark by addition of water. Immediately, 5.0 ml of working iodine solution to each of the tube was added and again the content was made up to 50.0 ml mark with water. Mixed well by shaking. Optical density was measured of tests and blank without delay against water at 660 nm on Spectronic- 20.

#### Calculations :-

$$\frac{\text{Optical density of Blank} - \text{Optical density of test}}{\text{Optical density of Blank}} \times 800$$

= amylase units 100 ml.

800 indicates that complete hydrolyses of starch would correspond to serum amylase activity of 800 units/100 ml. One amylase unit is the amount of enzyme that will hydrolyse 10.0 mg of starch in 30.0 minutes to a stage at which no color is given by iodine.

#### 2.6. PHOSPHORYLASE ASSAY :-

Phosphorylase activity was determined by measurement of the rate of liberation of inorganic phosphate from glucose-1-phosphate in presence of glycogen i.e. phosphorylase activity was measured in the direction of synthesis of polysaccharides by the method of Sutherland (1951).



### Procedure :-

The reaction was started by the addition of 1.0 ml of glucose-1-phosphate, glycogen and sodium fluoride to 0.5 ml of enzyme 5-AMP- NaF solution. The 0.5 ml of enzyme mixture contained 0.1 ml of 5-AMP ( 0.02 M), enzyme dilution ( Liver and brain, 10% homogenate ), and 0.1 M sodium fluoride to the volume of 0.5 ml. The assay mixture was incubated for 10 minutes at 37°C. The reaction was stopped by the addition of 10% trichloroacetic acid. In-organic phosphate was determined by the method of Fiske and Subba Row, as adopted for the Klett Summerson photometer at 660 nm.

Units and specific Activity :- One unit of enzyme was defined as that amount which caused the liberation of 1.0 mg of inorganic phosphate in 10 minutes when the percent conversion of glucose-1-phosphate was in the range of 12 to 22 percent.

### 2.7. PHOSPHOGLUCOMUTASE :-

The measure of phosphoglucomutase activity is based on the different chemical properties of two hexose phosphates. Glucose-1-phosphate is a nonreducing sugar with acid labile phosphate group. By contrast, glucose-6-phosphate has acid stable phosphate and reduces copper reagents.

For measuring enzyme activity, glucose-1-phosphate is used as the substrate because of the favourable equilibrium toward the forward reaction. The rate of reaction which is

dependent on the amount of enzyme present can be followed by measuring the reduction in hydrolysable phosphate. This principle was employed by Victor et.al. (1954).

Procedure :- 0.1 ml of  $Mg SO_4$  ( $6 \times 10^{-2}$  M), 0.1 ml glucose-1-phosphate ( $2 \times 10^{-2}$  M, pH 7.5) and 0.1 ml of cysteine (0.1 M, pH 7.5), were pipetted in a test tube and placed in a water bath at  $30^{\circ}C$ . After temperature equilibration, the reaction was started by adding 0.1 ml of the enzyme dilution. After incubation for 5 minutes, the reaction was stopped by the addition of 1.0 ml of 5 N  $H_2SO_4$ . The volume was made up to 5.0 ml with water, and the reaction tubes were placed in a boiling water bath for 3 minutes to hydrolyse the remaining glucose-1-phosphate. To the sample 1.0 ml of molybdate solution followed by 0.5 ml of 1-amino-2-naphthol-4-sulfonic acid (ANSA) solution were added according to Fiske & Subba Row (1925). The volume was made up to 10.0 ml, and the intensity of color produced was measured at 660 nm in Klett Summerson photometer. The amount of stable phosphate formed is the difference between the hydrolysable phosphate added and that remaining at the end of 5 minutes.

Unit Activity :- One unit of enzyme is defined as that which, under the conditions described above, catalyses the formation of 1.0  $\mu g$ , acid stable phosphate.

## 2.8. FRUCTOSE 1-6 DIPHOSPHATASE :-

The method is that of Pogell and McGilvery (1955) in

which the inorganic phosphate liberated from Fructose 1-6-Diphosphate ( FDP) is determined.

Procedure :- 0.1 ml of FDP ( 0.05 M, pH 7.4), 0.4 ml of borate buffer ( pH 9.5), 0.1 ml of Mg SO<sub>4</sub> ( .05 M ), and 0.1 ml of MnCl<sub>2</sub> ( .005 M ), were pipetted in centrifugation tubes. After warming 0.2 ml of enzyme was added to the tube, followed immediately by 0.1 ml of cysteine ( .05 M, pH 9.5 ).

Then this mixture was incubated at 38°C for twenty minutes and to it was added 1.0 ml of trichloroacetic acid and inorganic phosphate was determined by the method of Fiske and Subba Row (1925).

Unit Activity :- One unit of enzyme is defined as the amount which will liberate 1 mg of inorganic phosphate with linear kinetics of 20 minutes under the above conditions.

## 2.9. ADENOSINE TRIPHOSPHATASE :-

The assay of adenosine triphosphatase is most simply carried out by estimating colorimetrically the inorganic phosphate liberated by the enzyme from ATP under the specified conditions as stated by ( Seth et.al. 1966).

Procedure :- 0.2 ml of Tris buffer ( 0.5 M, pH 8.0), 1.5 ml of water, 0.1 ml of 10% tissue homogenate, 0.2 ml of ATP (55mg/10 ml) was pipetted in the centrifugation tube. The assay mixture was incubated at 37°C for 15 minutes. The reaction



The reaction was stopped by adding 1.0 ml of 10% trichloroacetic acid. The mixture was centrifuged and to 1.5 ml of supernatant 1.0 ml ammonium molybdate ( 2.5 % in 10 N  $H_2SO_4$ ) 10 N  $H_2SO_4$  was added followed by the addition of 7.1 ml of water and 0.4 ml of ANSA and optical density was noted at 660 nm.

One unit of enzyme is that which liberates 1.0 mg of inorganic phosphates in 15 minutes.

#### 2.10. SUCCINIC DEHYDROGENASE :-

The activity of Succinic Dehydrogenase ( SDH) was determined by the method of Kun and Abowd ( 1949), by its ability to oxidize succinic acid and reducing equivalents liberated in turn reduce the 2,4,6 triphenyl tetrazolium chloride, (TTC).

Procedure :- To a 15.0 ml caliberated centrifuge tubes, 0.5 ml of 0.1 M phosphate buffer ( pH 6.7) and 1.0 ml of 10% tissue homogenate were pipetted. Finally, 1.0 ml of freshly prepared 0.1% TTC solution was poured in. After shaking, the tubes were placed in a water bath at  $38^{\circ}C$  for a period of exactly 30 minutes. Immediately, after removal of tubes from water bath, 3.0 ml of acetone was added and the tubes were stoppered and shaken vigorously. Then tubes were centrifuged and the clear supernatant was read at 420 nm on spectronic-20. Sodium - dithionite (1% ) is used as standard reducing substance and mili-gram of TTC dye reduced is calculated.

## 2.11. BIOGENIC AMINES :-

The monoamines, dopamine ( DA), norepinephrine ( NE) and 5-hydroxytryptamine ( 5-HT) were estimated on Aminco-Bowman spectrofluorometer. DA and NE were extracted and oxidized by the method of Chang et.al.(1964) and 5-HT was extracted and estimated by the method of Curzon and Curzon (1970).

### Reagents :-

- (i) Acidified butanol :- Analytical grade of n-butanol was prepared by adding 0.85 ml of concentrated HCl per liter of n-butanol.
- (ii) 0.1 M EDTA :- 37.2 G of disodium ethylene diamine tetraacetate ( EDTA) was dissolved in 1 M sodium acetate solution and diluted to one liter. The pH of the resultant solution was adjusted between 6.7 to 7.0.
- (iii) Alcoholic Iodine Solution :- 1.27 G iodine dissolved in 100 ml of absolute alcohol.
- (iv) Alkaline sulfite reagent :- 1 ml of 25%  $\text{Na}_2\text{SO}_3$  ( anhydrous) solution diluted with 9.0 ml of 5 N NaOH. This solution was always made fresh immediately before use.
- (v) Standard solutions :- 100 ug/ml. solutions of NE, DA and 5-HT in 0.01 N HCl were kept at low temperature in a refrigerator. The stock standard was diluted when needed, with .02 M acetic acid to a concentration of 5 ug/ml. for DA and NE and 1 ug/ml for 5-HT.

Extraction of DA and NE :-

Whole rat brain, a weighed portion of liver, whole heart, a weighed portion of lungs were homogenised separately in acidified butanol using glass homogenising tubes and a motor driven teflon pestle. 10% solution of homogenates were prepared and were centrifuged for 10-15 minutes at 4000 rpm at  $-7^{\circ}\text{C}$  ( $\pm 2^{\circ}\text{C}$ ).

To 5.0 ml of supernatant, was added 10.0 ml of n-heptane (AR Grade) and 5 ml of water. These were shaken for 2 minutes on vortex mixer. 5.0 ml of aqueous phase was taken, to that was added to 0.2 gm alumina. Adjusted the pH between 7 to 7.5 by adding 1.0 ml of 2 M sodium acetate to each tube. Again the tubes were shaken for 5 minutes and centrifuged. DA and NE were adsorbed on activated alumina. The alumina bound catecholamines were eluted by shaking with 3.0 ml of 0.1 M acetic acid. The pH of eluate was always adjusted around 4.0 to 4.3.

1.0 ml of eluate was used for oxidation of DA and NE. To a 0.1 ml of eluate, 0.2 ml of EDTA reagent was added and the mixture was adjusted to a pH of 6.5. To the resultant solution, 0.1 ml of 0.1 N alcoholic iodine solution was added to oxidise catecholamines (CA). Exactly after two minutes, 0.2 ml of alkaline sulfite solution was added to stop the reaction. Again after 20 minutes, pH of the solution was adjusted to about 5.4 by addition of 0.2 ml of 5 N acetic acid.



### Tissue Blank :-

To 1.0 ml of acidified eluate 0.2 ml of EDTA reagent was added and the mixture was adjusted to a pH of 6.5. The iodine solution and alkaline sulfite reagent were added in a reverse manner in contrast to standard i.e. first 0.2 ml alkaline sulfite reagent was added followed by 0.1 ml of iodine solution. The solutions of standard catecholamines were prepared by taking 0.1 to 2 ug of standard solution, oxidized as described above.

Internal standard solutions equivalent from 20 to 200 ng of NE and DA were added to the tissue supernatant and were carried through extraction and oxidation procedures as described above. Therecovery of exogenous amines added to the tubes prior to the first shaking step was 90 to 95% for 5-HT and 75- 80% of NE and DA. These solutions were heated in a boiling water bath for 30 minutes, cooled and read in aminco bowman's spectrofluorometer as follows :-

For NE- excitation of the solution at 395 mu and emission read at 485 mu. Band pass was kept at 2, slit width at 2. For DA, excitation of the solution at 345 mu and emission read at 410 mu. Band Pass was always kept at 2 with slit width at 2.

### 2.12. EXTRACTION OF 5-HYDROXYTRYPTAMINE :-

10% homogenate prepared in acidified butanol as prepared

for CA extraction, was taken. To 2.5 ml of supernatant was added 5.0 ml of n-heptane, 0.4 ml of 0.1% cysteine in 0.1 N HCl. The contents were mixed thoroughly and centrifuged. To the aqueous phase 0.6 ml of .004 % ortho pthalaldehyde, prepared in 10 N HCl, was added. The contents were kept in boiling water bath for 15 minutes and then cooled in water. The fluorescence was read on Aminco-bowman Spectrofluorometer by activation wavelength at 295 nm and emission wavelength at 470 nm. Band Pass was kept at 2, and slit width at 4. ( Curzon and Curzon 1970 ).

### 2.13. MONOAMINE OXIDASE :-

The activity of monoamine oxidase ( MAO) was determined by its action on benzylamine hydrochloride which resulted in the formation of benzaldehyde and the extinction of benzaldehyde at 250 nm was used to determine the activity of this enzyme ( Tabor et.al. 1955 ).

#### Procedure :

0.4 ml of phosphate buffer ( 0.5 M, pH 7.2 ) was pipetted into a centrifugation tube followed by addition of 1.3 ml of water. 0.2 ml of 10% homogenate of tissues were poured, followed by addition of 0.1 ml of benzylamine-hydrochloride ( 0.1 M). This assay mixture was incubated for 30 minutes at 37°C. The reaction was stopped by addition of 1.0 ml of 10% perchloric acid. 3.0 ml of water was added, then centrifuged and read at 250 nm on U.V. Spectrophotometer, (Unicam SP-500). Extinction

of benzaldehyde at 250 nm is 11,300.

#### 2.14. GLUTATHIONE- REDUCED : (G-SH ) :-

The method of Ellman (1959) was used to assay sulphhydryl groups of reduced glutathione.

##### Procedure :-

To 10% homogenate was added equal volume of 10% trichloroacetic acid (TCA) and then centrifuged. To 0.1 ml of supernatant 0.4 ml of 5-5' dithiobis 2-nitrobenzoic acid ( DTNB) was added which was prepared as 5 mg per 100 ml of 0.5 M phosphate buffer ( pH 8.0). The optical density was taken at 412 nm.

#### 2.15. ANALYSIS OF ZINC, COPPER AND MANGANESE :-

Analysis of these trace elements in various organs of rat in CS<sub>2</sub> treated and in control was done by atomic absorption spectrophotometer ( Perkin-Elmer ) Model- 303.

##### Sample Preparation :-

Sample preparation was done by wet ashing method. Placed about 0.5 gm ( accurately weighed ) of the tissues in a 25 ml erlenmeyer flask, having glass beads and 2.5 ml of deionised water. Added 1 ml of a mixture (1:1) of concentrated HNO<sub>3</sub> and HClO<sub>4</sub>. The sample was boiled until the solution was clear. The final volume of the mixture was made to 10 ml with deionized water. This extract was used for analysis of zinc, copper and manganese on specified conditions and settings of



the instrument for each element, are described below.

Standard Conditions for Zinc :-

Stock Standard Solution :- Zinc 500 ug/ml. 0.5 gm of Zinc metal was dissolved in minimum volume of (1+1) HCl, and diluted to one liter with 1% ( v/v) HCl.

Operating Parameters :-

Wavelength was set at 214 nm, slit was kept at 4 ( 0.7 nm). Light source was Hollow cathode lamp. Air - acetylene oxidising flame ( lean and blue ) was used.

Linear Working Range :-

Zinc was linear upto concentrations of approximately 1 ug/ml in aqueous solution. A standard zinc solution containing 0.5 ug/ml. showed an absorbance reading of about 0.12 absorbance i.e. about 25% absorption.

Standard Conditions for Copper :-

Stock Standard Solution :- ( Copper, 1000 ug/ml) Dissolved 1.0 gm of copper metal in a minimum volume of ( 1+1)  $\text{HNO}_3$  and diluted to one liter with 1% ( v/v)  $\text{HNO}_3$ .

Operating Parameters :-

Wavelength was set at 325 nm and slit was set at 4 ( 0.7 nm). Light source used for estimation was Hollow cathode lamp and air acetylene flame when employed showed a linear working

range upto concentrations of approximately 5 ug/ml in aqueous solution.

Standard Conditions for Manganese :-

Stock Standard solution :- (Manganese 1000 ug/ml) Dissolved 1.0 gm of manganese metal in a minimum volume of ( 1+1)  $\text{HNO}_3$  and diluted to one liter with 1% (v/v) HCl.

Operating Parameters :-

Wavelength was set at 279 nm. Slit was set at 3(0.2 nm). Light source was Hollow cathode lamp and air acetylene oxidizing flame was used.

The working range for manganese was linear upto concentrations of approximately 3 ug/ml in aqueous solution. A standard containing 2 ug/ml manganese was specified, as it gave an absorbance reading of about 0.16 absorbance units i.e. about 31% absorption.

\*\*\*\*\*

CHAPTER - 3.

RESULTS AND OBSERVATIONS



### 3.1. BEHAVIOURAL OBSERVATIONS :-

A. Acute Treatment :- The animals exhibited an intense stimulation immediately after the administration of  $CS_2$ . There was itching at the site of injection and the spontaneous activity was reduced very much, afterward the animal became sedated and the eyelid size became slightly less. The animals acquired mild to intense rigidity and became placid and akinesic which increase in intensity and ultimately passed on to comma and death within 5 hours of administration of  $CS_2$ .

B. Chronic Treatment :- The animal showed a slight stimulation immediately after the injection of  $CS_2$ . But after some time the animals became normal. After a few weeks of treatment there body hair became rough and eye pupil dilated. There was no significant changes in physical observation in these rats.

### 3.2. EFFECT OF SINGLE DOSE OF CARBONDISULFIDE ( ACUTE TREATMENT) IN RAT ON SOME BIOCHEMICAL PARAMETERS :-

The following changes in various parameters were noticed after an acute dose of carbondisulfide.

#### 3.2.1. Tissue Glycogen level :-

There was an appreciable fall in the liver glycogen content as after one hour of the injection the liver glycogen

decreased by 30.65%, at two hours it was decreased by 83.80% at three hour it was decreased by 89.43% and after four hours glycogen was almost completely depleted from the liver, (97.31%) (Table-1; Fig.1 ). CS<sub>2</sub> treated rats showed an initial rise in the brain glycogen content, as after the first and second hour of the injection the glycogen content was increased by 8.33% and 52.38% respectively, whereas brain glycogen decreased by 12.0% and 17.36% in third and fourth hour after the injection of CS<sub>2</sub> ( Table-2; Fig.2).

### 3.2.2. Tissue Protein Level :-

A significant reduction in the total proteins of liver was found in CS<sub>2</sub> treated group of rat when compared to control group ( untreated ). The liver total proteins at various intervals after the CS<sub>2</sub> injection have been shown in Table-3; Fig.3. Liver proteins continuously reduced after 1,2,3 and 4 hour of CS<sub>2</sub> injection by 19.22%, 45.40%, 54.9% and 70.0% respectively (Table-3; Fig.3). In Table 4, Fig.4, the effect of CS<sub>2</sub> on brain protein level at various intervals is shown. Total brain proteins pattern varies quite in contrast to that of liver proteins. There is an initial rise in brain total proteins, in first hour after CS<sub>2</sub> injection, ( 56.5%). After 2,3 and 4 hours the brain proteins increased but not significantly as the rise in comparison to controls was only 14.3%, 11.5% and 2.7% respectively. One can say that brain proteins increased in first hour and then decreased in 2,3 and 4th hour gradually.!

### 3.2.3. Plasma Proteins, Blood Glucose and Serum Alpha-Amylase :-

A marked effect of acute carbondisulfide treatment on plasma proteins was observed because a significant decrease in plasma proteins have been found in first, 2nd, 3rd and 4th hour after the injection of CS<sub>2</sub>. (Table-5, Fig.5) shows the decrease of plasma proteins level in CS<sub>2</sub> treated rat by 19.13%, 41.4%, 48.84% and 65.89% respectively in comparison to control group. Blood glucose is also severely affected by CS<sub>2</sub> intoxication. First hour after the CS<sub>2</sub> injection blood glucose increased highly and rose from 140.5 mg % to 234.37 mg % . But from second hour to fourth hour, glucose level reduced gradually by 28.8%, 49.02% and 60.24% in comparison to control values ( Table-6; Fig.6).

In Table-7 Fig.7, the effect of CS<sub>2</sub> on serum alpha-amylase activity has been given. In treated rat the serum alpha amylase activity was found to be increased very appreciably as compared to untreated rat. In first hour after treatment, there was 86.40% rise, in second 250.35%, in third 205.94% and in fourth hour the rise was 246.05%. So in the first two hours there was a steep rise in serum alpha-amylase activity after which it did not rise but was similar to that of second hour level.

The changes in the weight of the organs after one month treatment with CS<sub>2</sub> is also represented in Table-8; Fig.8. The liver weight has increased significantly without any significant change in the weight of other organs.



### 3.3. EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ON BIOCHEMICAL PARAMETERS :-

#### 3.3.1. Tissue Glycogen :-

After one month treatment with CS<sub>2</sub>, the glycogen level of liver was 74.88% higher than the control group. Similar was the case of glycogen level in rat brain which was 52.2% higher (Table-9; Fig.9).

#### 3.3.2. Tissue Proteins :-

Table 10; Fig.10, shows the effect of chronic treatment on protein level of liver and brain. In liver a fall of protein level was observed by 15.93%, but brain protein level increased by 55.39% when compared to control group. So one can assume that after CS<sub>2</sub> treatment protein accumulate in brain to a greater extent.

#### 3.3.3. Plasma proteins, blood glucose and serum alpha-amylase activity :-

Plasma protein level decreased in CS<sub>2</sub> treated rat by 41.5% whereas blood glucose level increased in treated group by 44.19% as compared to untreated group. Serum alpha-amylase activity was similarly found to be higher in treated group by 49.3% as compared to normal untreated rat ( Table-11; Fig. 11).

### 3.3.4. Effect of Chronic Carbondisulfide Treatment on some Enzymes :-

The results of various estimations of enzymes, related to carbohydrate metabolism, are described below :

Enzymes activity was estimated both in liver and brain which have been found to be the main metabolic organs in this study.

3.3.4.1. Phosphorylase :- The liver phosphorylase activity was decreased by 58.43% in treated group as compared to untreated group, whereas brain phosphorylase is not effected significantly which decreased only by 9.99% (Table-12; Fig.12).

3.3.4.2. Phosphoglucomutase :- Phosphoglucomutase is an important enzyme of glycogen biosynthesis and its alteration observed due to CS<sub>2</sub> has been represented in Table-13; Fig.12. Liver phosphoglucomutase activity increased steeply by 248.59% in the CS<sub>2</sub> treated group in comparison to control group. Brain phosphoglucomutase also showed an upward trend as it increased by 34.61% as compared to untreated group. (Table-13; Fig.12).

3.3.4.2. Fructose 1-6, Diphosphatase :- Fructose- 1-6 diphosphatase ( FDPase ) an enzyme of importance in gluconeogenesis was also observed to be altered by carbondisulfide treatment for one month, as shown in Table -14; Fig.12. Liver FDPase activity is decreased by 42.02% and brain FDPase activity is decreased by 21.60% when compared to untreated group. The changes are statistically significant (  $P < .05$  ,  $< .025$  )

### 3.3.4.4. Adenosine triphosphatase ( ATPase ) :-

Adenosinetriphosphatase is significantly effected by chronic CS<sub>2</sub> toxicity. Table-15; Fig.13, shows the ATPase alteration in different tissues. Brain ATPase activity was found to be decreased by 24.73% while liver ATPase activity decreased by 48.24%.

### 3.3.4.5. Succinic Dehydrogenase :-

Succinic dehydrogenase, an important enzyme of Kreb's cycle, was also found to be altered by carbondisulfide toxicity as shown in Table-16; Fig.13. Liver succinic dehydrogenase decreased by 36.80% as compared to control and the brain it decreased by 28.54% when compared with control group. The decrease is significant in both the organs.

### 3.3.5. EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ON BIOGENIC AMINES, MONOAMINE OXIDASE AND GLUTATHIONE ( G.S.H. ) :-

Chronic treatment of carbondisulfide showed a severe alterations in the metabolism of neurotransmitters in various tissues. Following are the variations observed after CS<sub>2</sub> treatment.

3.3.5.1. Dopamine :- Dihydroxyphenylethylamine ( Dopamine) level is changed in all the tissues studied as shown in Table-17; Fig.14. Brain, heart and liver of treated group



showed a rise of dopamine by 161.24%, 412.65% and 262.94% respectively, where as the concentration of dopamine is decreased by 71.89% in lungs. So dopamine metabolism is effected variiently but sharply by CS<sub>2</sub> toxicity.†

### 3.3.5.2. Norepinephrine ( NE ) :-

Norepinephrine, an important neurotransmitter and an important metabolic regulatory hormone is effected by carbondisulfide toxicity. In brain a rise of 86.37% in heart a rise of 138.2% in liver a rise of 56.69% and in lungs a rise of 78.57% of norepinephrine content in treated group was observed as compared to nontreated group. (Table-18;Fig.14,16) These changes in NE tissue concentration may be responsible for many metabolic alterations in the living system.†

### 3.3.5.3. Serotonin ( 5-Hydroxy Tryptamine ):-

Serotonin, a metabolite of tryptophan metabolism and a neurotransmitter, altered by carbondisulfide toxicity as shown in Table-19;Fig.14,16. There was a significant decrease in brain serotonin by 45.88%. But 5-HT level of heart, liver and lungs increased by 41.95%, 240.0% and 367.1% respectively.†

### 3.3.5.4. Monoamineoxidase :-

Monoamine-oxidase is an enzyme which is responsible for the metabolism of neurotransmitters.† Carbondisulfide shows

a different effect in different organs on monoamine oxidase activity. There was a marked fall of MAO activity in brain, which was observed to be 42.57% lesser than control value. However, MAO in lungs decreased by 4.21% which is not a significant change. However, a marked rise of 47.18%, 71.25% was observed in heart and liver MAO activity (Table-20; Fig. 17).

#### 3.3.5.5. Reduced Glutathione ( G-SH) :-

Glutathione is an important molecule responsible for the covalent binding of various toxicants. Carbondisulfide was found to effect the level of glutathione ( G-SH) appreciably in various organs ( Table-21; Fig. 18). Glutathione was observed to be decreased in all organs studied in treated group as in brain there was a decrease of 17.91%, in heart, 90.87%, in liver 14.30% and in lungs 86.18 %.

#### 3.3.6. EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ON TISSUES ZINC, MANGANESE AND COPPER LEVEL.

Carbondisulfide toxicity is found to alter the various trace elements in the living system. Following is the observed changes in Zinc, Manganese and Copper levels in various tissues i.e., liver pancreas, kidneys, testis and brain.

##### 3.3.6.1. Zinc :-

Zinc concentration was observed to be altered by CS<sub>2</sub>

toxicity in various tissues as shown in Table-22;Fig.19. Zinc concentration in liver was decreased by 26.43%, in pancreas by 33.73% in testis by 34.87%, in brain 17.77% where as it is not significantly changed in Kidney ( 2.97% decrease only ).

### 3.3.6.2. Manganese :-

Manganese distribution was also altered by CS<sub>2</sub> treatment in various tissues as shown in Table-23;Fig.20. Manganese concentration was increased in liver by 34.81% in pancreas by 150.38% and in Kidneys by 18.42%. But in testis the concentration of manganese was decreased by 36.90% which was significant, ( P=0.005 ). Change in brain was N.S.

### 3.3.6.3. Copper :-

Copper concentration of liver is increased a little i.e. about 14.5% by CS<sub>2</sub> treatment whereas copper concentration in pancreas was altered markedly i.e. 157.83% more than the control group ( Table-24 ; Fig. 21). Kidney, Testis and Brain Copper concentration was decreased by 30.32%, 59.79% and 27.22% respectively.



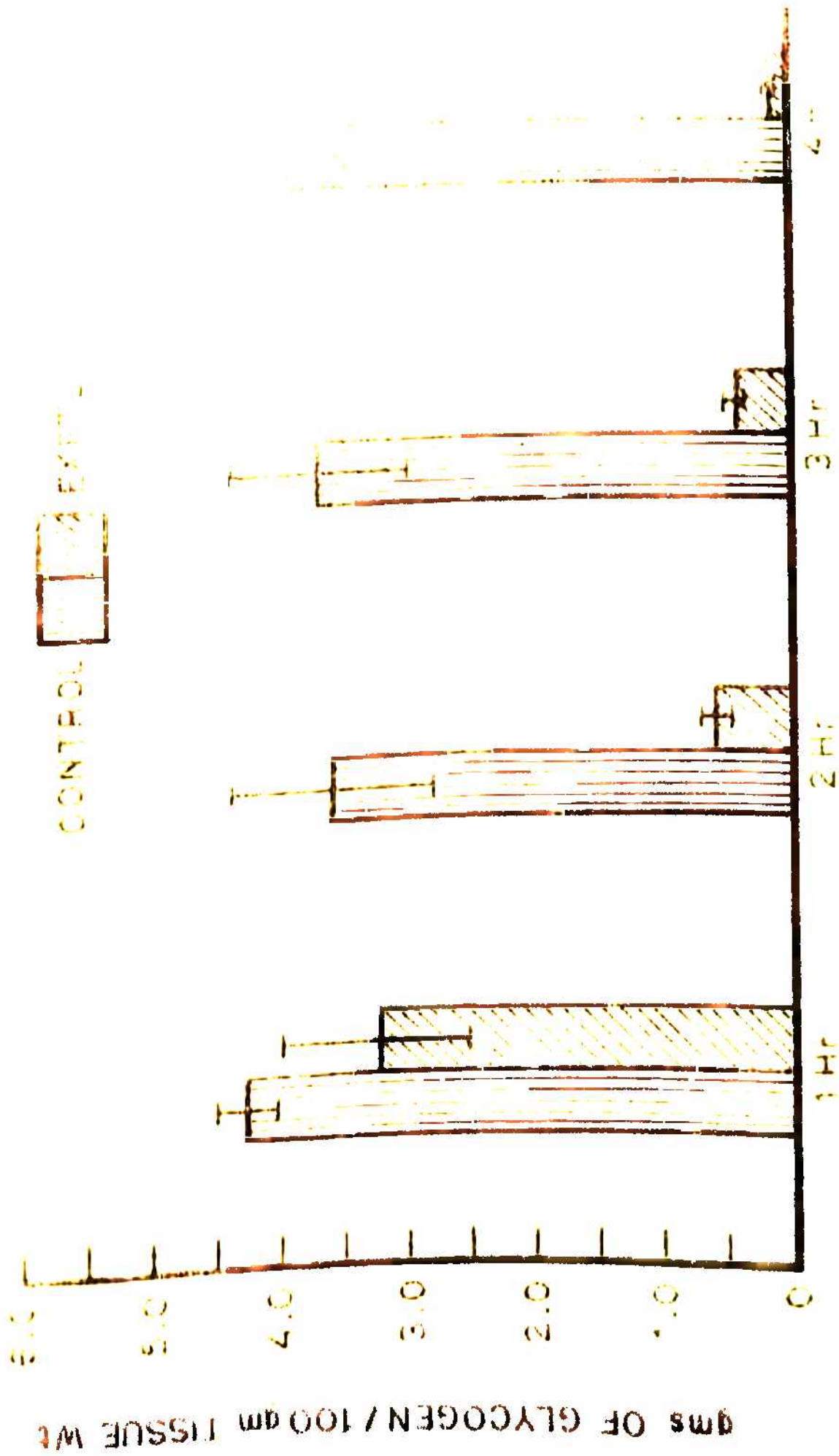
TABLE : 1

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1890 mg/kg body weight)  
ON GLYCOGEN LEVEL ( G/100 G tissue weight ) OF LIVER.

Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
1	5	4.37	3.28	-30.65	< .05
		± 0.21	± 0.70		
2	5	3.52	0.57	-83.80	< .005
		± .87	± .10		
3	5	3.58	0.41	-89.43	< .005
		± .67	± .07		
4	5	3.93	.107	-97.50	< .005
		± .58	± .056		

+ Shows a n increase , - Shows a decrease.

All experimental values are , Mean ± Standard error (S.E.)



LIVER GLYCOGEN

FIG: 1

TABLE : 3

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1890 mg/kg body weight )  
ON GLYCOGEN LEVEL ( G/100 G tissue weight ) OF BRAIN.

Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level
1	5	0.24 ± .04	0.26 ± .04	+ 8.33	N.S.
2	5	0.21 ± .06	.32 ± .07	+ 52.38	N.S.
3	5	0.25 ± .047	0.22 ± .03	- 12.0	N.S.
4	5	0.19 ± .02	.157 ± .02	- 17.36	N.S.

+ Shows an increase, - Shows a decrease.



FIGURE: 2

SHOWS THE EFFECT OF ACUTE DOSE OF CARBONDISULFIDE  
ON GLYCOGEN LEVEL OF BRAIN AFTER 1 HOUR, 2 HOUR, 3  
HOUR AND 4 HOUR OF INTRAPERITONEAL INJECTION.

TABLE - 3

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1890 mg/kg body weight )  
ON TOTAL PROTEIN LEVEL ( mg/G Tissue weight ) OF LIVER.

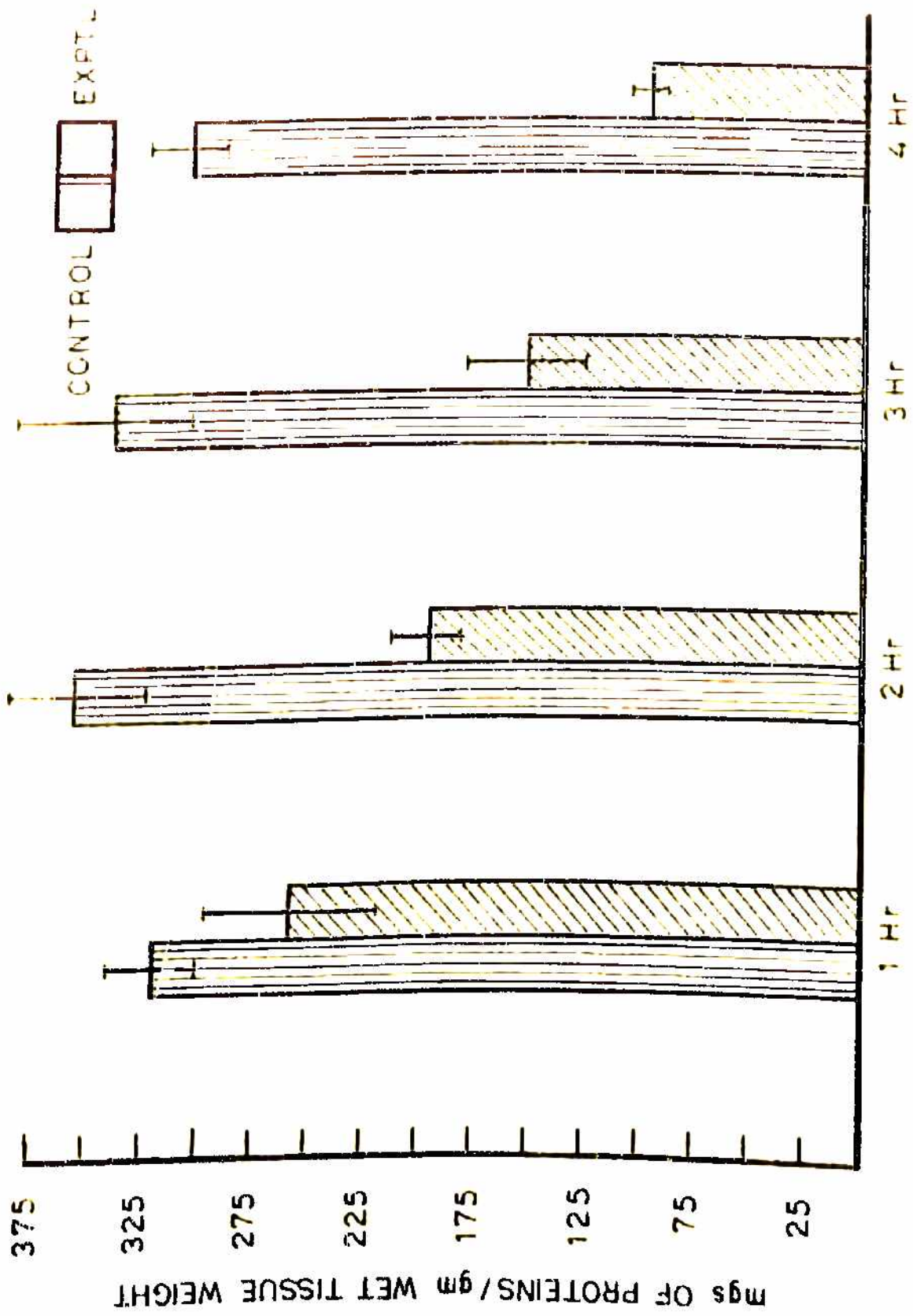
Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.																				
1	5	320.40	259.24	-19.22	<.05																				
		± 20.56	± 39.88			2	5	356.42	194.66	-45.10	<.005	± 32.16	± 16.96	3	5	338.90	152.73	-54.9	<.005	± 44.84	± 26.06	4	5	309.24	92.48
2	5	356.42	194.66	-45.10	<.005																				
		± 32.16	± 16.96			3	5	338.90	152.73	-54.9	<.005	± 44.84	± 26.06	4	5	309.24	92.48	-70.00	<.005	± 10.06	± 8.45				
3	5	338.90	152.73	-54.9	<.005																				
		± 44.84	± 26.06			4	5	309.24	92.48	-70.00	<.005	± 10.06	± 8.45												
4	5	309.24	92.48	-70.00	<.005																				
		± 10.06	± 8.45																						

+ Shows an increase , - Shows a decrease.

FIGURE: 3

SHOWS THE EFFECT OF ACUTE DOSE OF CARBONDISULFIDE  
ON TOTAL PROTEIN LEVEL OF LIVER AFTER 1 HOUR, 2  
HOUR, 3 HOUR AND 4 HOUR OF INTRAPERITONEAL  
INJECTION.





TOTAL PROTEINS IN LIVER

**FIG: 3**

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1890 mg/kg body weight )  
ON TOTAL PROTEIN LEVEL ( mg/G Tissue weight ) OF BRAIN.

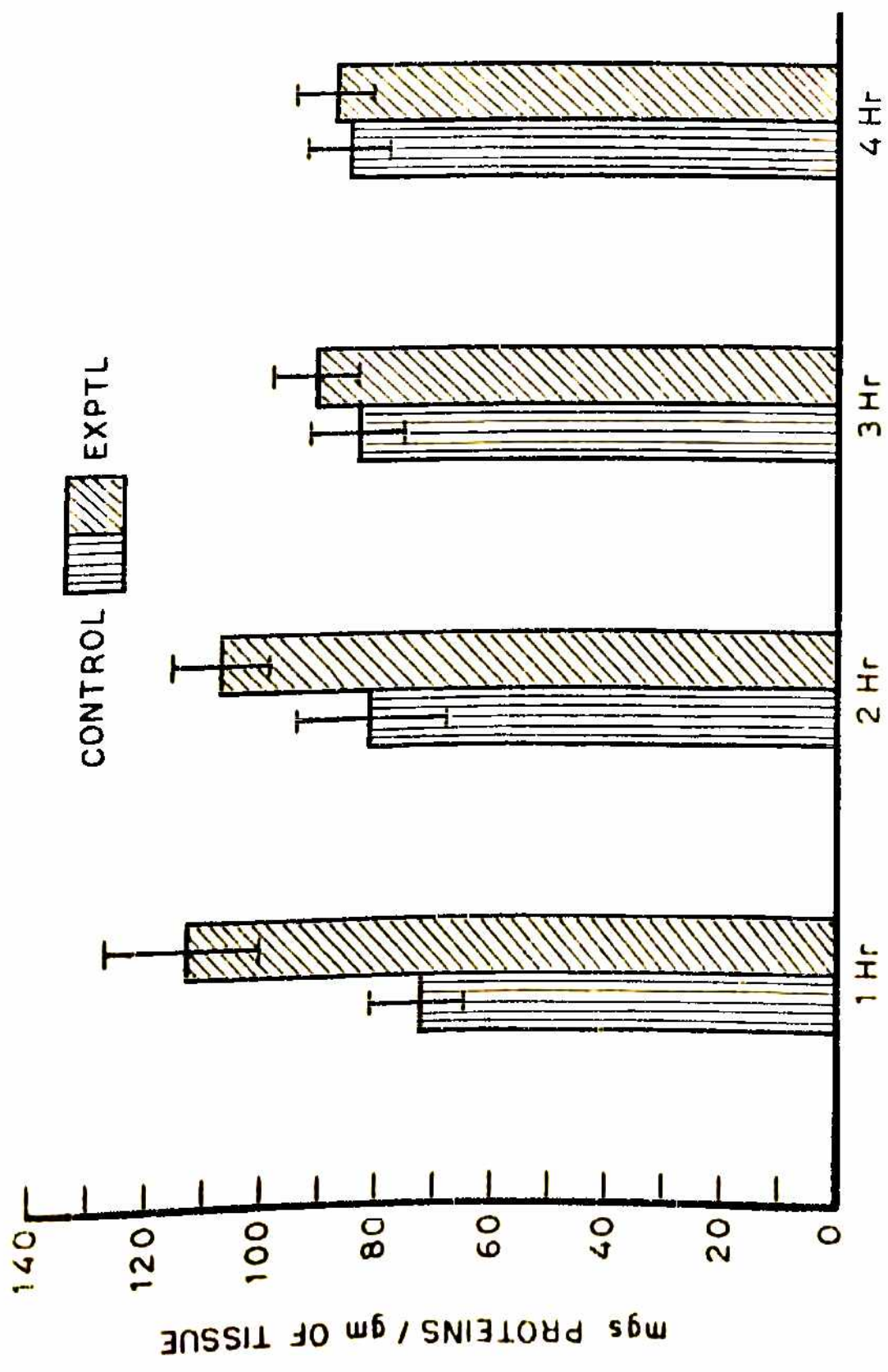
Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level,
1	5	72.70	113.81	+ 56.5	L.01
		± 8.60	± 13.11		
2	5	85.38	107.74	+ 14.3	N.S.
		± 14.60	± 8.02		
3	5	82.42	91.94	+ 11.5	N.S.
		± 8.83	± 7.15		
4	5	86.10	88.49	+ 2.7	N.S.
		± 7.32	± 6.01		

+ Shows an increase , - Shows a decrease.

FIGURE: 4

SHOWS THE EFFECT OF ACUTE DOSE OF CARBONDISULFIDE  
ON TOTAL PROTEIN LEVEL OF BRAIN AFTER 1 HOUR, 2  
HOUR, 3 HOUR AND 4 HOUR OF INTRAPERITONEAL  
INJECTION.





TOTAL PROTEINS IN BRAIN

**FIG: 4**

TABLE : 5

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1890 mg/kg body weight )  
ON TOTAL PLASMA PROTEIN LEVEL ( G/100 ml ) .

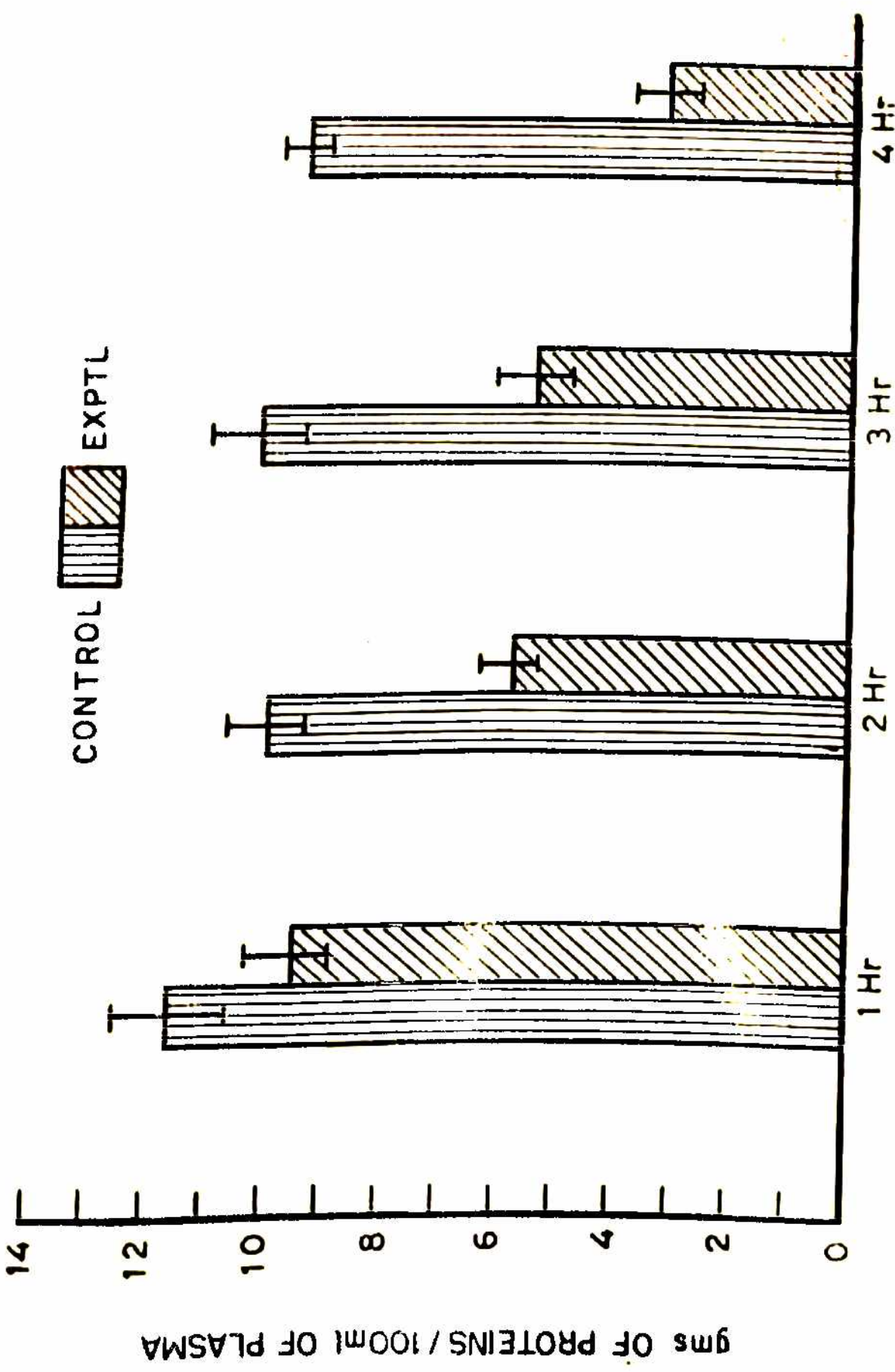
Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
1	5	11.76	9.51	-19.13	<.025
		± .92	± .74		
2	5	10.00	5.86	-41.4	<.005
		± .61	± .55		
3	5	10.40	5.32	-48.84	<.005
		± .84	± .69		
4	5	9.56	3.26	-65.89	<.005
		± .38	± .53		

+ Shows an increase, - Shows a decrease.

FIGURE: 5

SHOWS THE EFFECT OF ACUTE DOSE OF CARBONDISULFIDE  
ON TOTAL PLASMA PROTEIN LEVEL AFTER 1 HOUR, 2  
HOUR, 3 HOUR AND 4 HOUR OF INTRAPERITONEAL  
INJECTION.





TOTAL PLASMA PROTEINS

FIG: 5

TABLE : 6

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1500 mg/kg body weight )  
ON BLOOD GLUCOSE LEVEL (mg/100 ml).

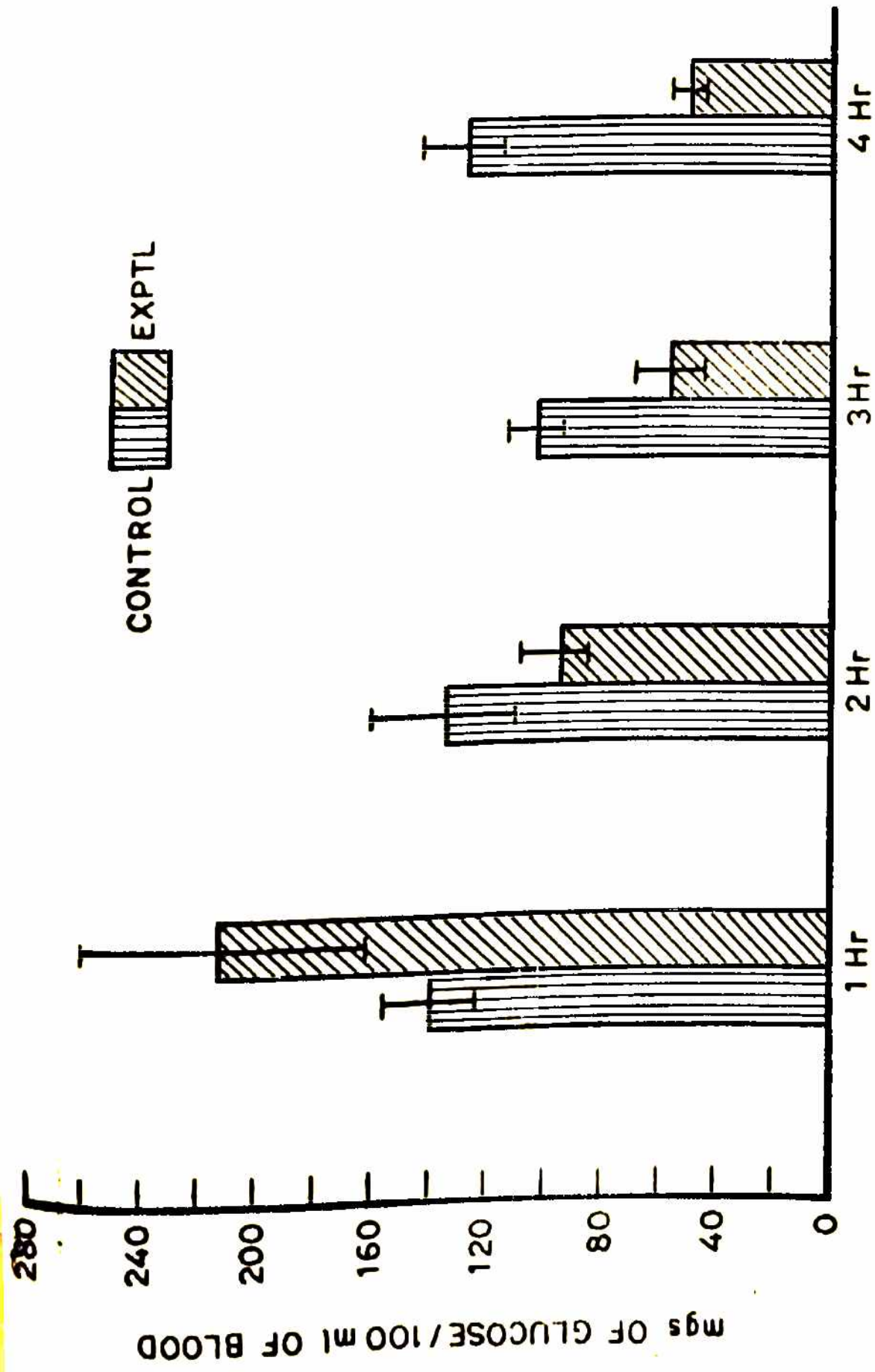
Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level,
1	5	140.5 ± 15.5	234.57 ± 45.83	+ 66.0	<.025
2	5	135.5 ± 25.7	96.42 ± 12.57	- 28.84	<.01
3	5	115.30 ± 9.50	58.78 ± 13.4	- 49.02	<.005
4	5	129.1 ± 14.4	51.58 ± 6.66	- 60.24	<.005

+ Shows an increase , - Shows a decrease.

FIGURE: 6

SHOWS THE EFFECT OF ACUTE DOSE OF CARBONDISULFIDE  
ON BLOOD GLUCOSE LEVEL AFTER 1 HOUR, 2 HOUR, 3  
HOUR AND 4 HOUR OF INTRAPERITONEAL INJECTION.





BLOOD GLUCOSE

FIG : 6

TABLE : 7

EFFECT OF SINGLE ACUTE DOSE OF CARBONDISULFIDE ( 1800 mg/kg body weight )  
ON SERUM ALPHA AMYLASE ACTIVITY. (\* Units/100 ml.)

Hours After Injection	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
1	5	295.6	551.00	+ 86.40	.1
		± 43.20	± 184.61		
2	5	274.5	961.72	+250.35	.005
		± 38.30	± 133.51		
3	5	354.7	1085.20	+ 205.04	.005
		± 62.5	± 72.30		
4	5	393.4	1361.4	+ 246.05	.005
		± 74.3	± 245.08		

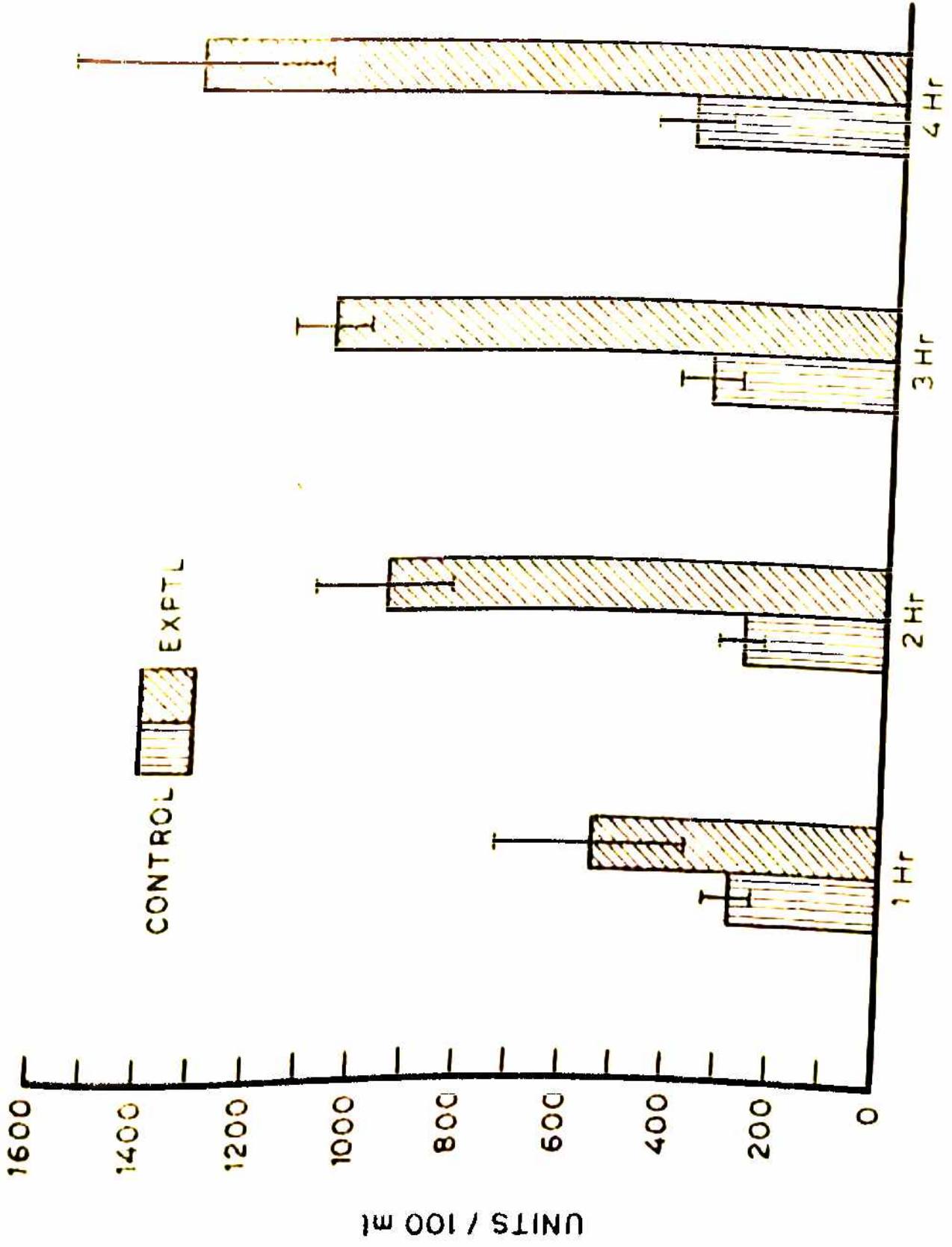
+ Shows an increase , - Shows a decrease.

\*Amylase Unit :- Is the amount of enzyme that will hydrolyze 10.0 mg of Starch in 30.0 minutes to a stage at which no color is given by iodine.

FIGURE: 7

SHOWS THE EFFECT OF ACUTE DOSE OF CARBONDISULFIDE  
ON SERUM ALPHA AMYLASE ACTIVITY ( UNITS/100 ML )  
AFTER 1 HOUR, 2 HOUR, 3 HOUR AND 4 HOUR OF  
INTRAPERITONEAL INJECTION.





SERUM -  $\alpha$  - AMYLASE

**FIG: 7**

TABLE : 8.

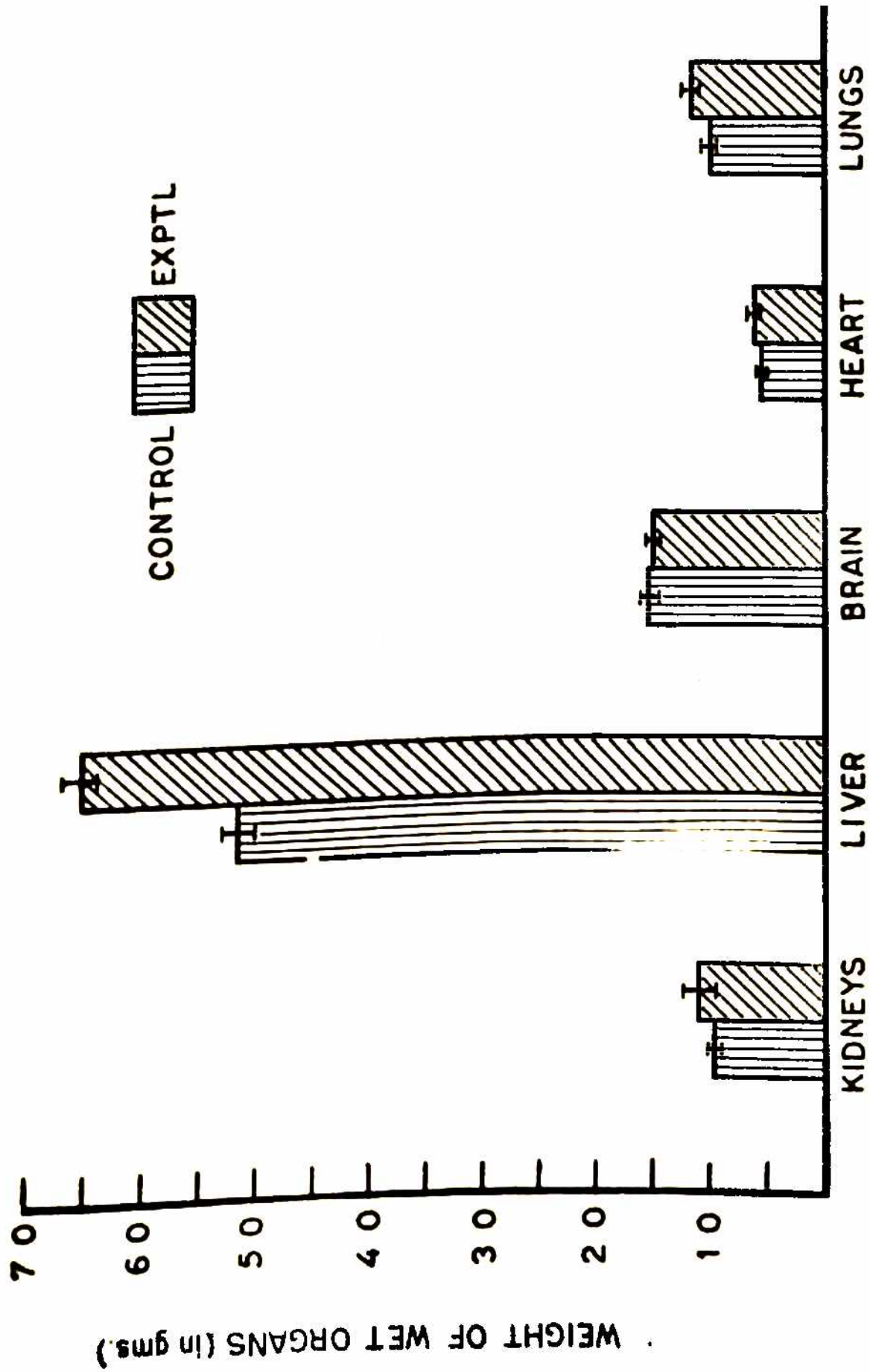
EFFECT OF CARBONDISULFIDE TREATMENT ( 25.0 mg/kg body weight ) FOR ONE MONTH  
ON THE WEIGHT OF ORGANS ( in g ms. )

Organs	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
Liver	5.351 ± .277	6.585 ± .279	+ 23.06	< .005
Brain	1.523 ± .107	1.597 ± .086	+ 4.84	< N.S.
Heart	.616 ± .033	.625 ± .088	+ .113	N.S.
Kidneys	.967 ± .04	1.112 ± .260	+ 13.03	N.S.
Lungs	1.077 ± .127	1.292 ± .129	+ 1.14	N.S.

FIGURE: 8

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT  
FOR ONE MONTH ON THE WEIGHT OF THE ORGANS OF RAT.





ORGANS

FIG: 8

TABLE: 9.

EFFECT OF CARBONDISULFIDE TREATMENT ( 25.0 mg/kg body weight) FOR ONE MONTH ON GLYCOGEN LEVEL( G/100 G tissue weight ) OF LIVER.

ORGANS	No. of Rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
LIVER	6	2.102.04 ± .327	3.676.12 ± .327	+ 74.88	< .005
BRAIN	6	.977. ± .177	1.556 ± .251	+ 52.2	< .025

FIGURE: 9

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE  
TREATMENT ON THE GLYCOGEN LEVEL OF LIVER  
AND BRAIN.



TABLE : 10.

EFFECT OF CHRONIC TREATMENT ( 25.0 mg/kg body weight) FOR ONE MONTH ON TOTAL PROTEIN LEVEL ( milligrams of Proteins/ gm tissue weight ) OF LIVER AND BRAIN.

ORGANS	Mo. of Rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significant Level.
LIVER	6	136.25 ± 4.71	114.543 ± 4.29	- 15.93	<.005
BRAIN	6	55.454 ± 7.653	86.171 ± 22.613	+ 55.39	<.1

FIGURE: 10

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE  
TREATMENT FOR ONE MONTH ON THE TOTAL PROTEIN  
LEVEL OF LIVER AND BRAIN.

TABLE : 11.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg. body weight) FOR, ONE MONTH,  
ON SERUM ALPHA AMYLASE ACTIVITY, BLOOD GLUCOSE LEVEL, PLASMA PROTEIN.

Parameter	No. of rats	* Control Units/100 ml.	CS <sub>2</sub> Treated *Units/100 ml.	Percentage Change	Significance Level.
Serum Alpha Amylase Activity	6	534.66 ± 51.52	793.33 ± 18.81	+ 49.51	<.005
Parameter	No. of rats	Control mg/100 ml.	CS <sub>2</sub> Treated mg/100 ml	Percentage Change	Significance level.
Blood Glucose	6	90.50 ± 5.60	132.50 ± 6.20	+ 44.19	<.005
_____ gm/100 ml.					
Plasma Proteins	6	10.10 ± .61	5.90 ± .54	- 41.5	<.005

\*Amylase Unit :- Is the amount of enzyme that will hydrolyze 10.0 mg of Starch in 30.0 minutes to a stage at which no color is given by iodine.



FIGURE : 11

SHOWS THE EFFECT OF CHRONIC CARBOHYDRATE TREATMENT  
FOR ONE MONTH ON THE BLOOD GLUCOSE LEVEL AND SERUM  
ALPHA AMYLASE ACTIVITY.

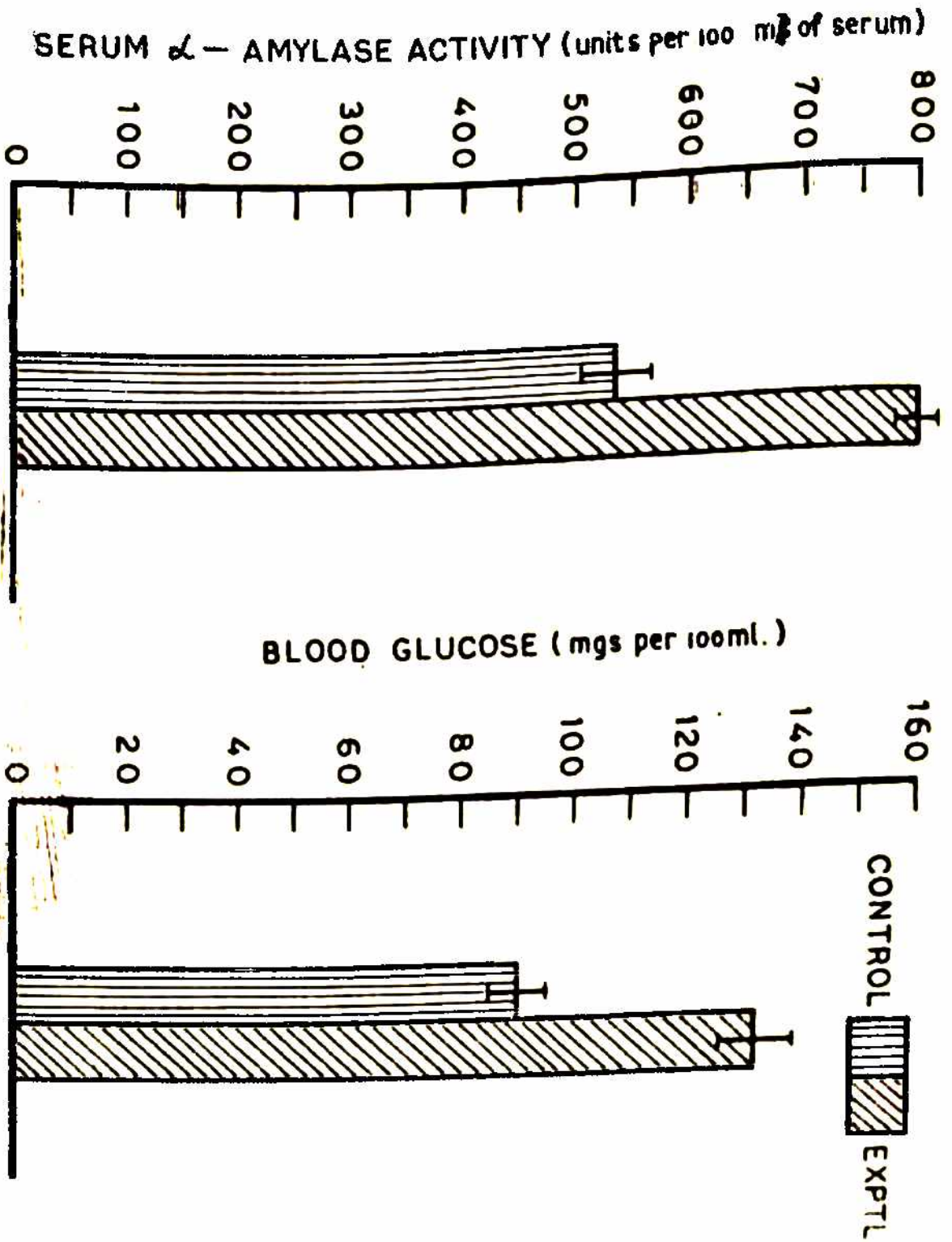


FIG: 11

TABLE : 12 .

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight), FOR ONE MONTH ON ENZYME, PHOSPHORYLASE IN LIVER AND BRAIN.

ORGANS	No. of Rats	Control Units.	CS <sub>2</sub> Treated *Units.	Percentage Change	Significance Level.
LIVER	5	1.201 ± .042	0.499 ± .0522	- 58.43	<.005
BRAIN	5	1.196 ± .101	1.076 ± .057	- 9.99	<.15 (N.S.)

One\*Unit of enzyme is that which caused the liberation of 1.0 mg of inorganic phosphorous per gm. of tissue weight in 10 minutes at 37°C.



TABLE: 13.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/ gm. body weight) , FOR ONE MONTH ON ENZYME, PHOSPHOGLUCOMUTASE ACTIVITY IN LIVER AND BRAIN.

ORGANS	No. of rats	*Control Units.	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
LIVER	6	0.489 ± 0.266	1.707 ± 0.370	+ 248.59	<.005
BRAIN	6	0.719 ± .170	1.100 ± 0.202	+ 54.61	<.1

\* Units = milligram of acid stable inorganic phosphorous liberated per gram of tissue weight per 30 minutes at 30°C.

TABLE : 14.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON ENZYME FRUCTOSE 1-6 DIPHOSPHATASE, IN LIVER AND BRAIN.

ORGANS	No. of rats	Control *Units	CS <sub>2</sub> Treated *Units.	Percentage Change	Significance Level
LIVER	6	0.476 ± 0.115	0.252 ± .080	- 42.02	<.05
BRAIN	6	0.231 ± 0.019	0.181 ± 0.015	- 21.60	<.025

\*Units = milligram of inorganic phosphorous liberated/gm of tissue weight per 20 minutes at 37°C.

FIGURE: 12

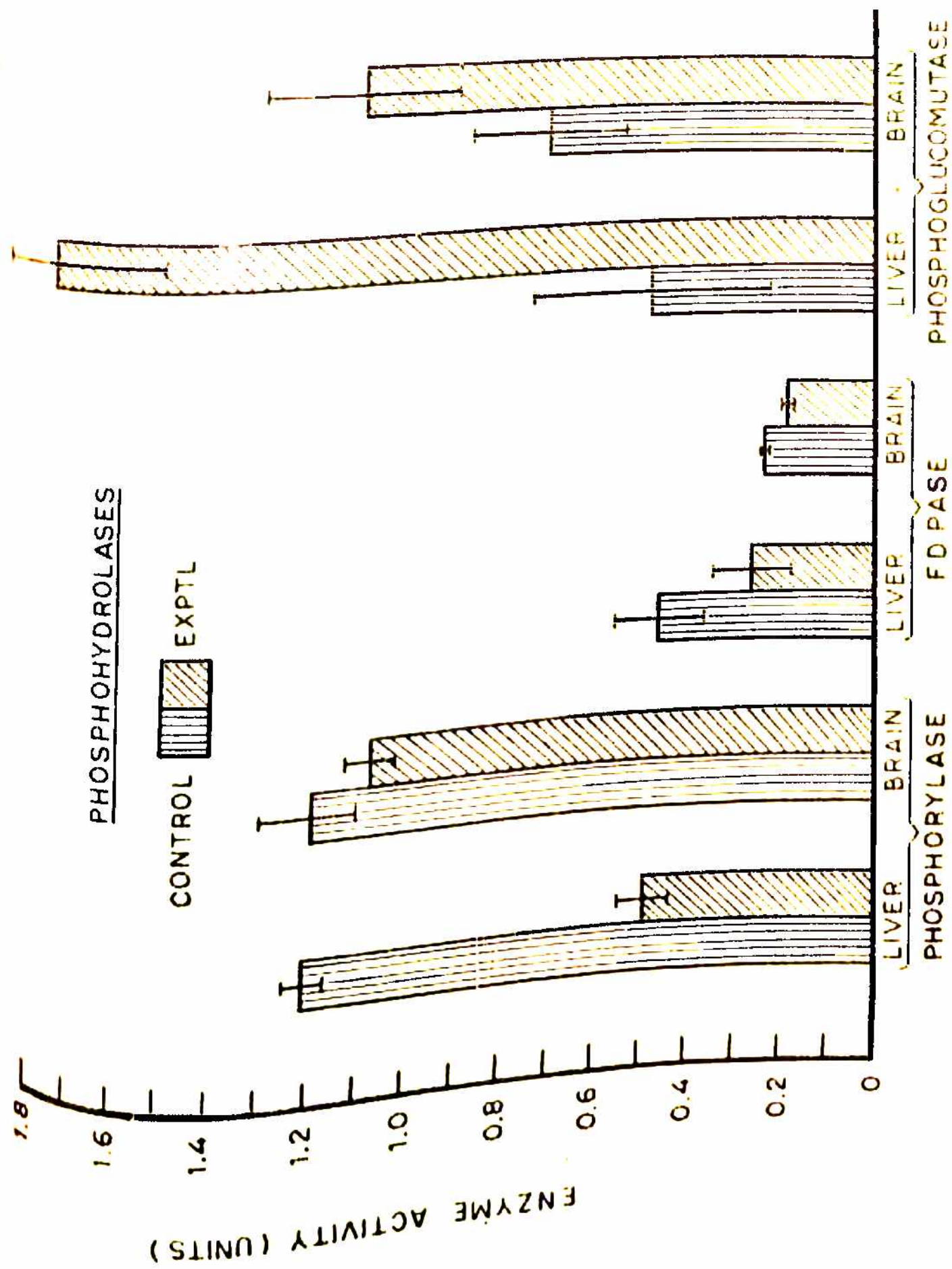
SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT FOR ONE MONTH ON THE ACTIVITY OF PHOSPHORYLASE, FRUCTOSE, 1-6 DIPHOSPHATASE ( FDPASE ) AND PHOSHO-GLUCOMUTASE IN LIVER AND BRAIN.

PHOSPHORYLASE UNIT : One unit of enzyme is that which caused the liberation of 1.0 mg of inorganic phosphorus per gram of tissue weight in 10 minutes at 37°C.

FDPase Unit : Unit activity is defined as milligrams of inorganic phosphorus liberated per gram of tissue weight in 20 minutes at 37°C.

Phosphoglucomutase Unit : Unit is defined as milligrams of acid stable inorganic phosphorus liberated per gram of tissue weight in 30 minutes at 30°C.





**FIG: 12**

TABLE : 15.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON ENZYME ADENOSINE TRIPHOSPHATASE IN LIVER AND BRAIN.

ORGANS	No. of rats	*Control Units.	CS <sub>2</sub> Treated * Units.	Percentage Change	Significance Level.
BRAIN	6	0.975 ± 0.171	0.734 ± 0.040	- 24.75	< .1
LIVER	6	1.083 ± .234	0.5606 ± .072	- 48.24	< .025

\* ATPase Unit : milligram of inorganic phosphorous liberated per gram of tissue weight in 15 minutes at 37°C.

TABLE : 16.

EFFECT OF CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON  
SUCCINIC DEHYDROGENASE ACTIVITY IN LIVER AND BRAIN.

ORGANS	Control * Units.	CS <sub>2</sub> Treated * Units.	Percentage Change	Significance Level.
LIVER	8.458 ± 1.28	5.345 ± .787	- 36.80	< .05
BRAIN	1.137 ± .258	.812 ± .108	- 28.54	< .1

\* Unit Activity is expressed as mgs of the dye (TTC) reduced per 30 minutes at 37°C.

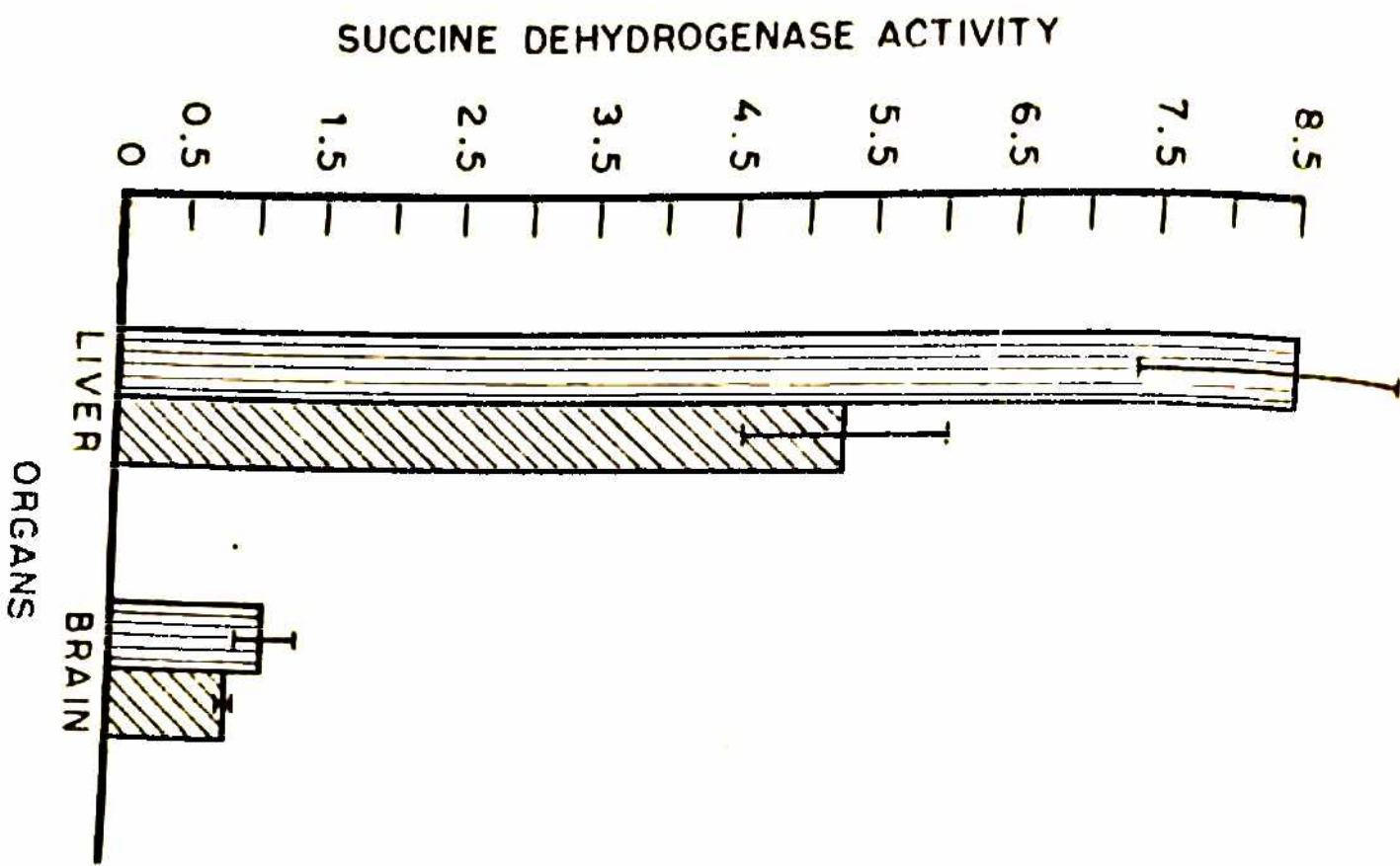
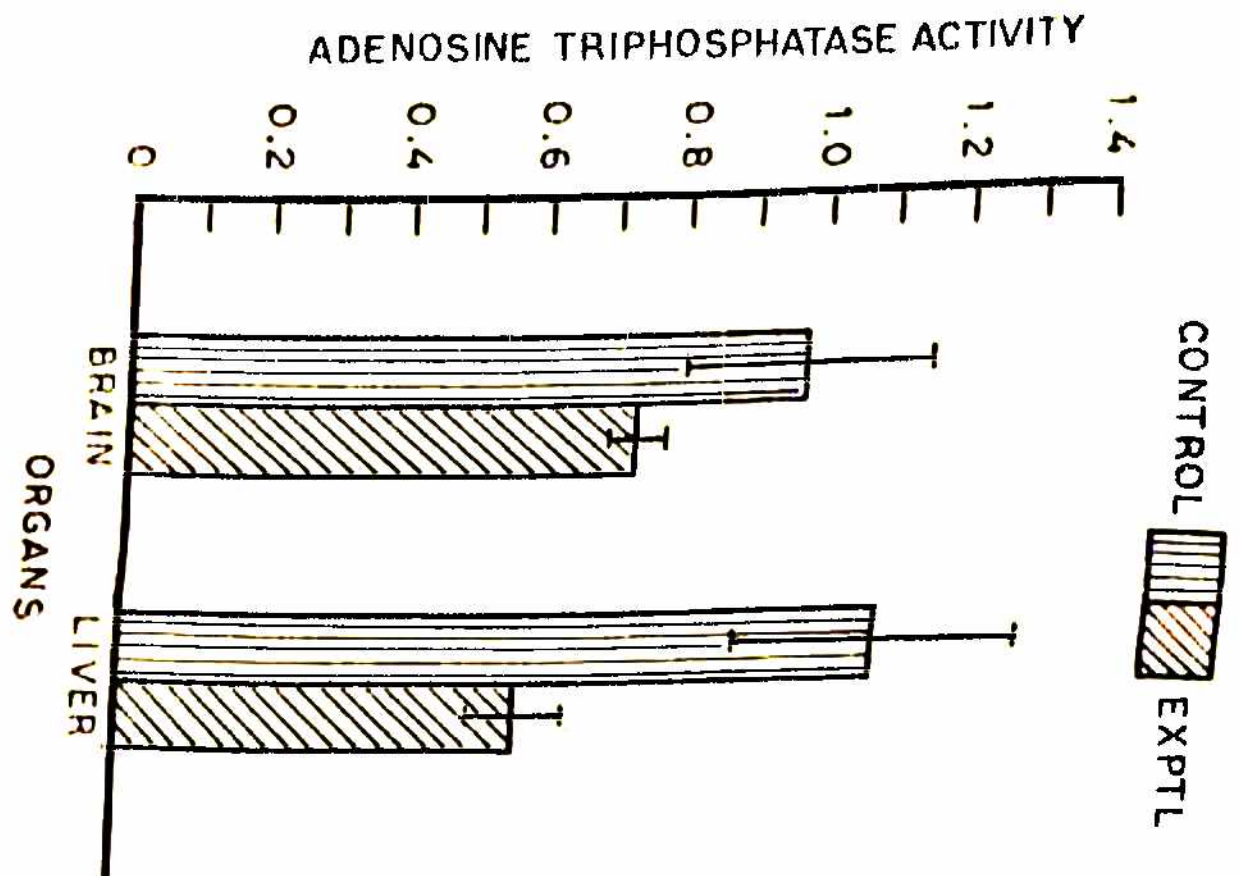
FIGURE: 13

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT FOR ONE MONTH ON THE ACTIVITY OF ADENOSINETRIPHOSPHATASE (ATPase) AND SUCCINIC DEHYDROGENASE (SDH) IN LIVER AND BRAIN.

ATPase Unit : Unit is defined as milligrams of inorganic phosphorous liberated per gram of tissue weight in 15 minutes at 37°C.

SDH Unit : Unit activity is expressed as milligrams of the dye (TTC) reduced in 30 minutes at 37°C.





**FIG : 13**

TABLE : 17.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg body weight ) FOR ONE MONTH ON DOPAMINE LEVEL ( ug/G. Tissue weight ) OF BRAIN, HEART, LIVER AND LUNGS.

ORGANS	No. of rat	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
BRAIN	5	3.094 ± .057	8.082 ± .111	+ 161.24	<.005
HEART	5	1.264 ± .089	6.418 ± .074	+ 412.65	<.005
LIVER	5	2.24 ± .045	8.13 ± .048	+ 262.94	<.005
LUNGS	5	7.542 ± .043	2.12 ± .028	- 71.89	<.005

TABLE : 18.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight ) FOR ONE MONTH, ON NOREPINEPHRINE LEVEL ( ug/G. Tissue weight ) OF BRAIN, HEART, LIVER AND LUNGS.

ORGANS	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
BRAIN	5	1.116 ± .031	2.03 ± .049	+ 86.37	<.005
HEART	5	0.39 ± .036	2.12 ± .321	+ 138.2	<.005
LIVER	5	1.136 ± .0811	1.78 ± .068	+ 56.69	<.005
LUNGS	5	0.84 ± .08	1.50 ± .054	+ 78.57	<.005

TABLE : 19.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON 5-HYDROXYTRYPTAMINE LEVEL ( ug/G.Tissue weight) OF BRAIN , HEART, LIVER AND LUNGS.

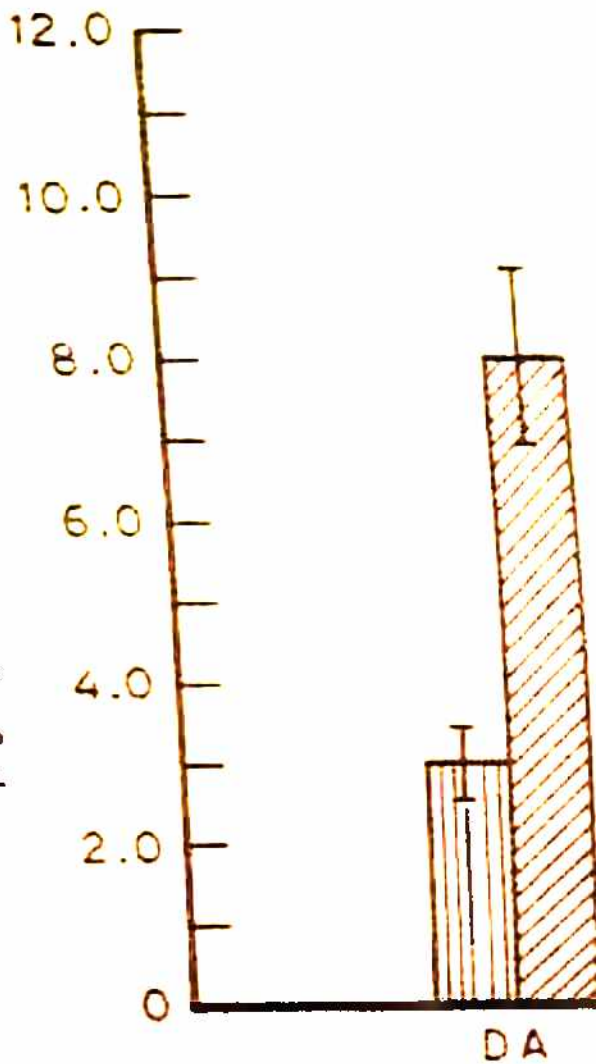
ORGANS	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
BRAIN	5	3.77 ± .08	5.50 ± .26	+ 45.88	< .005
HEART	5	2.86 ± .037	4.06 ± .185	+ 41.95	< .005
LIVER	5	.60 ± .178	2.04 ± .272	+240.0	< .005
LUNGS	5	2.20 ± .167	5.24 ± .87	+367.1	< .005



PLATE: 14

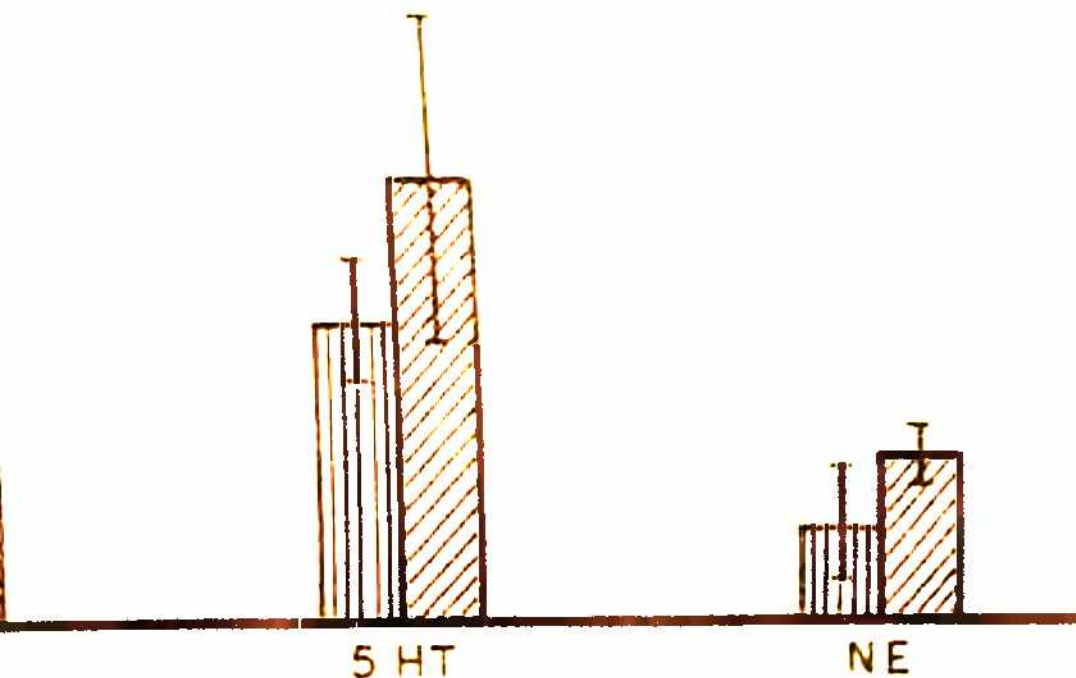
SHOWS THE AP END OF CAVITY OF SPERMATOPHYTES IN TRANSVERSE  
SECTION OF THE END OF THE SPERMATOPHYTES IN TRANSVERSE

$\mu\text{g} / \text{gm}$  WET TISSUE Wt.



BRAIN

CONTROL  EXPTL



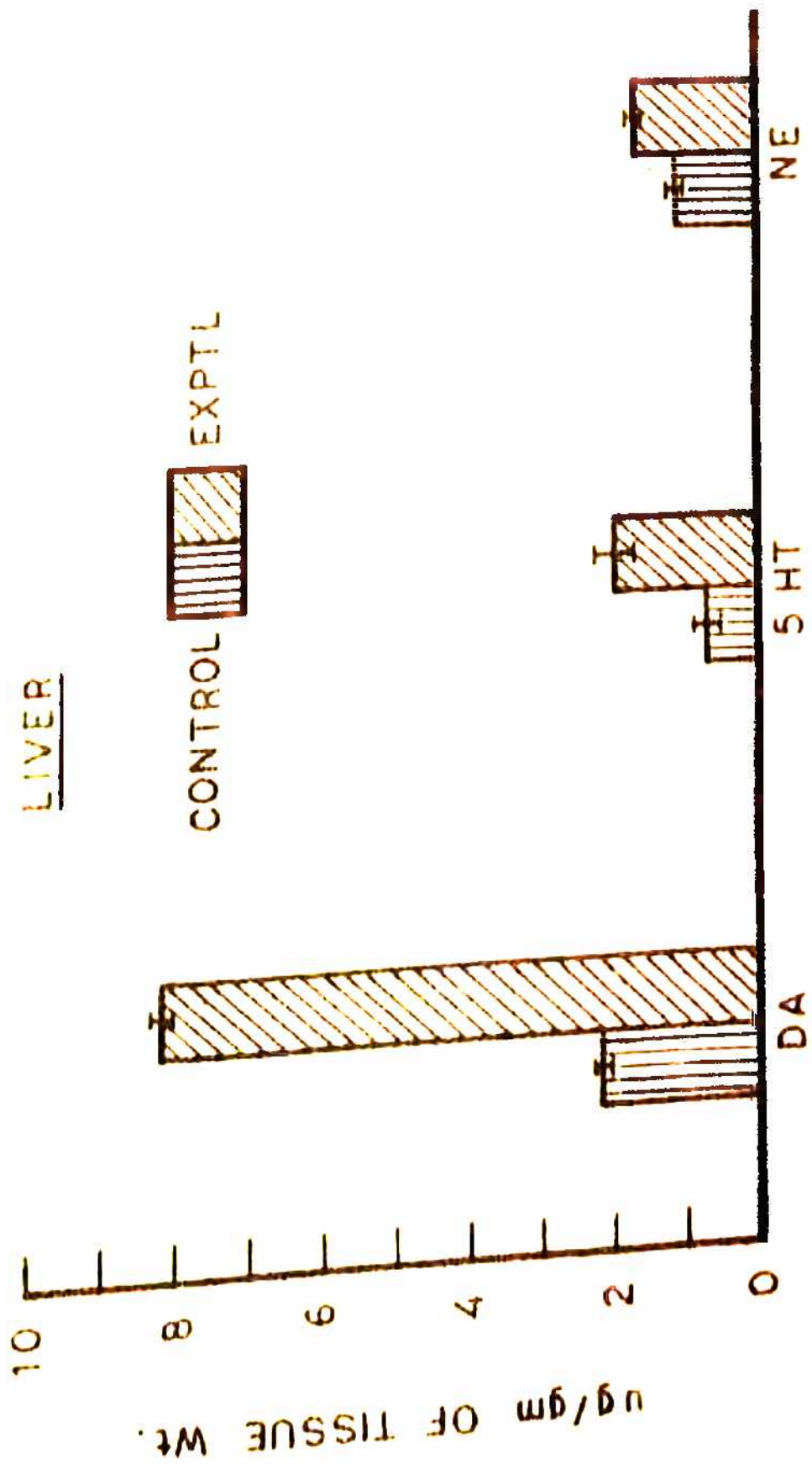
BIOGENIC AMINES

**FIG : 14**

PLATE 15

SHOWS THE SCENE OF THE OLYMPIAN TEMPLE AT ATHENS  
FOR OUR MONTH OF THE YEAR, AND THE TEMPLE OF PALLAS





BIOGENIC AMINES

FIG: 15

FIGURE 16

SHOWS THE NUMBER OF CIRCUITS OF EACH TYPE IN THE SYSTEM FOR ONE MONTH OF RA, 5-51, AND THE LEVELS OF HEAVY & LIGHTS.

WT.

ug/gm OF TISSUE

10

8

6

4

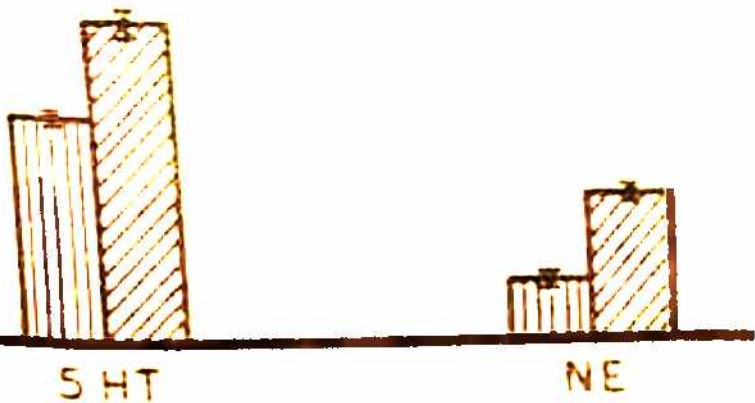
2

0

DA



HEART



BIOGENIC AMINES

**FIG: 16 A**



TABLE : 20.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight ) FOR ONE MONTH, ON MONOAMINE-OXIDASE ACTIVITY (MAO) OF BRAIN, HEART, LIVER AND LUNGS.

ORGANIS	No. of rats	Control * Units	CS <sub>2</sub> Treated * Units.	Percentage Change	Significance Level.
BRAIN	5	1.01 ± .050	0.58 ± .028	-42.57	<.005
HEART	5	.061 ± .003	.091 ± .004	+ 49.18	<.005
LIVER	5	2.35 ± .085	4.024 ± .092	+71.25	<.005
LUNGS	5	0.95 ± .036	.91 ± .040	- 4.21	N.S.

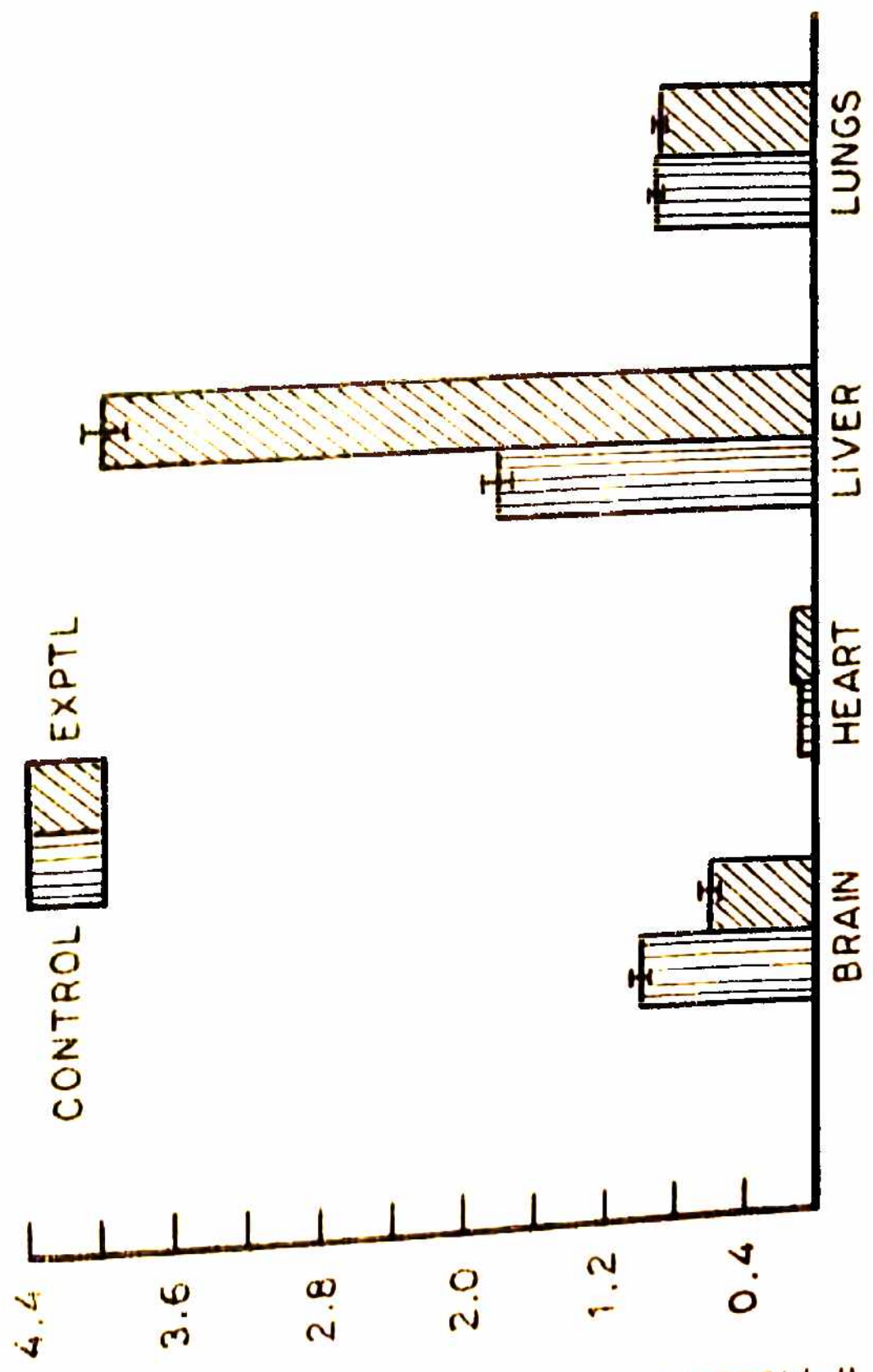
\* MAO Activity nanomoles of benzaldehyde release / gm of tissue weight/ per minute.

FIGURE: 17

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT  
FOR ONE MONTH ON MONOAMINE OXIDASE ACTIVITY OF BRAIN,  
LIVER, LUNGS AND HEART.

Monoamine Oxidase ( MAO ) Unit : Monoamine Oxidase Unit  
is defined as nano-moles of benzaldehyde produced per  
Gram of tissue weight per minute.

n- MOLES OF BENZALDEHYDE FORMED/min/gm TISSUE wt



MONO AMINE OXIDASE

FIG: 17

TABLE : 21.

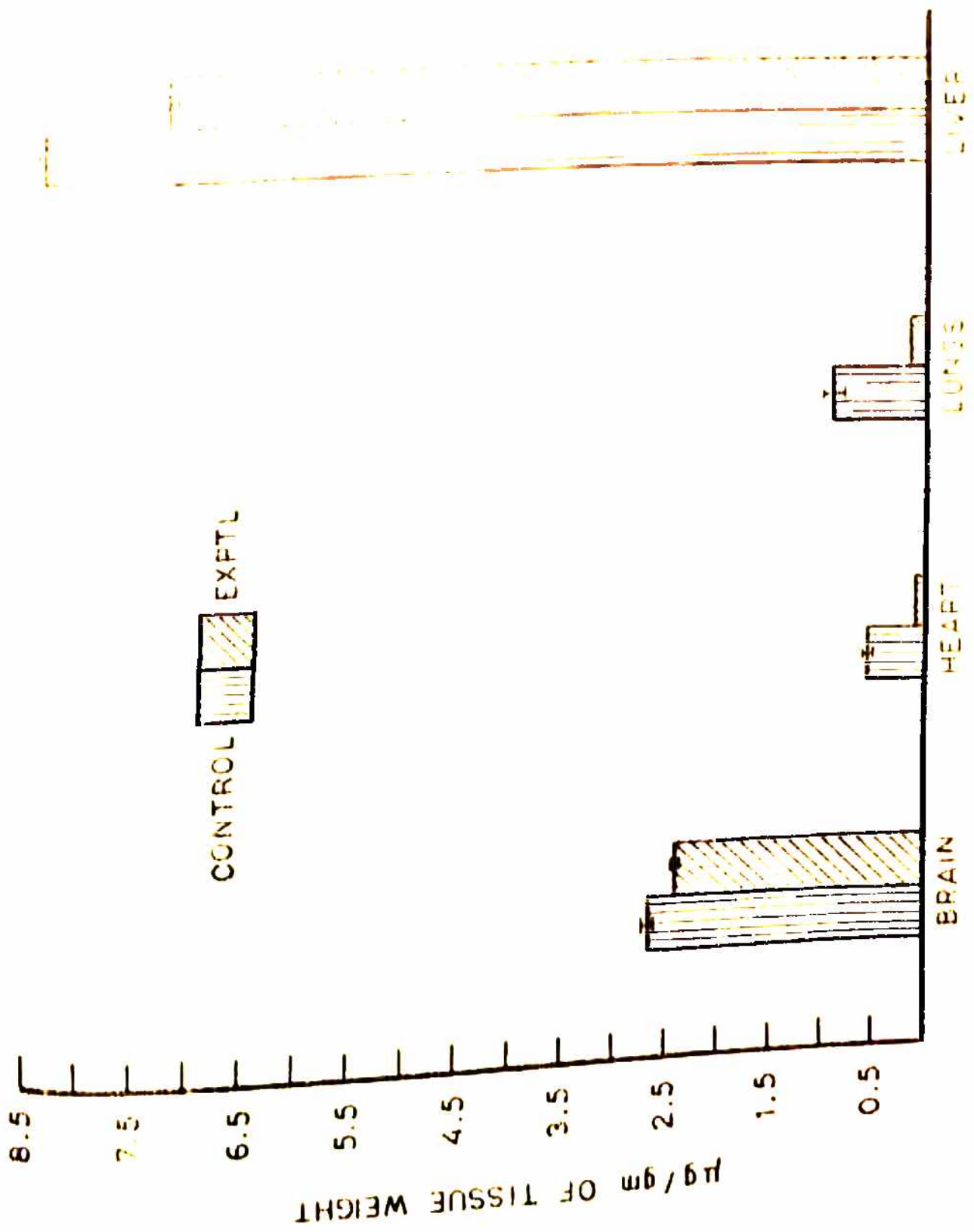
EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON GLUTATHIONE ( ug/G.Tissue weight) LEVEL OF BRAIN, HEART, LIVER AND LUNGS.

ORGANS	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance level.
BRAIN	5	2.680 ± .054	2.20 ± .028	- 17.91	<.005
HEART	5	.570 ± .034	.52 ± .0036	- 90.87	<.005
LIVER	5	8.460 ± .045	7.250 ± .070	- 14.30	<.005
LUNGS	5	.948 ± .087	.131 ± .005	- 86.18	<.005



FIGURE : 18

SHOWS THE EFFECT OF CHRONIC CARBON DISULFIDE TREATMENT  
FOR ONE MONTH ON GLUTATHIONE LEVEL OF BRAIN, LIVER,  
LUNGS AND HEART.



GLUTATHIONE (µ-SH)

FIG: 18

TABLE : 22.

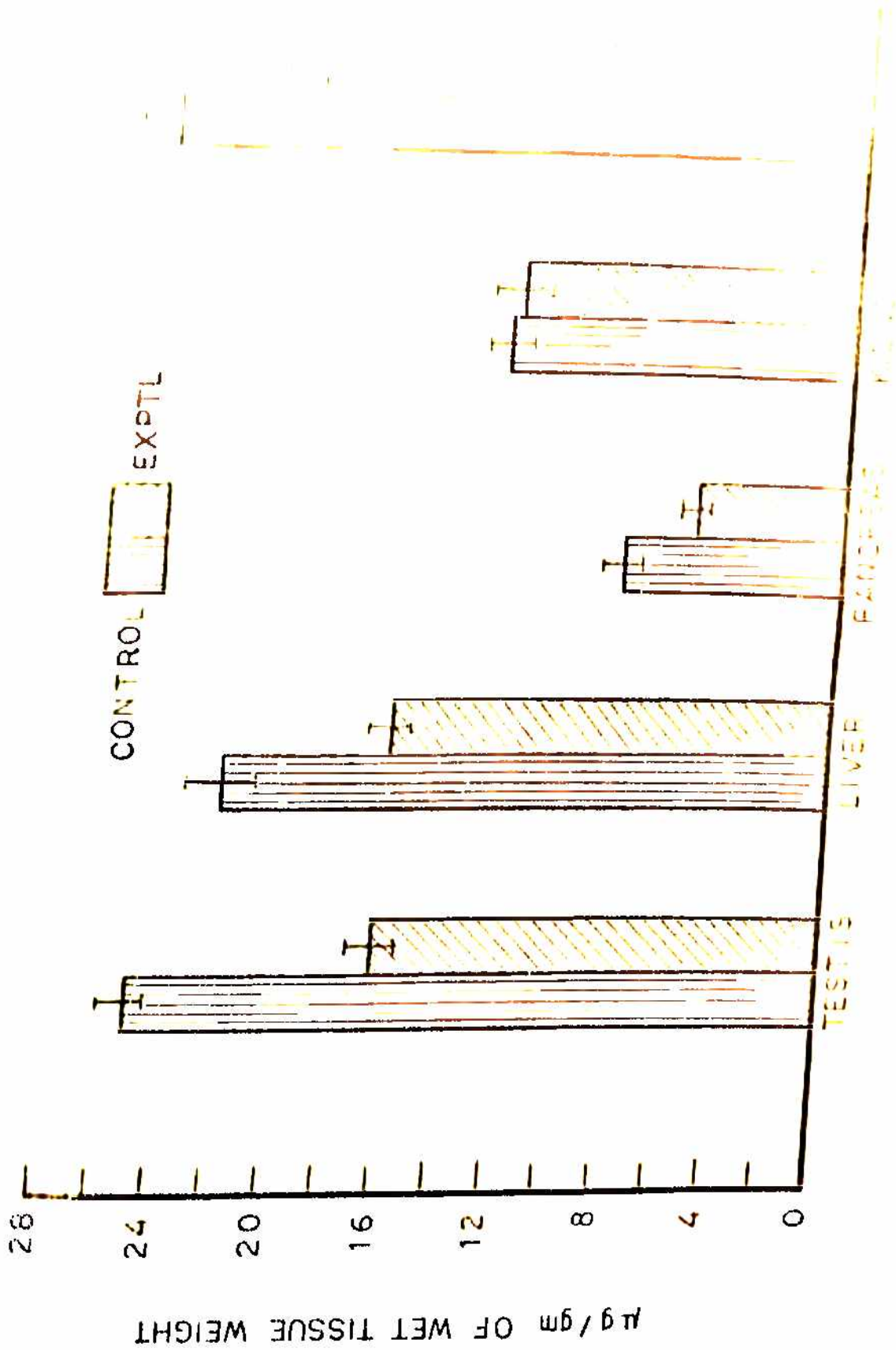
EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON ZINC LEVEL ( ug/g. Tissue weight) OF LIVER, PANCREAS, KIDNEY, TESTIS, BRAIN.

ORGANS	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
LIVER	5	22.02 ± .139	16.20 ± .066	- 26.43	< .005
PANCREAS	5	8.24 ± .0667	5.46 ± .0509	- 33.73	< .005
KIDNEY	5	12.76 ± .088	12.33 ± .090	- 2.97	N.S.
TESTIS	5	25.12 ± .082	16.36 ± .095	- 34.87	< .005
BRAIN	5	25.32 ± 1.32	20.82 ± 1.39	- 13.77	< .05

FIGURE : 19

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT  
FOR ONE MONTH ON ZINC LEVEL OF LIVER, BRAIN, TESTIS,  
PANCREAS AND KIDNEYS.





DISTRIBUTION OF ZINC IN TISSUES

**FIG: 19**

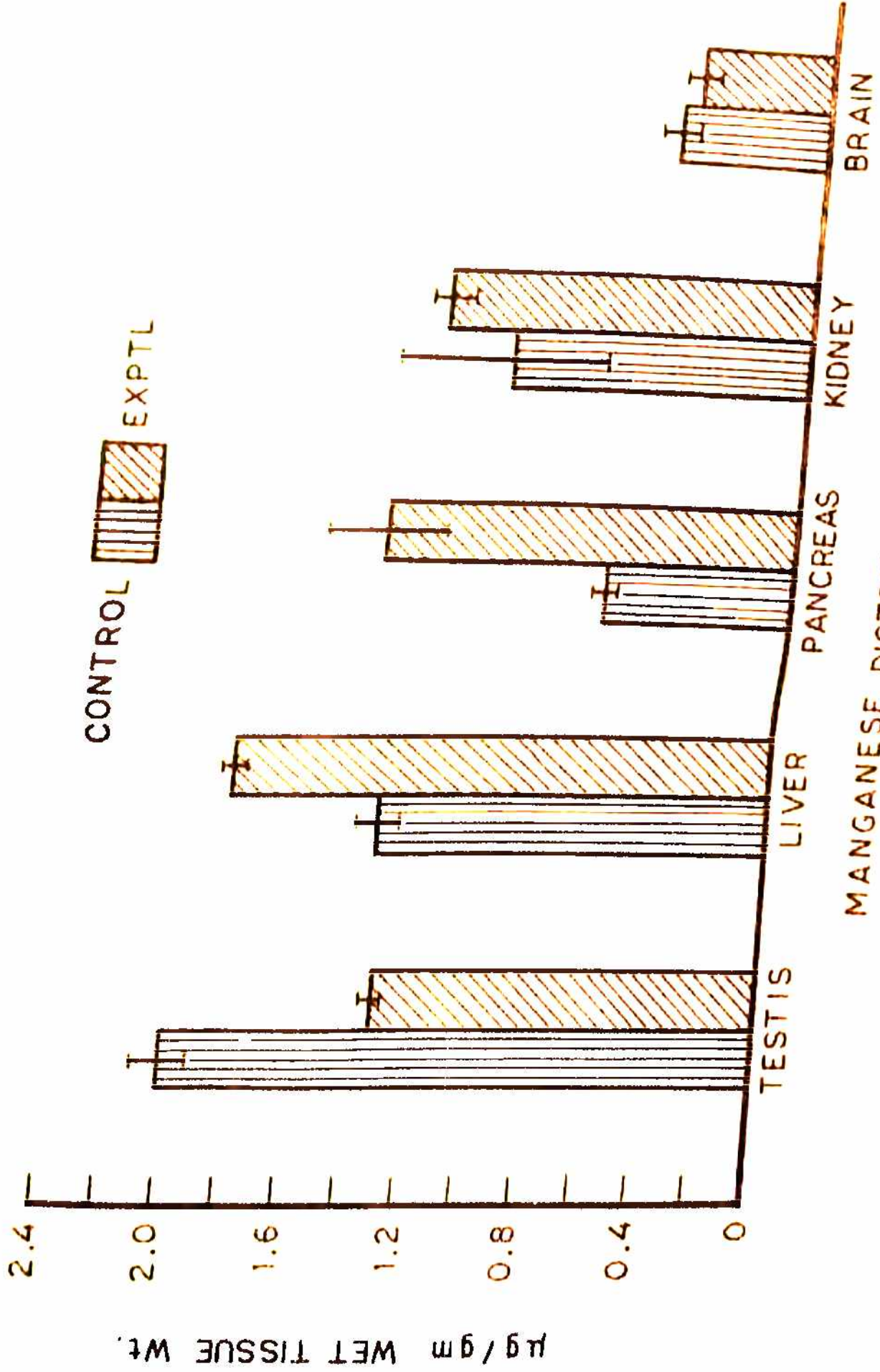
TABLE : 23.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON MANGANESE LEVEL ( ug/G. Tissue weight ) OF LIVER, PANCREAS, KIDNEY, TESTIS, BRAIN.

ORGANS	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
LIVER	5	1.35 ± .079	1.82 ± .035	+ 34.81	< .005
PANCREAS	5	0.65 ± .035	1.4 ± .254	+150.38	< .005
KIDNEY	5	1.20 ± .42	1.26 ± .035	+ 18.42	N.S
TESTIS	5	2.092 ± .126	1.32 ± .04	- 36.90	< .005
BRAIN	5	.50 ± .06	0.44 ± .05	- 12.0	N.S.

FIGURE : 20

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT  
FOR ONE MONTH ON MANGANESE LEVEL OF LIVER, BRAIN,  
TESTIS, PANCREAS AND KIDNEYS.



MANGANESE DISTRIBUTION IN TISSUES  
**FIG : 20**



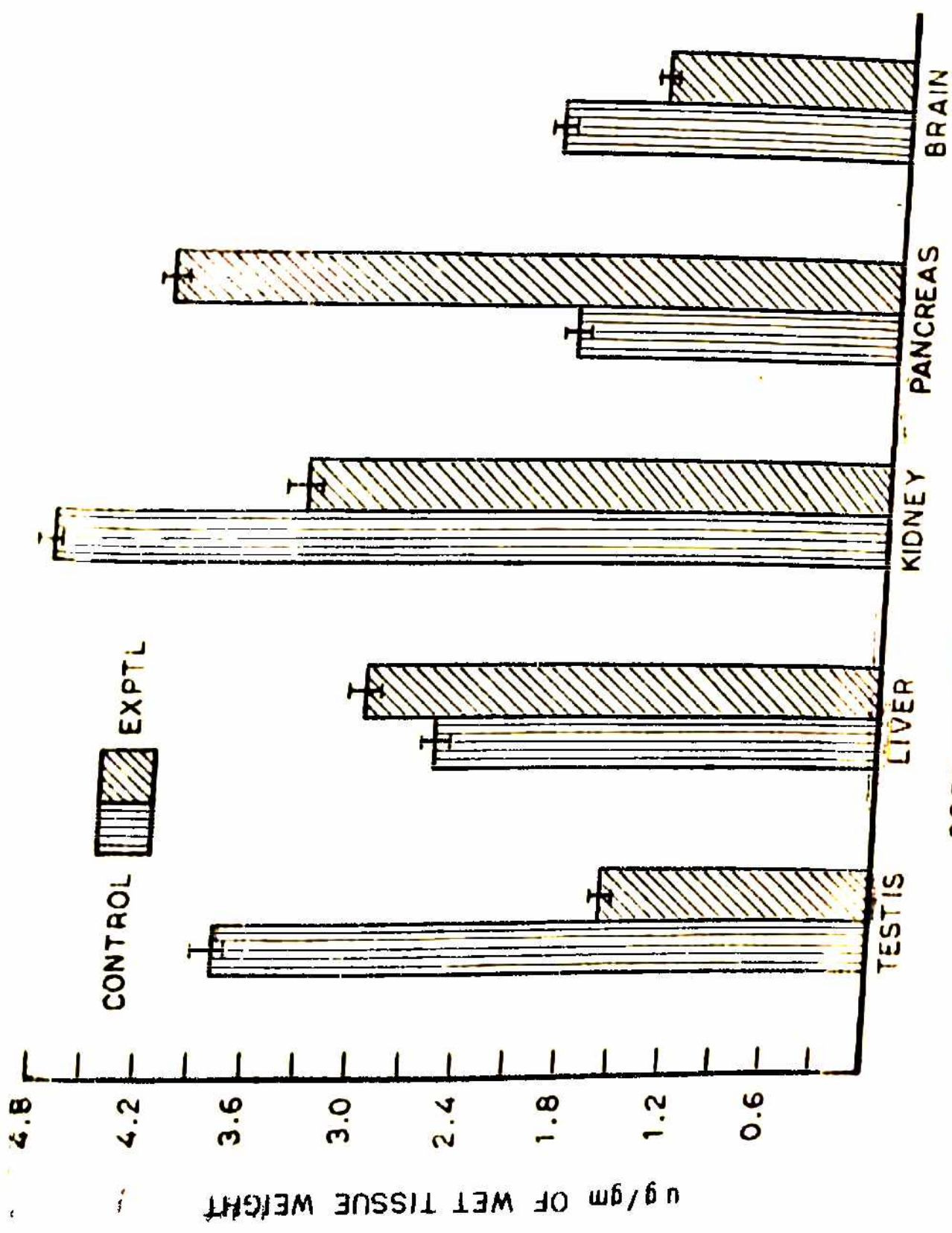
TABLE : 24.

EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT ( 25.0 mg/kg, body weight) FOR ONE MONTH ON COPPER LEVEL ( ug/G. Tissue weight) OF LIVER, PANCREAS, KIDNEY, TESTIS, BRAIN.

ORGANS	No. of rats	Control	CS <sub>2</sub> Treated	Percentage Change	Significance Level.
LIVER	5	2.62 ± .076	3.00 ± .085	+ 14.50	< .005
PANCREAS	5	1.66 ± .073	4.23 ± .060	+ 157.83	< .005
KIDNEY	5	4.88 ± .066	3.40 ± .088	- 30.32	< .005
TESTIS	5	3.88 ± .063	1.56 ± .055	- 59.79	< .005
BRAIN	5	2.02 ± .056	1.47 ± .048	- 27.22	< .005

FIGURE : 21

SHOWS THE EFFECT OF CHRONIC CARBONDISULFIDE TREATMENT  
FOR ONE MONTH ON COPPER LEVEL OF LIVER, BRAIN, TESTIS,  
PANCREAS AND KIDNEYS.



COPPER DISTRIBUTION IN TISSUES  
**FIG : 21**

CHAPTER - 4

DISCUSSION



#### 4.1 EFFECT OF ACUTE CARBONDISULFIDE TREATMENT :-

##### 4.1.1. GLYCOGEN :-

The acute damage to liver and brain have been considered to be the important manifestations of acute CS<sub>2</sub> poisoning ( Seppalainen, 1977 ; DeMatteis and Seawright, 1976; Jarvisalo and Gibbs, 1978; Mckenna and Victor, 1977). The damage to the liver appeared to be more intensive because of apparent sudden release of stored glycogen in the animals from 1 to 4 hour after CS<sub>2</sub> injection ( P < .005 ). This finding support the observations of Kurzinger and Freundt (1969), who reported a sharp decline in liver glycogen by CS<sub>2</sub> intoxication.

The brain glycogen has been, however, found to be unaffected during the period of observation, which reflects that brain glycogen might be bound more firmly and CS<sub>2</sub> cannot affect the bound glycogen. Kacew and Singhal (1973) have also demonstrated that administration of p - p', DDT produced a marked decrease in concentration of hepatic glycogen. The acute administration of CS<sub>2</sub> in a large dose appears to produce the damage to the adrenals and thus results in the release of catecholamines. It is, therefore, possible that these endogeneously liberated catecholamines have produced the breakdown of liver glycogen.

#### 4.1.2. BLOOD GLUCOSE :-

Blood glucose is effected drastically by acute CS<sub>2</sub> treatment as it significantly increased in first hour ( P < .005) The fall of glucose level which has its onset from second hour persisted till the fourth hour, confirmed the earlier reports of the fall in blood glucose level by CS<sub>2</sub> toxicity in rat (Cunningham, 1975), Cunningham and Daphne (1977), also studied the turnover of blood glucose under CS<sub>2</sub> intoxication. Katz et.al. (1974) concluded that fall in blood glucose has been at triosephosphate level and gluconeogenesis is not stimulated. The initial rise in blood glucose in first hour may therefore, be due to imbalance of released glucose and insulin to rapid hepatic glycogen breakdown as have been suggested earlier. But in the later hours i.e. from second hour onward perhaps, glucose utilisation have increased or muscular glycogen deposition has enhanced due to increased insulin in blood.

#### 4.1.3. SERUM ALPHA AMYLASE :-

Serum alpha amylase activity has been found to be increased sharply after CS<sub>2</sub> treatment ( P < .005 ). This increased serum alpha amylase activity has been a proof of the damage to pancreas, which is involved in carbohydrate metabolism (Carway, 1959 ). Therefore, the sharp decline in blood glucose during acute intoxication alongwith parallel rise in serum alpha amylase activity confirms our assumption of the sudden release of insulin from pancreatic storage granules. Hence,

this mechanism is responsible for higher circulating level of insulin and increased glucose turnover (Cunningham et.al. 1976).

#### 4.1.4. PROTEINS :-

Total protein level in tissues, as well as in the plasma showed a marked fall during first hour of acute CS<sub>2</sub> intoxication. This finding confirms the earlier suggestion of Bond and De Matteis (1969) that CS<sub>2</sub> intoxication decreased the synthesis of liver proteins or depletion of proteins along with hyperaminociduria and hyperaminoacidemia (EL'Dessoukey et.al. 1977) or the increase turnover of proteins (Jarvisalo 1977); or higher level of urea in the plasma (Cunningham, 1975). These observations tend this study to propose that protein degradation is enhanced under acute CS<sub>2</sub> toxicity, which may be providing fuel for energy and gives evidence for weight loss in man and animals exposed to CS<sub>2</sub>.

The acute studies of CS<sub>2</sub> toxicity therefore, lead to suggest that carbohydrate metabolism is immediately affected. The liver appears to be the main site of action of CS<sub>2</sub> and liver glycogen gets depleted rapidly which in turn lead to hyperglycemia. The hypoglycemia along with glycogen depletion from liver indicates the rapid utilization of glucose which could be attributed to the observed rapid and faster respiration rate. Further, the mobilization of glycogen from liver to muscles



might also play some role. The increased serum alpha amylase activity is an indicator of the pancreatic damage which caused the excessive release of insulin and hence may increase peripheral utilization of glucose. The decreased liver proteins and plasma proteins also support the assumption of mobilization of proteins from liver to muscles. Further, this study has provided evidence that CS<sub>2</sub> might be responsible for the enhanced degradation of proteins in liver for energy metabolism, and probable rise of blood urea. (Cunningham et.al. 1975).

#### 4.2. EFFECT OF CHRONIC CARBONDISULFIDE TOXICITY :-

##### 4.2.1. GLYCOGEN :-

Glycogen of liver ( P <.005 ) and brain ( P <.025 ) increased significantly with one month chronic treatment of CS<sub>2</sub>. This observation, however, are in contrast to the effect of p-p'. DDT observed by Kacew and Singhal (1973), who observed a decrease in hepatic glycogen after its chronic treatment. Therefore, the sites of hepatic toxicity of CS<sub>2</sub> and p-p'. DDT could be different and the rise in glycogen in CS<sub>2</sub> poisoning might be due to increased glycogen synthesis or/and decreased glycogen breakdown as is revealed by the study of enzymes described further in this work.

##### 4.2.2. BLOOD GLUCOSE :-

The permanent rise in blood glucose may be due to



the uniformly decreased peripheral utilization of glucose or inhibition of glycolysis in liver and muscles as also observed with p-p'- DDT by Kacew. and Singhal (1973). This could be due to inhibition of glucose metabolising enzymes and increased serum alpha amylase activity. The increase in enzyme activities has been attributed to pancreatic damage, which probably is responsible for decreased glucose utilization for the energy production and increased level of glucose in blood. These observations, therefore, tend to give some clue to the observed higher blood glucose and thus diabetogenic tendencies in workers exposed to CS<sub>2</sub> for long duration of time ( Gotto et.al. 1971; Hernberg et.al. 1977).

#### 4.2.3.1 TOTAL PROTEINS :-

The chronic treatment of CS<sub>2</sub> is not found to effect the liver and brain proteins significantly whereas plasma proteins have been found to be decreased significantly,  $P < .005$  (Jarvisalo et.al. 1977; Cunningham et.al. 1975 ). The reduction in plasma protein level was related to the increased turnover of proteins and higher urea excretion and urea concentration in plasma by earlier workers ( Cunningham et.al. 1977 ; Jarvisalo et.al. 1977). These observations also confirm our previous contention that liver is more susceptible than brain in CS<sub>2</sub> toxicity.

FIGURE: 22

SHOWS THE PATHWAY OF GLYCOGEN SYNTHESIS AND STIMULATION CAUSED BY CARBONDISULFIDE OF PHOSPHOGLUCOMUTASE AND INHIBITION OF PHOSPHORYLASE AND FRUCTOSE, 1-6 DIPHOSPHATASE IN LIVER.

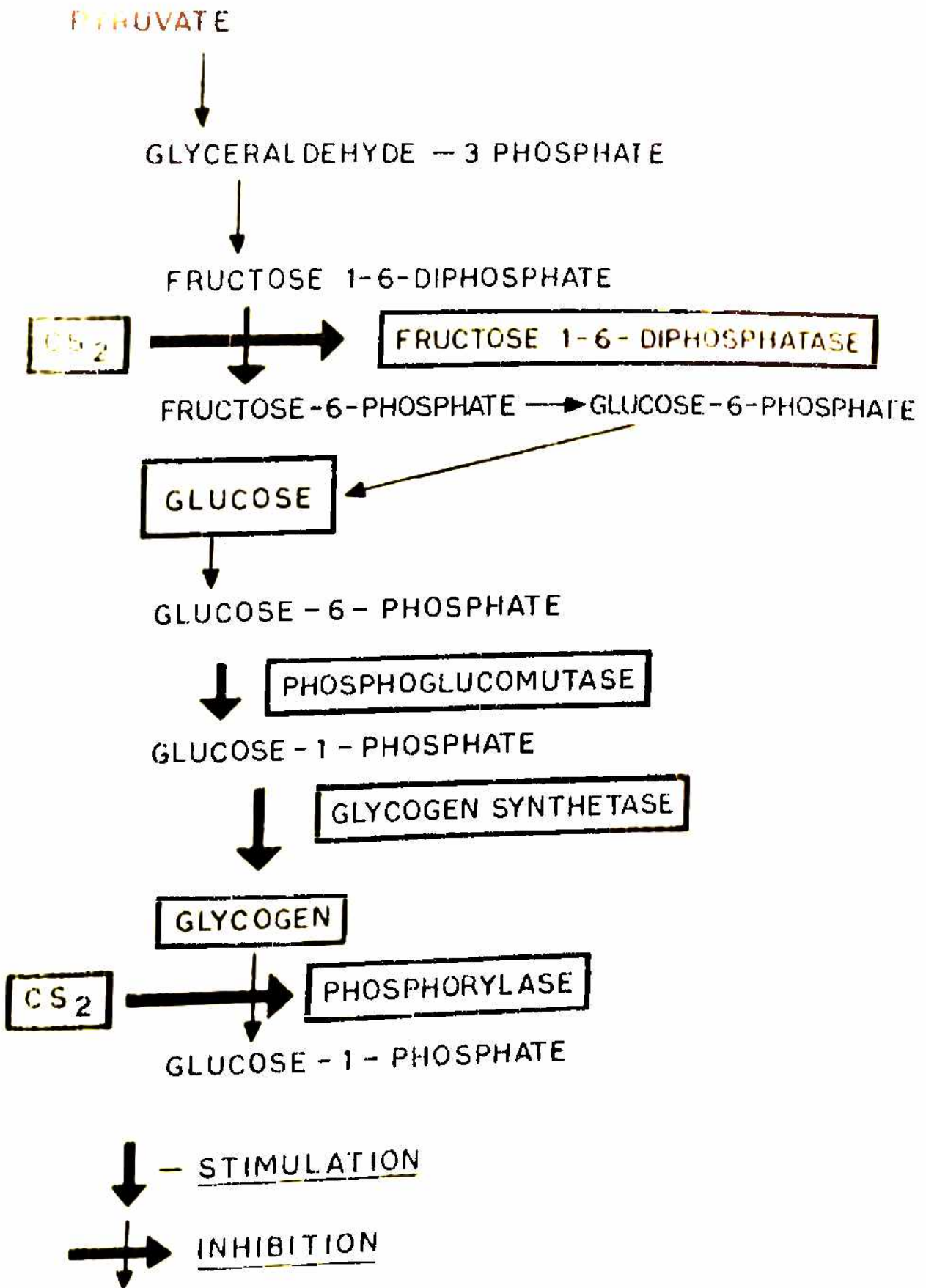


FIG: 22

#### 4.2.4. PHOSPHORYLASE :-

The reduction of liver phosphorylase (  $P < .005$  ), supported the earlier observations of Chandra et.al. (1972), who too observed the reduction of phosphorylase histochemically. The fall in phosphorylase activity leading to retardation of glycogen degradation may therefore be responsible for the accumulation of glycogen in the liver. The brain phosphorylase activity again, observed to be unaffected which confirms further, that the increase in brain glycogen is due to increased glycogen synthesis rather than decreased breakdown as observed in liver.

#### 4.2.5. PHOSPHOGLUCOMUTASE :-

Glucose is converted to glucose 6-phosphate and then to glucose 1-phosphate by phosphoglucomutase, which then get transformed to glycogen by glycogen synthetase system. Hockin et.al. (1968) reported a decrease in the activity of glucose 6-phosphate dehydrogenase, hexokinase and glucokinase in rabbits after  $CS_2$  exposure. These results showed that there is a decrease in catabolism of glucose by glycolysis and hexose monophosphate shunt ( HMP ) pathway. Therefore, the present observation of decreased phosphorylase and enhanced phosphoglucomutase with chronic  $CS_2$  treatment tempts one to assume that more of glucose is channelised for the formation of glycogen rather than its normal degradation. Thus, the increased glycogen content in liver and brain can be attributed to the rise in phosphoglucomutase.



#### 4.2.6. ADENOSINETRIPHOSPHATASE :-

Sidrorwicz et.al.' (1977), while studying the effect of CS<sub>2</sub> on red cell metabolism in viscose rayon workers, reported a decreased level of ATP and increased level of AMP, ADP. The present study has also observed a decrease in ATPase, thus leading to the reduction of oxidative phosphorylation phenomena. This observation may contribute an explanation to hypoxia observed by Sidrorwicz et.al.' (1977) by CS<sub>2</sub> toxicity. The ATPase inhibition by CS<sub>2</sub> has been found to affect the membrane transport of metabolites and thus be responsible for the disturbance in metabolic pathways (Srivastava et.al. 1975 ).

#### 4.2.7. FRUCTOSE, 1-6 DIPHOSPHATASE ( FDPase ) :-

Ol'Khovaskaya (1976) found the inhibited activity of aldolase due to inhibition of glycolysis in erythrocytes in CS<sub>2</sub> toxicity. The present study has revealed the fall in FDPase activity which, therefore, suggests the inhibition of gluconeogenesis ( Cunningham and Holt, 1977). Therefore, the chronic treatment with CS<sub>2</sub> can be observed to reduce the glucose formation by gluconeogenesis from noncarbohydrate precursors. The inhibition of FDPase confirms that increased glucose level may be due to decreased glycolysis rather than increased gluconeogenesis.

#### 4.2.8. SUCCINIC DEHYDROGENASE :-

Succinic dehydrogenase is a key enzyme of Kreb's cycle

and links with energy utilisation and  $CS_2$  inhibits the energy metabolising enzymes of liver and brain ( i.e., brain and liver ATPase and succinic dehydrogenase ). The SDH and ATPase are considered to be important since these are membrane bound enzymes (Murasko and Gubskis, 1976 ). The inhibition of these enzymes showed an alteration of membrane transport, which can be an interaction of  $CS_2$  with mitochondrial membrane. This chemical intoxication may thus, lead to some kind of derangement in the membrane pattern of lipids of the mitochondria, which could lead to alteration in the energy metabolism of carbohydrates.

It may, therefore, be proposed from the study of chronic treatment that  $CS_2$  inhibits the glucose breakdown caused by glycolysis or HMP pathway alongwith inhibition of gluconeogenesis. This phenomena has been due to the decreased glucose breakdown and causes the reduction in the synthesis of glucose. It could be the responsible phenomenon for the stimulation of the biosynthesis of glycogen and also reduction in glycogen breakdown. The membrane bound mitochondrial enzymes inhibition resulted in the reduced respiration rate and normal functioning of Kreb's cycle or electron transport chain. However, the energy production could be due to an increased protein breakdown rather than glucose metabolism.

FIGURE : 23

SHOWS THE RELATIONSHIP BETWEEN GLYCOGEN, PHOSPHORYLASE  
AND PHOSPHOGLUCOMUTASE UNDER THE STRESS OF CARBONDI-  
SULFIDE.

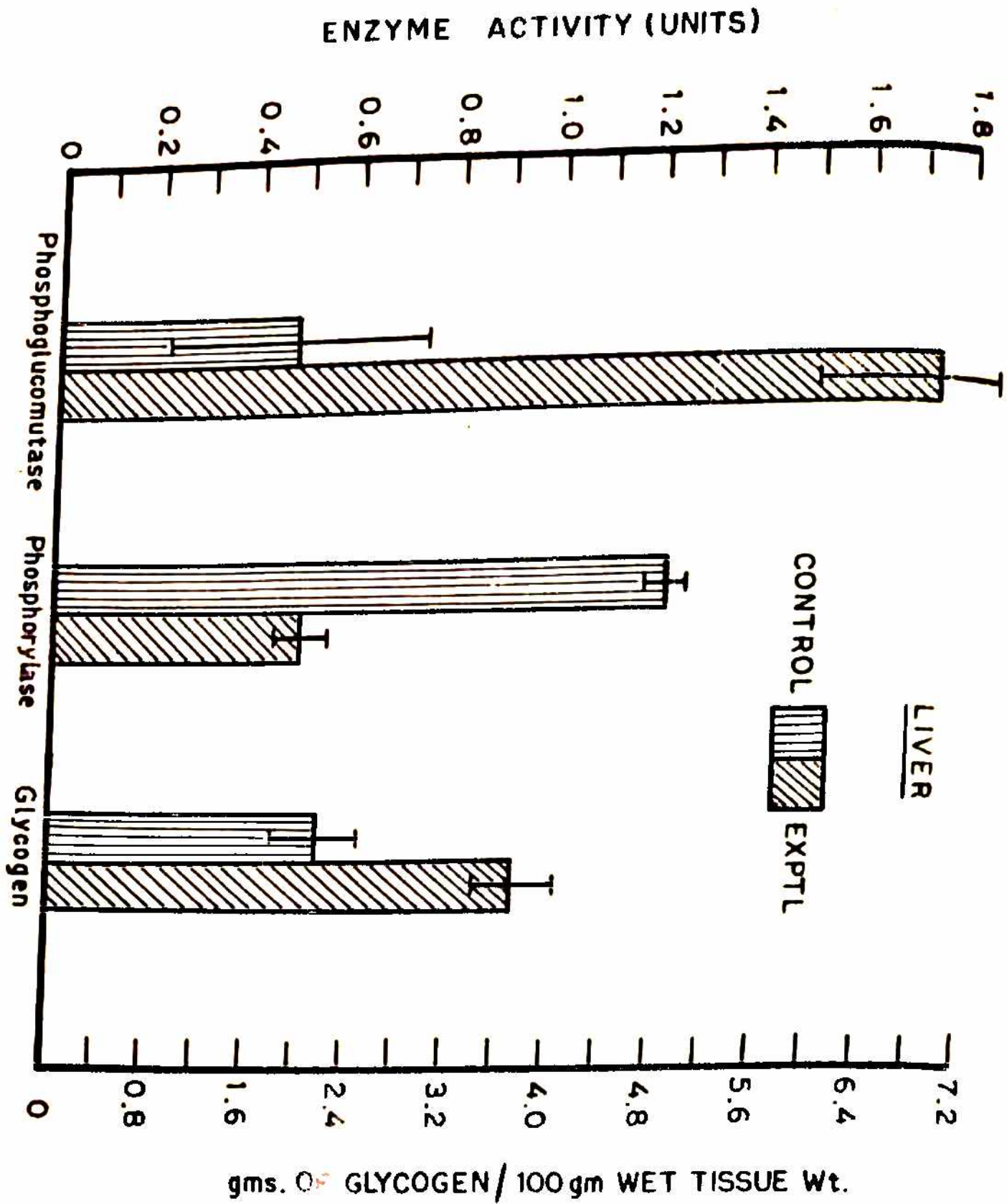


FIG: 23



#### 4.2.9. ACUTE VS CHRONIC CS<sub>2</sub> TOXICITY :-

Chronic biochemical changes are more firm qualitatively and quantitatively in contrast to that of acute studies. The chronic studies revealed that CS<sub>2</sub> might be inhibiting glycolysis as well as gluconeogenesis which have been shown by the inhibition of FDPase, SDH and ATPase. But the rise in glycogen level is observed which is attributed to the increased phosphoglucomutase and decreased phosphorylase activity. In acute studies, the glycogen depletion is noticed along with hypoglycemia in later hours which suggested a quick mobilization of glycogen from liver to peripheral tissues. The glycolysis may be supposed not to be inhibited under acute toxicity because of lowering of blood glucose level which occur suddenly in response to insulin release from pancreas.

#### 4.3. ALTERATIONS OF BIOGENIC AMINES BY CHRONIC CARBONDISULFIDE TREATMENT :-

##### 4.3.1. CARBONDISULFIDE TOXICITY AND CATECHOLAMINES :-

It is well established that CS<sub>2</sub> forms dithiocarbamate with catecholamines in the tissues undergoing metabolic conversion. Most of the catecholamine metabolising enzymes are also reported to be inhibited by the action of these dithiocarbamates (Stripp et al. 1969). A few of the studies on acute treatment

with CS<sub>2</sub> have revealed that brain NE level is decreased while observations with brain DA are varied (Stripp et.al. 1969). Maj and Vetulani (1970) found that N-N'-disubstituted dithiocarbamates synthesized in the nerve granules, by the action of catecholamines with CS<sub>2</sub>, have been responsible for the reduction of brain NE content and spontaneous activity in animals. Magos and Jarvis (1970) found that rats exposed to CS<sub>2</sub> for two days showed a decrease in NE and increase in DA, and found no change in monoamine oxidase (MAO) activity. This may be attributed to the increased DA uptake, (McKenna and Distefano 1977). They suggested that the reduction in rat brain NE and DA level has been due to the formation of diethyldithiocarbamate complex formed by interaction of CA with CS<sub>2</sub> on chronic or prolonged treatment. But the present chronic treatment study could not confirm their contention as there has been an increase of DA level in chronic treatment which indicated that granular DA, more firmly bound, may not be available for complex formation. Further, the rise in brain NE concentration may be partially attributed to the decreased brain MAO activity ( P < .005) and unaffected granular DA.

The simultaneous studies on the estimation of biogenic amines of liver and heart have focussed to the interesting observations of increased MAO activity alongwith an increase of both DA and NE. These observations suggest that dopamine



beta-hydroxylase of liver may not be inhibited with chronic CS<sub>2</sub> treatment as observed in plasma under stress (Schanberg and Krishner, 1976 ; Mckenna and Distefano 1977). This suggested that chronic CS<sub>2</sub> intoxication caused stress and does not appear to affect the dopamine beta-hydroxylase as much seen with acute treatment. This statement is further augmented by an observed increase of copper made available for DBH in liver.

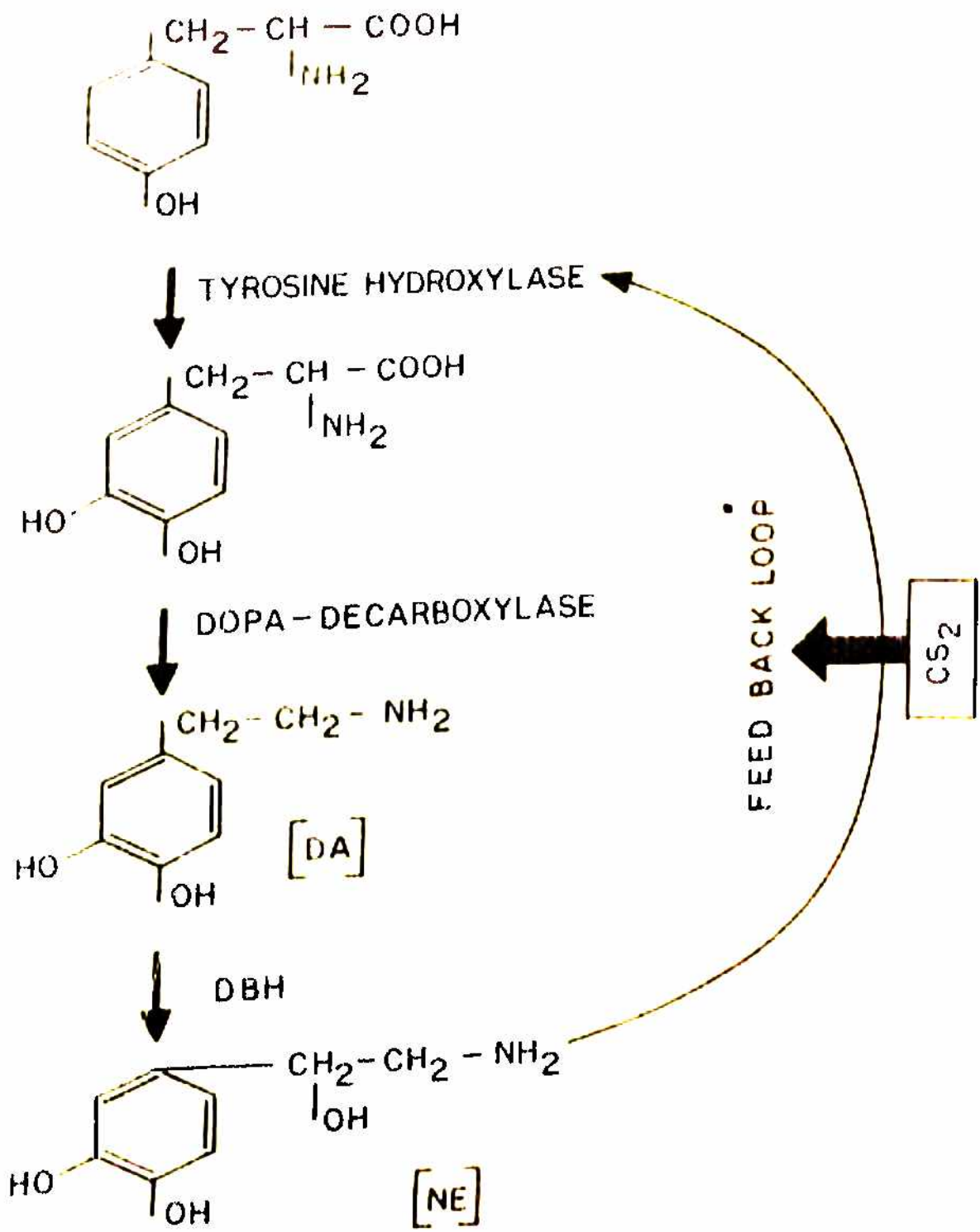
Reduced glutathione (G-SH), a protecting agent against many toxicants has been known to prevent the covalent binding of toxic substances to the important enzymes and proteins. The glutathione was reported to be decreased appreciably in the present studies as observed by earlier workers (Corsini, 1972; Read et.al. 1972; Jallow et.al. 1971; Glende, 1972). The reduction in G-SH may therefore, contribute for the protective action against the reduction in enzymes like dopamine beta-hydroxylase and MAO in liver and heart. It is hence possible that the increased available tyrosine is directed for the synthesis of biogenic amines and thus causing an increase in NE and DA synthesis. This hypothesis is based on the results of Teisinger (1971), who reported that chronic CS<sub>2</sub> poisoning caused the deficiency of pyridoxalphosphate, a coenzyme responsible for decarboxylation and transamination reactions, required for normal tyrosine metabolism.

On the similar basis the elevation of NE and reduction

FIGURE : 24

SHOWS THE EFFECT OF CARBONDISULFIDE ON THE PATHWAY OF DOPAMINE SYNTHESIS AND INTERRUPTION OF FEED BACK MECHANISM BY CARBONDISULFIDE.





**FIG: 24**

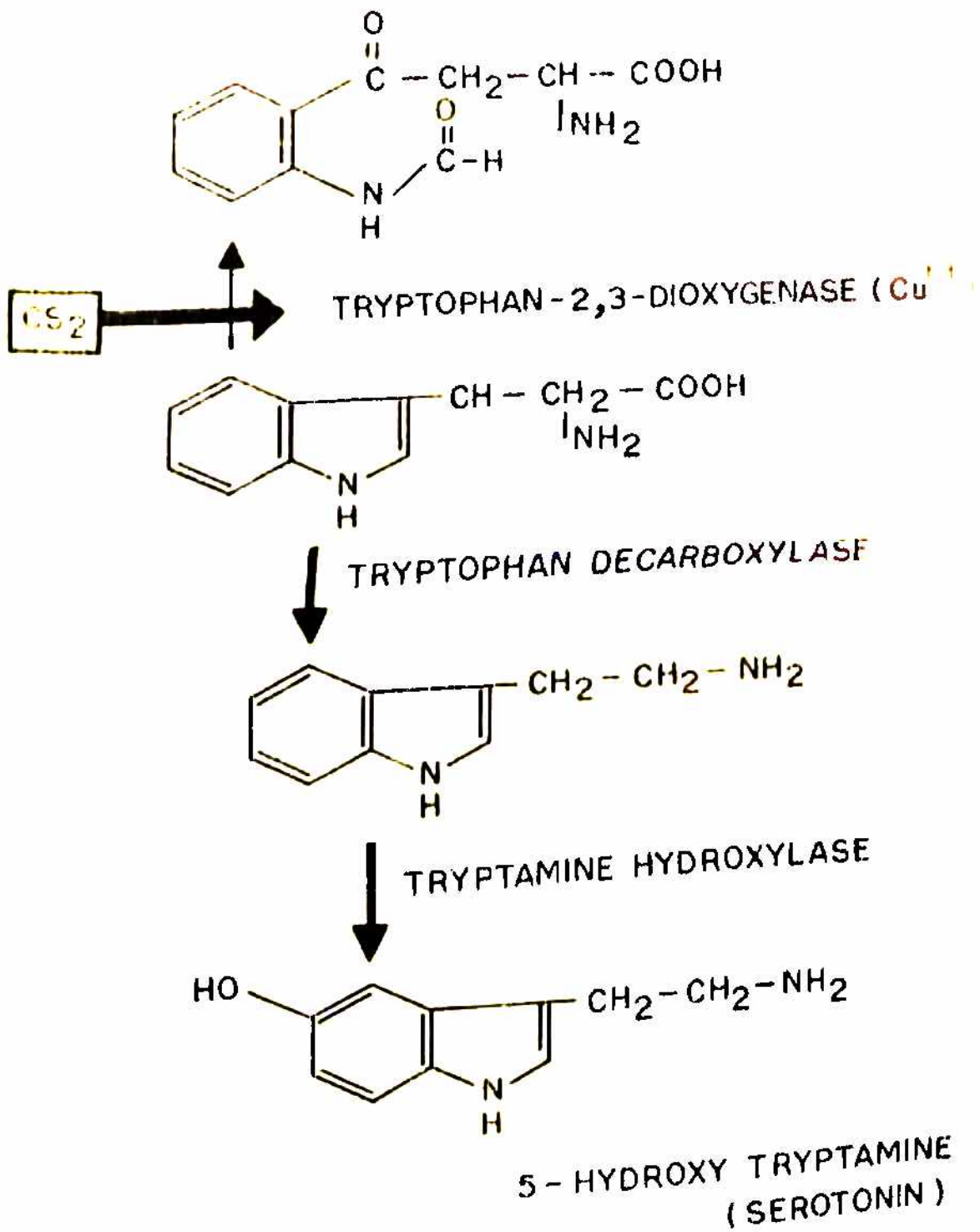
in DA levels in lungs after chronic CS<sub>2</sub> treatment may be responsible for the probable increase in the activity of dopamine beta-hydroxylase and decreased level of MAO activity ( P < .005 ) observed in this study.

#### 4.3.2. CARBONDISULFIDE TOXICITY AND 5 -HYDROXY-TRYPTAMINE :-

Aleksander (1966) and Meiner et.al. (1977) have reported an increased excretion of 5-HIAA in the urine of workers exposed to CS<sub>2</sub> and ozone respectively, and thought an increased degradation of 5-HT. Maj'Jerzy et.al. (1970) reported a rise in the level of brain 5-HT in toxicity studies with the disulfiram and diethyldithiocarbamate as observed in the present studies in brain, heart, liver and lungs of rats treated chronically with CS<sub>2</sub>. However, the weight of lungs have not been increased which reflects that CS<sub>2</sub> does not effect either the tissues water retention as observed with other toxicant under similar conditions ( Meiner et.al. 1977; Skillen et.al. 1961), or might not produce any injurious effect to kidneys to produce abnormal excretion of water. Therefore, the increase in the level of 5-HT can partially be explained on the basis of an increased tryptophan decarboxylase activity ( Skillen et.al. 1961, with ozone ). Besides, the rise in 5-HT level may also be in response to the inhibition of MAO and any MAO inhibitors is liable to cause an increase in the level of 5-HT in brain ( Brodie and Shore 1957). In

FIGURE : 25

SHOWS THE EFFECT OF CARBONDISULFIDE ON THE PATHWAY  
OF TRYPTOPHAN CATABOLISM AND 5-HYDROXY TRYPTAMINE  
SYNTHESIS.



**FIG: 25**



liver and heart, as there was not any significant inhibition of MAO, so the accumulation of 5-HT, may be wholly due to its increased synthesis. Normal tryptophan degradation pathway may, therefore, be assumed to be diverted for the increased synthesis of 5-HT because of presumed increase of tryptophan concentration. The increased synthesis of 5-HT leads to accumulation of 5-HT and the increased utilisation of this amino acid to the conversion of 5-HT. These findings lead one to think that tryptophan pyrrolase activity perhaps, is inhibited alongwith other enzymes of tryptophan degradation by CS<sub>2</sub> chronic treatment - of amines. The deficiency of pyridoxal phosphate caused by CS<sub>2</sub> toxicity may be another reason of 5-HT accumulation (Teisinger 1971). The decrease of tissue glutathione (G-SH) levels cannot also be overlooked since glutathione provides a defensive mechanisms to the body and various enzymes which react with CS<sub>2</sub> permanently.

Concludingly, we assume that simultaneous rise in DA and NE in most of the organs may be attributed to the increased activity of enzymes responsible for the catalysis of the conversion of Tyrosine to NE. From this it is suggested that feedback mechanisms have also been disrupted by chronic CS<sub>2</sub> toxicity which caused the rise in DA, NE and 5-HT in organs.

4.4.1.

ALTERATION OF TRACE ELEMENT METABOLISM BY  
CARBONDISULFIDE TOXICITY :-

4.4.1.1.

ZINC AND CARBONDISULFIDE TOXICITY :-

Earlier studies on zinc deficient rats have shown a reduced activity of various independent enzymes in testis bones, oesophagus and kidney ( Prasad, 1966 ; Oberleas 1971). In the present work also, chronic CS<sub>2</sub> treatment causes a fall in SDH activity in liver and brain confirming the earlier view that this enzyme is a zinc dependent as it decreases with observed reduction of zinc. Therefore, supporting the earlier suggestion that SDH activity reduction causes focally occurring elective parenchymal necrosis in liver (Minder and Rothe, 1970). Similar fall of SDH activity in tissues of zinc deficient rat was also observed by Burch et.al. (1973) which is in support of our finding that CS<sub>2</sub> treatment causes zinc deficiency on the basis of another zinc metallo enzyme, aldolase study (Olkhovaskaya 1976), It can be proposed that CS<sub>2</sub> exposure can cause reduction of SDH and probably all other zinc dependent enzymes. Further, in this study the reduction in these enzymes activity bear a direct relationship to the decreased level of zinc and glycolysis in tissues.

The reduction of zinc in pancreas by chronic CS<sub>2</sub> treatment could be an indication of pancreatic dysfunction



because a considerable amount of zinc binds with insulin in pancreatic storage granules ( Scott, 1934). On these basis, glucose tolerance of zinc deficient animals was reported to be depressed as compared to that of pair fed controls ( Quarterman et.al. 1966; Boquis and Lernmark, 1969 ; Hendrick and Mahoney, 1972; Huber and Gerschoff, 1973). Since zinc participates in the synthesis and storage of insulin in beta-cells of langerhans, therefore, the amount of insulin present in animals treated with CS<sub>2</sub> could be expected to be lowered, with reduced glucose tolerance which has been caused by an increased rate of insulin degradation ( Hendricks and Mahoney, (1972) .'

The observed rise in liver glycogen with an increased activity of phosphoglucomutase, shows that possibly, there is activation of the process of glycogen synthesis and reduction of glycolysis.' This may partially or wholly be due to zinc deficiency caused by CS<sub>2</sub> treatment in liver, brain and pancreas.' Hyper glycemia and decreased glucose catabolism in liver may, therefore, be due to the increased glycogen synthesis and its storage.'

The depletion of total zinc of body by CS<sub>2</sub> chronic treatment may also be one of the reasons in the fall of protein level ( Macpinlac et.al. 1968 ; Somers and Underwood, 1969 ; Sandstead et.al. 1971; Prasad et.al.' 1971 ).' Abnormality in protein metabolism pattern of plasma and tissues could indicate

that zinc deficiency can stimulate protein wastage by quick protein catabolism as suggested earlier (Fox and Hamson, 1965, Miller et.al. 1968 ; Tao and Hurlby, 1971 ; Macpinlac et.al. 1968; Somers and Underwood, 1969). It has been indicated further, that impaired protein biosynthesis could also contribute to the deficiency of proteins in animals ( Theuer and Hoekstra, 1968 ; Mills et.al. 1967 ). The decreased protein biosynthesis under zinc deficiency has been further related to lower RNA/DNA ratios in liver, kidneys and pancreas ( Prasad et.al. 1971) and higher non-protein nitrogen ( Somers and Underwood, 1969). The reduction of zinc in the body by CS<sub>2</sub> chronic treatment could be attributed to the reduction of its storage capacity of liver because of the degeneration produced by CS<sub>2</sub> ( Unpublished observation, Subhash and Chopra, 1978).

#### 4.4.2. MANGANESE AND CARBONDISULFIDE TOXICITY :-

Manganese homeostasis appears to be regulated at the excretory level rather than at the site of absorption ( Maynard and Cotzias, 1958 ; Briton and Cotzias , 1966 ; Hughes et.al. 1966 ; Papavasiliou et.al. 1966; Berthicamps et.al. 1966). The regulation of bile from liver has been reported to be affected by manganese level, therefore, manganese has been considered to be an important element affected in disease and health.

A steep rise in manganese with an increase of liver



FIGURE : 26

SHOWS THE INTERRELATIONSHIP OF ZINC AND MANGANESE IN  
LIVER AND PANCREAS UNDER THE STRESS OF CARBONDISULFIDE.

and brain glycogen evolves a clue of involvement of certain manganese dependent enzymes responsible for carbohydrate metabolism. Most of glycosyl transferases involved in the transfer of sugar moiety from sugar nucleotide require manganese ion for the activity ( Leach, 1971). The accumulation of manganese in liver, therefore, may have some influence on the increased glycogen biosynthesis which results due to an enhancement of the metalloenzyme, UDP- glycosyl transferase activity. This view supports the findings of Jarvisalo and Savolainen (1977) who also reported the increased activity of UDP- glucuronyl transferase of liver under CS<sub>2</sub> toxicity. An increase in tissue manganese and proposed increased in glycosyl transferase activity may cause increased phosphoglucomutase activity noticed in the present study. This could however, be explained only if the equilibrium is shifted in favour of the synthesis of glycogen in liver supporting further our earlier contention of increase in the synthesis of glycogen. Other alterations in carbohydrate metabolism related to manganese changes are (i) a rise of manganese in pancreas, (ii) a depletion of zinc in this tissue (iii) pancreatic necrosis and (iv) an increased serum alpha amylase activity, (Shrader and Everson 1968 ; Everson and Shrader 1968 ). The increased manganese level may therefore be considered as a protective action of the body against pancreatic damage showing thereby that manganese and zinc may have inverse relationship in liver and pancreas. The similarity between

the symptoms of chronic manganese poisoning and Parkinson's disease prompted that a study should be conducted to draw actual possible relationship between tissue manganese and biogenic amines. Mens et.al. (1970) have reported that the administration of L-dopa to patients with chronic manganese toxicity resulted in disappearance of rigidity and hypokinesia, improvement of postural reflexes and restitution of balance. Papavasiliou et.al. (1968) have proposed that cyclic AMP is the link between biogenic amines and manganese metabolism. They suggested that altered cyclic AMP increased manganese level and increased biogenic amines, as evidenced by increased liver retention of manganese and decreased biliary excretion, are interrelated. Mushtafa and Chandra (1975) reported reduction of DA and NE level while 5-HT level remained unaltered in animals fed with toxic dose of manganese. However, the present study with CS<sub>2</sub> toxicity depicted no true picture of biogenic amines in relation to manganese level because DA, NE and 5-HT levels were elevated while there was no significant change in brain manganese level. These observations with CS<sub>2</sub> could possibly be explained, on the basis that CS<sub>2</sub> alters blood brain barrier by producing drastic disturbances in physiological and biochemical picture of an animal and increasing the biogenic amines. This is supported further, by the observation of depletion of liver zinc which bear an inverse relation with manganese in all other tissues. A rise in liver manganese level alongwith simultaneous increase in biogenic



amines level of liver support our contention of brain biogenic amine and manganese level alterations. It may therefore, be suggested that CS<sub>2</sub> in rat brings about the changes in tissue manganese levels in various organs effecting carbohydrate metabolism and thereby brings about toxic influence on biogenic amines of brain and liver.

#### 4.4.3. COPPER AND CARBONDISULFIDE TOXICITY :-

Copper has been advocated as an essential element for the elaboration of heme, a component of cytochrome oxidase, thus establishing that copper is a biochemical catalyst, The increase of copper in liver and pancreas has suggested that copper metalloenzymes quantities are increased in these organs thus causing increased metabolism. Simultaneously, CS<sub>2</sub> intoxication forms dithiocarbamates by the interaction with biogenic amines in these tissues by utilising the available copper in the process which results in the increase of the dithiocarbamates synthesis and thus CS<sub>2</sub> toxicity ( Mckenna and Distefano, 1977). The interaction of CS<sub>2</sub> with copper and biogenic amines appears to be on the similar or same sites as observed with another inhibitor 2-mercaptoethyl guanidine an inhibitor of DBH (Diliberto and Distefano, 1973) because both the treatment caused increased DA level.

The fall of copper in brain, testis and kidneys



however, suggests that either there is no complexation of amines with  $CS_2$  in these tissues or if the complex is being formed is not much dependent on copper. There may be other elements available in these tissues which chelates dithiocarbamates in these tissues and cause the inhibition of enzymes. The observation that the increase of copper level in liver and pancreas leads to suggest that  $CS_2$  induced DBH inhibition can be due to its high lipid solubility, its ability to cross granular membrane and increased dithiocarbamate formation by binding the available biogenic amines in presence of enzymic copper ( Mckenna and Distefano, 1977 ). However, the increase in DA and NE level seen in liver with an increase in copper level appears to be contrary to this hypothesis. Therefore, it can be speculated that liver CA granular storage may be different from brain CA storage sites.

The elevation of biogenic amines ( DA, NE ) is observed in all organs under  $CS_2$  intoxication. The rise in DA level is in confirmity with the earlier findings ( Mckenna and Distefano , 1977 ; Magos, 1971 ; Maj and Vetulani, 1970 ; Diliberto and Distefano, 1973 ; Magos and Jarvis, 1976 ). They attributed this rise of DA due to inhibition of DBH. The rise in brain NE may, therefore, be attributed only to the plausible inhibition of MAO, a cuproenzyme ( Buffoni and Blaschko, 1964 ). The decrease of copper level in brain

further confirms the assumption that increased MAO activity of liver may be due to the increased copper level and the microsomal copper dependent enzyme system of liver. These enzymes may cause quick oxidation of biogenic amines and amino acids, viz. tyrosine and tryptophan. The increase of 5-HT in all organs may be related to inhibition of tryptophan pyrrolase, which is also a cuproenzyme and is essential for normal catabolism of tryptophan. The decreased protein synthesis may have diverted the tryptophan for the synthesis of 5-HT, which might be available for metabolism by increased MAO in the liver, thus confirming the existence of feed back mechanisms i.e. with the increased catabolising enzymes the biogenic amines level should be increased. Hence, the accumulation of 5-HT might be due to its increased synthesis and also decreased degradation in brain. (Maj and Maria, 1970).

Further, the reduction of copper in brain, kidney and testis also can be explained by considering the similar views of Massoud (1977), that useful copper in the living system becomes unavailable because of the firm interaction between copper and dithiocarbamates in liver. The reduction of copper in these tissues may lead to abnormal functioning of testis and kidneys, thereby giving an explanation for nephritis and infertility (Gondzik, 1976) in workers exposed to CS<sub>2</sub> environment. The decrease in usable copper with

decreased ceruplasmin has been found to alter the mobilisation of iron from reticuloendothelial cells ( Rolser et.al. 1970) This might lead to decreased heme synthesis, increased heme degradation and increased heme oxygenase activity in CS<sub>2</sub> treated rats ( Jarvisalo, et.al.' 1978) and hence may explain the causes of anemia seen in patients from industry using carbondisulfide.

In the existing literature there is a paucity of reports regarding the role of copper in carbohydrate metabolism. However, the observed changes in carbohydrate metabolism in the present study appear to be solely related to the zinc and manganese alterations but the role of copper level in various body organs and its relationship to carbohydrate metabolism cannot be ignored and needs further scientific probe in this direction.

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SUMMARY



The low health status and overall high susceptibility to various disorders of individuals exposed to  $CS_2$  have caused considerable doubts about the safety of this substance. The interaction between biological systems and  $CS_2$  has been a problem of only recent interest and few toxic manifestations in human beings and animals have been ascribed to it. The present work was undertaken to reveal the relationship, if any, between toxic manifestations of  $CS_2$  with observed abnormality of carbohydrate metabolism in relation to various enzymes and biogenic amines in rats.

Acute and chronic  $CS_2$  treatments caused a marked change only in liver glycogen rather than in the brain, which indicated that toxicity of  $CS_2$  might be a result of liver damage. Liver phosphorylase was found to be inhibited significantly while in the brain it was altered to an insignificant extent only. However, phosphoglucomutase activity of liver varied markedly, i.e. about eight times, than in brain which showed that the deposition of glycogen is much higher in liver than in the brain. The indirect approach of studying fructose, 1-6, diphosphatase also showed an inhibition trend and revealed decreased gluconeogenesis. The inhibition of adenosine triphosphatase and succinic dehydrogenase activity in liver and brain provided the proofs for the altered membrane structure, decreased activity of Kreb's cycle and electron transport chain. This, further, showed a reduction

in glycolysis and increase in glycogen deposition indicating that all processes depending upon the transport mechanisms are affected by  $CS_2$ . Therefore, increased susceptibility to infections and diseases could be attributed to the membrane structure alteration.

The depletion of plasma proteins and tissue proteins can be due to an increased utilisation of proteins by systems either for energy production or to overcome the stress induced by carbondisulfide.

The increase in brain, heart, liver and lungs biogenic amines i.e. dopamine, norepinephrine and 5-hydroxytryptamine (5-HT), provided further evidence of severe biochemical alterations by  $CS_2$  in chronically treated rats. The increased level of these biogenic amines support the assumption of the loss of granular permeability of the vesicles. However, the synthesis of these essential amines, could not be altered as the accumulation of tyrosine and tryptophan was found to be more than normal in these tissues. The simultaneous reduction in monoamine oxidase (MAO) activity in brain could explain further, the increase in the level of brain DA and NE. On the other hand, MAO activity in liver was increased which showed that the increase of biogenic amines in these organs may be due to an increase in their synthesis rather than inhibition



of catabolism. In liver, brain, heart and lungs there was a significant fall in the reduced glutathione which explains the protective action of glutathione against  $CS_2$  in these organs. This shows that biogenic amines-synthesising enzymes might not be inhibited substantially, because glutathione must be sparing the enzymes for covalent binding with  $CS_2$  and hence caused a rise in biogenic amines.

A study of trace elements metabolism showed that zinc was decreased in all these tissues. The pancreatic zinc reduction reflects pancreatic dysfunction or insulin reduction due to defective carbohydrate metabolism and in protein metabolism. The altered manganese concentration has been related to the increased glycogen biosynthesis and increased phosphoglucomutase activity in liver. Accumulation of manganese has been attributed to the increased level of DA and NE observed under  $CS_2$  intoxication. The manganese studies, therefore, provide increased evidence of anemia and other cardiovascular disorders in the workers handling  $CS_2$ .

Increased level of copper in liver and pancreas might be considered as an index of increased biogenic amines which formed complex with  $CS_2$  to give dithiocarbamates. On the other hand the decreased zinc level in testis, brain and kidneys may explain the mobilization of copper from these tissues to liver and pancreas so as to maintain Zn/Cu ratio

under the stress of  $CS_2$  thus may afford to justify the lessening of the stress.

Therefore, it can be concluded from the present study that  $CS_2$  toxicity causes a disturbance in carbohydrate metabolism of the body with alteration of paired trace elements "Zinc and Copper" and "Copper and Manganese". The paired elements have been found to bear an inverse relationship in liver and pancreas under the  $CS_2$  stress. The increased liver glycogen is related to increased phosphoglucomutase activity and decreased phosphorylase activity. Increased liver manganese is thought to be responsible for glycogen accumulation by increasing glycogen synthesis activity indirectly. Decreased Zinc level may inhibit the enzymes of glycolysis and Krebs' cycle. Though no relationship between trace elements and biogenic amines was established in this study, the rise of liver manganese and biogenic amines are interrelated confirming the earlier studies.

From this work it may, further, be suggested that the diet supplemented with trace elements may alleviate some toxic effects of carbondisulfide in workers.



CHAPTER -6

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